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**TITLE PAGE**  
**- Korean Journal for Food Science of Animal Resources -**  
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ARTICLE INFORMATION	Fill in information in each box below
<b>Article Title</b>	Effects of <i>Astragalus membranaceus</i> , <i>Adenophora triphylla</i> , and <i>Ulmus pumila</i> extracts on quality characteristics and storage stability of sous-vide cooked chicken breasts
<b>Running Title (within 10 words)</b>	Quality characteristics of sous-vide chicken breasts with herbal plant extracts
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<b>Conflicts of interest</b> List any present or potential conflict s of interest for all authors. (This field may be published.)	No potential conflict of interest relevant to this article was reported.
<b>Acknowledgements</b> State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available. (This field may be published.)	This research was supported by the Kyungpook National University Research Fund.
<b>Author's contributions</b> (This field may be published.)	Conceptualization: Park CH. Data curation: Lee B, Choi YM. Formal analysis: O H, Kim D. Methodology: Park CH, O H. Software: Kim JY, Cho DK. Investigation: Park CH, Lee B. Writing - original draft: Lee B, Park CH, Kim JY, O H, Kim D, Cho DK, Kim YS, Choi YM.
<b>Ethics approval (IRB/IACUC)</b> (This field may be published.)	The human ethics approval was granted by Bioethics Committee of KNU (protocol number: 2019-0027).

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10 **Effects of *Astragalus membranaceus*, *Adenophora triphylla*, and *Ulmus***  
11 ***pumila* extracts on quality characteristics and storage stability of sous-vide**  
12 **cooked chicken breasts**

14 **Abstract**

15 This study aimed to investigate the influence of *Astragalus membranaceus* (AM),  
16 *Adenophora triphylla* (AT), and *Ulmus pumila* (UP) extracts on the quality traits, palatability,  
17 and storage stability of sous-vide (SV) cooked chicken breasts. Chicken breasts were  
18 marinated in AM, AT, or UP extracts for 1 h, and then consistently cooked at a constant  
19 temperature of 60°C for 2 h. SV cooked chicken breasts with the UP extract exhibited lower  
20 lightness and higher yellowness values on the surface region compared to those with the AM  
21 and AT extracts ( $p < 0.05$ ). The control and UP groups displayed a similar overall visual  
22 acceptability ( $p > 0.05$ ), although the UP group had lower color acceptability ( $p < 0.01$ ). The UP  
23 group also had higher flavor and lower off-flavor intensities compared to the control group  
24 ( $p < 0.05$ ), although similar scores were observed in tenderness attributes and juiciness among  
25 the groups ( $p > 0.05$ ). Owing to these results regarding overall sensory acceptability, samples  
26 from the UP group were more preferred by the trained panelists compared to samples from the  
27 control group ( $p < 0.001$ ). On d 14 of cold storage, all the groups with herbal medicinal  
28 extracts exhibited a lower concentration of thiobarbituric acid-reactive substances than the  
29 control group ( $p < 0.05$ ), and the AT and UP groups showed lower values compared to the AM  
30 group due to their higher flavonoid contents ( $p < 0.001$ ). Therefore, meat marination with  
31 herbal plant extracts before SV cooking can be effective for enhancing the overall quality of  
32 SV cooked chicken breast.

33 **Key words:** Sous-vide cooking, Herbal medicinal extracts, Quality characteristics, Storage  
34 stability, Chicken breasts.

## 35 **Introduction**

36        Sous-vide (SV) cooking generally uses a vacuum packaging machine and precisely  
37 thermo-controlled water bath to provide efficient and uniform heat penetration into food  
38 products (Park et al., 2020). It is considered one of the suitable methods for home-meal  
39 replacements, since SV cooking has several benefits, including extending shelf life, enhancing  
40 product yield, and preventing the nutrient loss (Baldwin, 2012). However, due to the lower  
41 cooking temperature, meat cooked with SV method exhibited a less developed brown color  
42 and flavor compared to meat cooked with conventional method (Park et al., 2020). These  
43 results were associated with lack of Maillard reaction (MR) development, since the MR  
44 products related to desired flavor increase with increasing cooking temperatures (Cho et al.,  
45 2021). Therefore, additional treatments, such as searing and marination, are necessary to  
46 compensate for the drawbacks of SV cooking.

47        Recently, since consumers strongly believe that foods can directly contribute to their  
48 health, they are becoming more interested in healthier food products that use natural additives  
49 (Choi et al., 2012). Thus, the food industry is employing plant-based products, which contain  
50 various antioxidant compounds, such as carotenoid and flavonoid, as natural ingredients (Kim  
51 et al., 2009). The medicinal plant extracts with a higher amount of antioxidant not only can  
52 improve the functionality of meat products, but also inhibit the deterioration of food quality  
53 by preventing lipid oxidation during processing and storage (Pompella et al., 2014).  
54 Additionally, herbal medicinal extracts can be used as a flavoring agent for the development  
55 of meat products due to their specific flavors (Aminzare et al., 2019).

56        *Astragalus membranaceus* (AM; called as Hwanggi), *Adenophora triphylla* (AT; called  
57 as Jandae), and *Ulmus pumila* (UP; called as Ugeunpi), which are widely distributed  
58 throughout the world, and are used as herbal medicines in Asian countries for liver cirrhosis,  
59 chronic bronchitis, and inflammation, respectively (Kim et al., 2009; Sun et al., 2007; Zhou et

60 al., 2017). Also, these herbal medicines commonly contain greater polyphenolic compounds  
61 as like blueberry and rosemary, which have antioxidant properties (Kim et al., 2009; Li et al.,  
62 2014; Zhou et al., 2017). Thus, AM, AT, and UP extracts can be used an ingredient for  
63 improving the quality and shelf life of meat products. Therefore, to improve usability by  
64 enhancing the quality of SV chicken breast, this study investigated the effects of AM, AT, and  
65 UP extracts on the quality traits, palatability, and storage stability of SV cooked chicken meat.

66

## 67 **Materials and Methods**

### 68 **Sample preparation and treatments**

69 The roots of AM, AT, and UP were purchased from a local medicinal plant market  
70 (Geumsan, Chungcheongnam-do, Korea). At 400 g in 10 L of water, each herbal medicine  
71 was used and boiled at 100°C for 2 h to obtain the extract. Extracts from the three herbs were  
72 poured into plastic containers and stored at 4°C until the marinades were prepared.

73 A total of 123 fresh boneless and skinless fresh chicken breasts were purchased from a  
74 local retail market. All the chicken breasts belong to the normal quality condition according to  
75 the chicken quality classification (Park et al., 2020). The samples were randomly assigned  
76 into 1 of 4 groups, the control and 3 herbal medicinal extract (AM, AT, and UP) groups. The  
77 control group was immersed in water at the meat-to-fresh water ratio of 1: 2 without any  
78 addition of plant extracts, and the three experiment groups were marinated in AM, AT, or UP  
79 extracts at a ratio of 1:2 (meat:extract) for 1 h.

80 All the samples were weighed, put into a polyethylene pouch, and vacuumed using a  
81 vacuum packaging machine (Leepack, Hanguk Electronic, Incheon, Korea). Samples were  
82 then cooked in a circulating thermostatic water bath at 60°C for 2 h, the optimal condition for  
83 chicken breast cooked SV (Park et al., 2020). All the SV samples were cooled in an ice-slurry  
84 until equilibration; then, the quality traits were immediately examined using 24 samples. A

85 total of 24 samples (6 samples per treatment) were stored at  $-20^{\circ}\text{C}$  for the assessments of  
86 visual attributes and sensory quality traits. The contents of polyphenols and flavonoids were  
87 analyzed using 27 chicken breasts (9 samples per treatment without control group). The  
88 remaining 48 samples were stored at  $4^{\circ}\text{C}$  to measure storage stability during a cold storage  
89 (period from d 0 to 14).

90

### 91 **Quality measurements**

92 The pH of SV cooked samples was determined using a Testo 206-pH2 (Testo AG,  
93 Lenzkirch, Germany) with a penetration probe. Color parameters, including lightness ( $L^*$ ),  
94 redness ( $a^*$ ), and yellowness ( $b^*$ ), were measured using a colorimeter (CR-400, Minolta  
95 Camera Co., Osaka, Japan) at the surface and inner regions of SV samples according to the  
96 recommendations of the Commission Internationale de l'Eclairage (1978). Hue angle [ $\tan^{-1}(b^*/a^*)$ ]  
97 and saturation index [ $((b^{*2}+a^{*2})^{0.5})$ ] at the same regions were calculated. Cooking loss  
98 and Warner-Bratzler shear force (WBS) of each sample were measured based on a previous  
99 publication (Honikel, 1998). Samples were weighed before and after SV cooking to calculate  
100 the percentage of cooking loss. After measuring cooking loss, more than six core samples  
101 (1.27 cm diameter) were obtained parallel to the muscle fiber orientation for WBS  
102 measurement. The WBS value was collected using an Intron Universal Testing Machine  
103 (Model 1011, Instron Corp., Canton, MA, USA) with the Warner-Bratzler blade (crosshead  
104 speed, 200 mm/min).

105

### 106 **Visual attributes and eating quality characteristics**

107 For analyses of visual attributes and eating quality characteristics, a total of 24 samples  
108 were randomly coded with a 3-digit number and used during four sessions (six samples per  
109 session). Before each session, the frozen SV samples were thawed at  $4^{\circ}\text{C}$  overnight, then

110 heated to and maintained at a core temperature of 54°C in a water bath until further analyses.  
111 All the panelists (six women and five men aged 23 to 48) were trained according to the  
112 previous procedures (American Meat Science Association, 1995; Meilgaard et al., 1991), and  
113 evaluated the visual attributes and sensory quality characteristics of SV breasts to use a  
114 hedonic scale (1 to 9). Training of the panelists and sensory evaluations were conducted at the  
115 Kyungpook National University (KNU). Visual attributes, including color, moisture,  
116 appearance, and overall acceptability, were evaluated. A total of 11 eating quality attributes,  
117 including initial tenderness, rate of breakdown, amount of perceptible residue, juiciness,  
118 flavor intensity, off-flavor intensity, treatment flavor acceptability, sweetness, sourness,  
119 bitterness, and overall acceptability were assessed.

120

#### 121 **Total polyphenol and flavonoid contents**

122 A total of 27 SV cooked samples (3 treatments × 3 samples × 3 repetitions) were used to  
123 measure the polyphenol and flavonoid contents. One gram of SV sample was homogenized,  
124 and extracted using 10 mL of 70% ethanol (v/v) and methanol (v/v) solutions for the  
125 assessment of polyphenol and flavonoid contents, respectively. For the determination of total  
126 polyphenol content by the Folin-Ciocalteu procedure, the method described by Singleton and  
127 Rossi (1965). The flavonoid content for each sample was evaluated according to the method  
128 described by Song et al. (2014), with some modifications. The results of both polyphenol and  
129 flavonoid contents were expressed as mg/100 g of experiment sample.

130

#### 131 **Storage stability**

132 The levels of lipid oxidation in the SV cooked breasts during storage at 4°C was assessed  
133 by measuring the thiobarbituric acid-reactive substances (TBARS) according to the method  
134 described by Buege and Aust (1978) and Cho et al. (2020). A total of 48 samples (16 samples

135 per each repetition) were used on d 0, 3, 7, and 14 as three repetitions. The TBARS values  
136 were expressed as milligrams of malonaldehyde (MDA) per kg of SV sample.

137

### 138 **Statistical analysis**

139 The general linear model in SAS software (SAS Institute, Cary, NC, USA) was  
140 performed to compare the quality traits, visual attributes, palatability characteristics, and  
141 storage stability, including levels of polyphenol, flavonoid, and TBARS, among the SV  
142 cooked chicken breasts with different herbal medicinal extracts. A linear mixed model was  
143 used to identify the factors influencing the quality traits, visual attributes, palatability, and  
144 polyphenol and flavonoid contents. In the model, the fixed effect commonly included the  
145 herbal extracts, and the random effects included the number of experimental repetitions and  
146 panelists. A linear mixed model was also used to compare the TBARS values of the SV  
147 cooked meats among the groups, with the extracts and storage periods as the fixed effects and  
148 repetitions as the random effects. Significant differences among the groups were determined  
149 by the probability difference at 5%. All the data were presented as the least-squares means  
150 with standard errors.

151

## 152 **Results**

### 153 **Effect of herbal medicinal extracts on quality and palatability traits**

154 Meat quality characteristics among the SV cooked chicken breasts marinated with  
155 different herbal medicinal extracts were compared (Table 1). No difference was observed in  
156 pH between the control and herbal extract groups ( $p>0.05$ ). While all the groups displayed a  
157 similar redness value on the surface region ( $p>0.05$ ), the SV samples with UP extract showed  
158 the lowest lightness and highest yellowness values compared to the SV samples with other  
159 extracts and samples without extracts ( $p<0.05$ ). A similar hue angle was observed among the



160 groups ( $p>0.05$ ), and the UP group exhibited a higher saturation index compared to the other  
161 groups ( $p<0.05$ ). In the inner region of SV cooked breasts, there were no differences in any of  
162 the color parameters among the groups ( $p>0.05$ ). On the other hand, the control group  
163 exhibited a lower cooking loss compared to the AT group (17.4 vs. 18.5%,  $p<0.01$ ), and  
164 showed a similar loss compared to the AM and UP groups ( $p<0.05$ ). No difference was  
165 detected in WBS value among the groups ( $p>0.05$ ).

166 Comparison of the visual attributes and palatability characteristics among the SV cooked  
167 meats with different extracts are shown in Table 2. The AM group exhibited lower color  
168 acceptability compared to the other groups ( $p<0.01$ ), except for the UP group ( $p>0.05$ ). There  
169 was no difference in the moisture intensity and appearance acceptability among the groups  
170 ( $p>0.05$ ). The control group showed similar overall acceptability compared to the other  
171 groups ( $p>0.05$ ), except for the AM group ( $p<0.01$ ).

172 A similar score of initial tenderness was observed in the SV cooked meat with AM, AT,  
173 and UP extracts ( $p>0.05$ ), and the AM group had a lower value compared to the control group  
174 (7.63 vs. 8.22,  $p<0.05$ ). No differences were detected in rate of breakdown, amount of  
175 perceptible residue, and juiciness between the control and herbal extract groups ( $p>0.05$ ). SV  
176 breast added UP extract showed a higher flavor intensity compared to SV breast added AM  
177 extract (6.87 vs. 6.37,  $p<0.01$ ), and the other herbal treatments had similar scores compared to  
178 the control group ( $p>0.05$ ). In contrast, a marked difference was observed in off-flavor  
179 intensity among the groups. The herbal plant extract groups scored higher than the control  
180 group ( $p<0.001$ ). Similar to the pattern in flavor intensity, the level of treatment flavor  
181 acceptability did not differ among the control and herbal extract groups ( $p>0.05$ ), except for  
182 the UP group ( $p<0.05$ ). While all the groups had similar values of sweetness and sourness  
183 ( $p>0.05$ ), the control group scored higher on bitterness than the herbal extract groups  
184 ( $p<0.001$ ), except for the AT group ( $p>0.05$ ). The herbal plant treatments, except for the AM

185 group, showed a higher score of overall acceptability compared to the control group ( $p < 0.01$ ).

186

### 187 **Effect herbal medicinal extracts on polyphenols, flavonoids, and TBARS contents**

188 Contents of total polyphenols and flavonoids among the AM, AT, or UP groups were  
189 compared (Fig. 1A). Although there was no significant difference in the total polyphenol  
190 content among the groups ( $p > 0.05$ ), a considerable difference was observed in the flavonoids  
191 content among the groups ( $p < 0.001$ ). Samples from the UP group had the highest flavonoid  
192 content among the groups, and a lower content was observed in the AM group compared to  
193 the AT group (1.13 vs. 1.54 mg/100 g,  $p < 0.001$ ).

194 Changes in the TBARS values among the groups during the storage period are shown in  
195 Fig. 1B. The TBARS values of all the groups tended to increase during 0 to 14 d of the cold  
196 storage ( $p < 0.05$ ). After d 7 of storage, all the groups showed a higher value than that on d 0  
197 ( $p < 0.05$ ). A difference in the increase of TBARS values was observed in the control (0.73 vs.  
198 0.88 mg MDA/kg) or AM (0.69 vs. 0.79 mg MDA/kg) groups between 7 and 14 d of cold  
199 storage ( $p < 0.05$ ). However, no differences were observed in the AT (0.69 vs. 0.70 mg  
200 MDA/kg) or UP (0.73 vs. 0.75 mg MDA/kg) groups during 7 to 14 d of storage ( $p > 0.05$ ).

201

## 202 **Discussion**

203 The herbal plants are widely available and have been considered as a potential source for  
204 enhancing food functionality (Krishnan et al., 2014). The extracts from herbal plants have  
205 various colors and flavors, which can influence the appearance characteristics of meat  
206 products (Jin et al., 2015). For instance, turkey breasts with rosemary extract showed a lower  
207 lightness value compared to untreated breasts due to yellowish color of rosemary extract (Yu  
208 et al., 2002). However, the raw meat characteristics and cooking methods also influence the  
209 color of final meat products. For examples, no differences were observed in the lightness

210 value and appearance acceptability between the untreated beef meatballs and those with  
211 rosemary extract due to a darker color of raw beef (Fernandez-Lopez et al., 2005).  
212 Additionally, *Akebia quinata* extract did not influence the lightness value and sensory color  
213 score of emulsion-type pork sausage (Jin et al., 2015). In this study, the SV chicken breasts  
214 with UP extract exhibited darker and yellower color on the surface region compared to the  
215 other SV chicken breasts ( $p < 0.05$ ) due to a brown color of UP extract (Kim et al., 2016) and  
216 the lighter yellowish color of the other extracts (Li et al., 2015; Ny et al., 2021). However, the  
217 addition of UP extract influenced the color acceptability, and the control group was preferred  
218 to the UP group ( $p < 0.001$ ). In comparison, there were no differences in moisture, appearance,  
219 and overall acceptability between the control and UP groups ( $p > 0.05$ ). Additionally, all the  
220 groups did not differ in the cooking loss and WBS value ( $p > 0.05$ ) except for the AT group in  
221 cooking loss. Thus, herbal medicinal extracts in this study did not negatively affect the meat  
222 quality and visual attributes of SV cooked chicken breasts.

223 Poultry meat is more susceptible to quality deterioration mainly due to lipid oxidation  
224 with resulting off-flavors compared to red meat during storage and processing, (Jayasena et  
225 al., 2013). Marination using various herbal plant extracts that act as flavoring agents to  
226 compensate for the disadvantage by masking the off-flavor can be applied to diverse meat  
227 types, especially chicken meat (Embuscado, 2015). In a previous study, the pork patties with  
228 cassia bark extract had a lower rancid flavor compared to the patties without extract, although  
229 the overall acceptability was similar between the groups (Kong et al., 2010). Addition of  
230 0.02% rosemary extract to ground beef reduced the extent of warmed-over flavor compared to  
231 ground beef with distilled water (Ahn et al., 2002). However, herbal plant extracts generally  
232 have is a limited effect on the other sensory traits, such as tenderness and juiciness, of  
233 processed meat products (Hayes et al., 2011; Jin et al., 2015). A similar result was found in  
234 this study; the AM, AT, and UP groups did not differ in tenderness attributes and juiciness

235 compared to the control group ( $p>0.05$ ). UP extract demonstrated specific flavor, sweetness,  
236 and bitter taste as assessed by trained panelists (Lee et al., 2016). Consistent with the previous  
237 findings, this study found the SV breasts with UP extracts had better flavor and lower off-  
238 flavor intensities compared to SV breasts without herbal plant extract ( $p<0.05$ ). As these  
239 results, the trained panelists preferred the SV cooked meat with UP extract compared to the  
240 untreated meat ( $p<0.01$ ), although samples from the UP group tasted more bitter than samples  
241 from the other groups ( $p<0.001$ ) except for the AM group.

242 Lipid oxidation is well associated with protein oxidation, as the oxidation products of  
243 one substance can accelerate the oxidation of another substance (Cai et al., 2021). Thus, to  
244 assess storage stability, extents of these two or each oxidation are mainly measured. There is a  
245 need to suppress or delay the onset of lipid and protein oxidation in chicken products to  
246 increase shelf life. On the other hand, phytochemicals, especially flavonoids and phenolic  
247 acids derived from the herbal plant origins, are essential antioxidants due to their ability to  
248 scavenge free radicals (Embuscado, 2015). AM, AT, and UP as medicinal plants have also  
249 higher amounts of phenolic compounds (Kim et al., 2009; Li et al., 2014; Zhou et al., 2017);  
250 thus, these plant extracts can be used as food additives to improve the storage stability of meat  
251 products. In this study, the flavonoid contents of SV cooked chicken breasts were highest in  
252 the UP group, followed by the AT and AM groups ( $p<0.001$ ), although the total polyphenol  
253 contents were similar among the groups ( $p>0.05$ ). This finding may explain the previous  
254 observation that UP extract had better antioxidant (Im et al., 2017) and immunomodulatory  
255 properties (Chang and Woo, 2003) compared to AM and AT extracts, respectively. Due to its  
256 high flavonoid content, the UP group also exhibited lower TBARS concentrations compared  
257 to those the AM group after 14 d of cold storage ( $p<0.05$ ). Moreover, adding herbal plant  
258 extracts to the SV cooked breasts significantly inhibited the formation of TBARS compared  
259 to the control breasts at d 14 of storage ( $p<0.05$ ).

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## Conclusion

Taken together, the addition of AM, AT, and UP extracts before SV cooking enhanced the storage stability of chicken breasts during refrigeration without impairing the meat quality traits. In particular, the UP extract improved palatability of the chicken breasts by reducing the off-flavor and increasing the flavor intensities compared to the chicken breasts without plant extract. Therefore, herbal plant extracts, especially the UP extract, can be a good food additive for enhancing the overall quality of SV cooked chicken breasts and improving the utilization of plant extracts.

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359



360 **Figure caption**

361 **Fig. 1. Comparing the total polyphenol (TP), flavonoid (A), and 2-thiobarbituric acid**  
362 **reactive substance (TBARS; B) values among the sous-vide (SV) cooked chicken breasts**  
363 **with different herbal plant extracts. TBARS were measured during 14 d of storage at 4°C.**  
364 Control, SV cooked chicken breast with distilled water; AM, SV cooked chicken with the  
365 *Astragalus membranaceus* extract; AT, SV cooked chicken with the *Adenophora triphylla*  
366 extract; UP, SV cooked chicken with the *Ulmus pumila* extract. Bars indicate standard errors  
367 of least-square means, and different letters represents significant difference ( $p < 0.05$ ).

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**Table 1. Comparing meat quality characteristics among the sous-vide (SV) cooked chicken breasts with different herbal plant extracts**

	Control	Treatments <sup>1</sup>			SEM	Level of Significance
		AM	AT	UP		
Meat pH	6.19	6.22	6.16	6.17	0.03	NS
<i>Color – surface region</i>						
Lightness (L*)	81.7 <sup>a</sup>	81.7 <sup>a</sup>	82.0 <sup>a</sup>	78.6 <sup>b</sup>	0.62	**
Redness (a*)	3.12	3.11	2.86	3.05	0.37	NS
Yellowness (b*)	14.6 <sup>b</sup>	14.0 <sup>b</sup>	14.6 <sup>b</sup>	16.1 <sup>a</sup>	0.49	*
Hue angle <sup>2</sup>	77.3	77.5	79.0	79.1	1.31	NS
Saturation index <sup>3</sup>	14.4 <sup>b</sup>	13.7 <sup>b</sup>	14.9 <sup>b</sup>	16.4 <sup>a</sup>	0.50	*
<i>Color – inner region</i>						
Lightness (L*)	83.2	84.3	84.3	84.6	0.45	NS
Redness (a*)	3.91	3.93	3.63	3.43	0.33	NS
Yellowness (b*)	12.5	11.2	11.9	11.7	0.61	NS
Hue angle <sup>2</sup>	72.5	70.3	73.0	73.4	1.76	NS
Saturation index <sup>3</sup>	13.1	12.0	12.5	12.2	0.58	NS
Cooking loss (%)	17.4 <sup>b</sup>	17.1 <sup>b</sup>	18.5 <sup>a</sup>	16.5 <sup>b</sup>	0.36	**
Warner-Bratzler shear force (N)	18.2	19.9	17.7	19.8	1.17	NS

369 Level of significance: NS, not significant; \* p<0.05; \*\* p<0.01.

370 <sup>a-b</sup> Different superscripts in the same row represent significant differences (p<0.05).

371 <sup>1</sup> Control, SV cooked chicken breast with distilled water; AM, SV cooked chicken with the *Astragalus membranaceus* extract; AT, SV cooked  
372 chicken with the *Adenophora triphylla* extract; UP, SV cooked chicken with the *Ulmus pumila* extract.

373 <sup>2</sup> Hue angle =  $\tan^{-1}(b^*/a^*)$ ; <sup>3</sup> Saturation index =  $(b^{*2}+a^{*2})^{0.5}$ .

374

**Table 2. Comparing organoleptic characteristics among the sous-vide (SV) cooked chicken breasts with different herbal plant extracts**

	Control	Treatments <sup>1</sup>			SEM	Level of Significance
		AM	AT	UP		
<i>Visual attributes</i>						
Color <sup>2</sup>	6.08 <sup>a</sup>	5.17 <sup>c</sup>	5.67 <sup>ab</sup>	5.33 <sup>bc</sup>	0.17	**
Moisture <sup>3</sup>	6.33	6.17	6.25	6.33	0.16	NS
Appearance <sup>2</sup>	6.25	5.67	6.00	6.00	0.21	NS
Overall acceptability <sup>2</sup>	6.17 <sup>a</sup>	5.17 <sup>b</sup>	5.67 <sup>ab</sup>	5.67 <sup>ab</sup>	0.18	**
<i>Palatability characteristics</i>						
Initial tenderness <sup>4</sup>	8.22 <sup>a</sup>	7.63 <sup>b</sup>	7.79 <sup>ab</sup>	7.90 <sup>ab</sup>	0.15	*
Rate of breakdown <sup>5</sup>	7.81	7.28	7.45	7.58	0.16	NS
Amount of perceptible residue <sup>6</sup>	7.28	6.81	6.86	7.06	0.17	NS
Juiciness <sup>7</sup>	6.45	6.45	6.31	6.67	0.17	NS
Flavor intensity <sup>8</sup>	6.06 <sup>b</sup>	6.37 <sup>b</sup>	6.53 <sup>ab</sup>	6.87 <sup>a</sup>	0.15	**
Off-flavor intensity <sup>9</sup>	5.76 <sup>c</sup>	6.45 <sup>b</sup>	6.63 <sup>b</sup>	7.42 <sup>a</sup>	0.21	***
Treatment flavor acceptability <sup>2</sup>	6.25 <sup>b</sup>	6.45 <sup>ab</sup>	6.70 <sup>ab</sup>	6.87 <sup>a</sup>	0.16	*
Sweetness <sup>9</sup>	6.45	6.42	6.03	6.48	0.24	NS
Sourness <sup>9</sup>	7.50	7.03	7.38	7.18	0.20	NS
Bitterness <sup>9</sup>	7.55 <sup>a</sup>	6.81 <sup>bc</sup>	7.10 <sup>ab</sup>	6.41 <sup>c</sup>	0.19	***
Overall acceptability <sup>2</sup>	6.36 <sup>c</sup>	6.45 <sup>bc</sup>	6.83 <sup>ab</sup>	7.15 <sup>a</sup>	0.15	**

376 Level of significance: NS, not significant; \* p<0.05; \*\* p<0.01; \*\*\* p<0.001.

377 <sup>a-c</sup> Different superscripts in the same row represent significant differences (p<0.05).

378 <sup>1</sup> Control, SV cooked chicken breast with distilled water; AM, SV cooked chicken with the *Astragalus membranaceus* extract; AT, SV cooked  
379 chicken with the *Adenophora triphylla* extract; UP, SV cooked chicken with the *Ulmus pumila* extract.

380 <sup>2</sup> Score (1-9) = very unacceptable-very acceptable; <sup>3</sup> Score (1-9) = very dry-very moist; <sup>4</sup> Score (1-9) = very firm-very tender; <sup>5</sup> Score (1-9) =  
381 very slow-very fast; <sup>6</sup> Score (1-9) = very abundant-none; <sup>7</sup> Score (1-9) = not juicy-very juicy; <sup>8</sup> Score (1-9) = very weak-very strong; <sup>9</sup> Score (1-9)  
382 = very strong-very weak.

