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9 **Effect of modified casein to whey protein ratio on dispersion stability, protein quality**
10 **and body composition in rats**

11
12 **Abstract**

13
14 The present study was designed to investigate the effects of protein formula with different
15 casein (C) to whey protein (W) ratios on dispersion stability, protein quality and body
16 composition in rats. Modification of the CW ratio affected the extent of protein aggregation,
17 and heated CW-2:8 showed a significantly increased larger particle ($> 100 \mu\text{m}$) size distribution.
18 The largest protein aggregates were formed by whey protein self-aggregation. There were no
19 significant differences in protein aggregation when the CW ratios changed from 10:0 to 5:5.
20 Based on the protein quality assessment (CW-10:0, CW-8:2, CW-5:5, and CW-2:8) for four
21 weeks, CW-10:0 showed a significantly higher feed intake ($p < 0.05$), but the high proportion
22 of whey protein in the diet (CW-5:5 and CW-2:8) increased the feed efficiency ratio, protein
23 efficiency ratio, and net protein ratio compared to other groups. Similarly, CW-2:8 showed
24 greater true digestibility compared to other groups. No significant differences in fat mass and
25 lean mass analyzed by dual-energy x-ray absorptiometry were observed. A significant
26 difference was found in the bone mineral density between the CW-10:0 and CW-2:8 groups (p
27 < 0.05), but no difference was observed among the other groups. Based on the results, CW-5:5
28 improved protein quality without causing protein instability problems in the dispersion.

29
30 **Keywords:** protein quality, milk protein, casein-to-whey protein ratio, particle size, bone
31 mineral density

32 Introduction

33 Milk is one of the major food resources containing various essential nutrients (Haug
34 et al., 2007). In particular, milk protein, mainly consisting of casein (80%) and whey protein
35 (20%), usually accounts for approximately 3% of whole milk (Pereira, 2014). Milk protein has
36 showed higher digestibility than plant source protein (Gilani and Sepehr, 2003; Mathai et al.,
37 2017). Moreover, both caseins and whey proteins are important sources of branched-chain
38 amino acids and other bioactive peptides (Bos et al., 2000; Scholz-Ahrens and Schrezenmeir,
39 2000).

40 The amino acid sequence of milk proteins primarily influences the digestibility and
41 physicochemical characteristics of milk proteins and also leads to different digestion kinetics
42 (Gan et al., 2018). Caseins are easily coagulated by pepsin under a gastric condition so that it
43 is slowly digested whereas whey proteins rapidly pass through the stomach, are digested to
44 amino acids and peptides in the intestine, and increase the amino acid level in blood (Boirie et
45 al., 1997; Hall et al., 2003; Mahe et al., 1996; Ye et al., 2016).

46 Because of the difference between casein and whey protein characteristics, the casein
47 to whey protein ratio in milk formulation has affected *in vitro* digestion and physiological
48 activities in many aspects. A casein to whey protein ratio of 40:60 exhibited higher *in vitro*
49 digestion compared to 60:40 and 80:20 in infant formula (Phosanam et al., 2021). Similarly, as
50 the casein portion increased in milk protein from 20% to 100%, solid curd was easily formed
51 in simulated gastric conditions (Mulet-Cabero et al., 2020). Recently, Wood et al. (2021)
52 reported that modification of goat milk-based protein formulation from 80:20 to 40:60
53 (casein:whey) influenced food intake and hypothalamic neuronal activation in mice. In addition,
54 modification of the casein:whey protein ratio to 40:60 reduced the allergenic potential

55 compared to natural cow's milk (Lara-Villoslada et al., 2005). Taken together, it could be
56 suggested that modification of milk protein type may have different nutritional outcomes.

57 Although compelling evidence regarding the protein quality of each milk protein and
58 the effects of the casein to whey protein ratio on *in vitro* digestion and physiological activities
59 has existed, the effects of various blending ratios of casein to whey protein on physicochemical
60 properties and *in vivo* protein quality have not yet been fully elucidated. Based on the above
61 mentioned studies, we hypothesize that modified casein to whey protein ratios may play an
62 important role in protein quality including utilization and digestibility in rats.

63

64 Materials and Methods

65 **Materials**

66 Micellar casein isolate (MCI; Refit Micellar Casein isolate 88; Protein: 85%) and whey
67 protein isolate (WPI; Hilmar™ 902; Protein: 89.5%) were obtained from Friesland Campina
68 ingredients (Wageningen, The Netherlands) and Hilmar ingredients (Hilmar, CA, USA),
69 respectively.

70

71 **Preparation of protein dispersion with different casein:whey protein ratios**

72 Milk protein dispersions (5% protein, w/w) with different casein:whey protein ratios
73 (CW-10:0, CW-8:2, CW-5:5, and CW-2:8) were prepared by reconstitution of appropriate
74 amounts of MCI and WPI. The protein dispersions (2 L) went through a two-stage homogenizer
75 (Ariete NS 2006, GEA, Italia) at 110 bar and 50 bar, respectively. The aliquots of samples (1
76 L) were heated in a 95°C water bath (Chang Shin Science, Korea) for 30 min to simulate

77 pasteurization.

78

79 **Protein solubility**

80 The samples were placed on a multi-stirrer (MS-MP8, Wisd Laboratory Instrument,
81 Germany) for 1 h at 350 rpm and were subjected to centrifuge (Beckman Coulter, Fullerton,
82 CA, USA) at $6,000 \times g$ for 20 min. The protein solubility of the samples (unheated and heated
83 samples) were calculated by quantifying proteins before and after centrifugation. The protein
84 content of the samples was determined using the bicinchoninic acid assay (Cortes-Rios et al.,
85 2009). Briefly, sample (25 μL) was mixed with BCA solution (200 μL) in a 96-well plate and
86 placed in a plate reader (Biotek Instruments Inc., Winooski, VT, USA) for 30 min at 37°C . The
87 absorbance was taken at 562 nm and protein content was calculated from the standard curve
88 prepared using bovine serum albumin.

89

90 **Particle size distribution**

91 The changes in the particle size distribution of milk protein dispersions before and
92 after pasteurization were measured using a particle size analyzer (Horiba LA-960 Laser
93 Scattering Particle Size Analyzer, Japan) as previously described (Yun and Imm, 2021).

94

95 **Protein profile analysis**

96 The protein profile of CW-2:8 dispersion was analyzed since only CW-2:8 dispersion
97 showed significant changes in particle size distribution upon heat treatment. The freeze-dried

98 samples (CW-2:8 and heated CW-2:8; 20 mg/mL) were loaded onto a column (15 mm × 450
99 mm) packed with Sephacryl S-500HR (GE Healthcare Bioscience, Sweden). The sample was
100 eluted with Bis-Tris-Propane buffer (20 mM, pH 7.0) at a flow rate of 1 mL/min. The eluted
101 peak fraction detected at 215 nm was collected using multiple preparative liquid
102 chromatography system (LC-Forte/R, YMC, Japan).

103 The protein profile in the collected peak fraction was analyzed by SDS-PAGE. The
104 proteins in the samples were separated on a 4-20% acrylamide gradient gel (Biorad
105 Laboratories, Richmond, CA, USA) using a Biorad mini gel electrophoresis unit and a
106 ChemiDac XRS + system (Biorad Laboratories, Richmond, CA, USA) was used for the
107 visualization of the bands.

109 **Animals and experimental diets**

110 Male Sprague-Dawley rats (4 wk-old) were obtained from Koatech (Pyongtaek,
111 Korea). Animals were housed at a temperature of 23°C and relative humidity of 50 ± 10%, and
112 maintained under a 12-hour light-dark cycle, with feed and water available *ad libitum*. After a
113 week of acclimation, the rats were randomly divided into five groups ($n = 8$ for each group):
114 CW-10:0, CW-8:2, CW-5:5, CW-2:8, and nitrogen-free (N-free). The composition of
115 experimental diets is shown in Table 1. Diets based on AIN-93M (Saeronbio Inc., Uiwang,
116 Korea) were formulated to contain 10% protein according to the official PER AOAC 960.48
117 method. After four weeks, the rats were fasted overnight and anesthetized with 10 mg/kg
118 xylazine (Bayer Korea, Seoul, Korea) and 100 mg/kg ketamine (Yuhan Co., Seoul, Korea). The
119 animal experiment was conducted under the guidance of the Hanyang University Animal Care
120 and Use Committee (HY-IACUC-19-0159).

121

122 **Growth performance**

123 Body weight and feed intake were measured once a week throughout the experiment.

124 Body weight gain was calculated using body weight recorded at the beginning and the end of
125 the experiment. The feed efficiency ratio was calculated using Eq. (1).

$$126 \text{ FER} = \frac{\text{Wt. gain (g)}}{\text{Feed intake (g)}} \times 100 \quad (1)$$

127

128 **Protein quality evaluation**

129 To evaluate the protein utilization, the protein efficiency ratio (PER) and net protein
130 ratio (NPR) were assessed according to the official procedures recommended by the AOAC
131 Official Method 960.48 and calculated using Eq. (2, 3). The weight loss of the N-free group
132 was used to determine NPR. To evaluate the protein digestibility, the rats were housed
133 individually in metabolic cages to collect separate feces for three days at the second week. The
134 collected fecal samples were dried and ground before total nitrogen analysis. The total nitrogen
135 of the fecal samples was analyzed by the Kjeldahl method (AOAC). True digestibility (TD)
136 was calculated using Eq. (4). The result of the fecal sample from the N-free group was used to
137 confirm endogenous nitrogen.

$$138 \text{ PER} = \frac{\text{Wt. gain (g)}}{\text{Protein intake (g)}} \times 100 \quad (2)$$

$$139 \text{ NPR} = \frac{\text{Wt. gain (g)} - \text{Wt loss on N-free diet (g)}}{\text{Protein intake (g)}} \quad (3)$$

140
$$TD (\%) = \frac{N \text{ intake} - (\text{fecal N} - \text{endogenous fecal N})}{N \text{ intake}} \times 100 \quad (4)$$

141

142 **Body composition**

143 Body composition including fat mass (g), lean mass (g), and bone mineral density
144 (BMD) (g/cm^2) was measured by dual-energy X-ray absorptiometry (DEXA; InAlyzer,
145 Medikors Inc., Korea) before the sacrifice.

146

147 **Statistical analysis**

148 All data were analyzed using a one-way ANOVA followed by a Tukey's post hoc test
149 for multiple comparisons. Values at $p < 0.05$ were considered to be significant. GraphPad Prism
150 8 was used for the data analysis (GraphPad Software, La Jolla, CA, USA).

151

152 **Results and Discussion**

153 **Protein solubility of milk protein dispersion with different CW ratios**

154 High protein beverages are gaining popularity in the market and dairy proteins are one
155 of the attractive options for the production of high protein beverages. However, the decrease in
156 protein solubility by heat-mediated protein-protein interactions is a major factor to limit
157 product stability. The solubility of protein dispersions (5% protein, w/w) was measured by
158 quantifying proteins before and after centrifugation. Protein solubility increased as the
159 proportion of whey proteins increased in the dispersion (Fig. 1).

160 Micellar casein isolate (MCI) is a high protein dairy ingredient manufactured by
161 microfiltration. Since casein micelles in MCI are close to the native state, MCI has been
162 suggested as an alternative for traditional casein isolate prepared using acid or rennet (Carter
163 et al., 2021). Low reconstitution and solubility of MCI were reported and were found to be due
164 to the slow dissolution rate of casein micelles from the powder surface (Schokker et al., 2011;
165 Zhang et al., 2018). The lower storage temperature delayed loss of the rehydration property by
166 preventing surface hardening from the casein micelle surface (Burgain et al., 2016).

167 Heat treatment (95°C, 30 min) lowered solubility except for CW-10:0. The gap in
168 solubility before and after heating also increased with increasing whey proteins in CW-8:2 and
169 CW-5:5 but no further increase was observed in CW-2:8. This result suggests that MCI is quite
170 heat stable at the tested pH (pH 6.7) and concentration (5%, w/w). However, Sauer and Moraru
171 (2012) reported that high temperature treatment such as ultra high temperature (UHT) and
172 retort heating caused instability of the MCI dispersions (10%, w/w). The pH-induced alteration
173 of mineral balance and casein dissociation from the casein micelle surface was responsible for
174 the heat instability of MCI.

175 Heat-induced decreased solubility is probably associated with the formation of high
176 molecular weight protein aggregates. Liyanaarachchi et al. (2015) demonstrated that the
177 average particle size of heat-induced whey protein aggregates can be decreased by increasing
178 the proportion of casein in the protein dispersion (10% total solid). Caseins exerted chaperone-
179 like activity in heat-induced whey protein aggregation and cause aggregated whey protein to
180 be soluble.

181

182 **Changes in particle size distribution of protein dispersion with different CW ratios**

183 Particle size distribution of protein dispersions varied depending on the CW ratios.
184 Before heating, most particles in CW-10:0, CW-8:2, and CW-5:5 were present in the
185 submicron range while a small volume of larger particles of 2-6 μm was noted in CW-2:8
186 dispersion (Fig. 2). Substantial changes in particle size distribution by heating were found only
187 in CW-2:8 and displayed three broad peaks. This suggests particles with different levels of
188 whey protein aggregation are produced when sufficient whey proteins are present in the
189 dispersion.

190 Singh et al. (2019) reported that UHT processed CW-8:2 and CW-5:5 displayed similar
191 particle size distribution at the sub-micron range, but particle size distribution was significantly
192 increased when the proportions of whey proteins in the mixtures were greater than 50% (CW-
193 4:6, D (0.9) = 110 μm). They concluded that casein acted as a chaperon to inhibit the formation
194 of whey protein-mediated large protein aggregates. Our result was also consistent with a
195 previous report by Beaulieu et al. (1999) that heating (95°C, 5 min) of model milk protein
196 dispersion produced protein aggregates of various sizes, and the occurrence of heterogeneous
197 aggregates increased from CW-80:20 to 20:80. The formation of large aggregates probably
198 increases the risk of deposit accumulation on the heat exchanger (Khaldi et al., 2015).

200 **Protein profile analysis of the CW-2:8 dispersion**

201 Various sizes of large protein aggregates were formed by the heating of CW-2:8
202 dispersion. To analyze the involvement of individual proteins for aggregate formation,
203 unheated and heated CW-2:8 dispersion were separated using size exclusion chromatography.
204 The protein profile of the peak fractions was analyzed by SDS-PAGE.

205 Before heating, CW-2:8 eluted as one peak, and the intensity of the casein bands
206 decreased as elution time passed (Fig. 3 (A) and (C)). This indicated that whey proteins were

207 present mainly as unaggregated forms. The peak fraction of CW-2:8 decreased by heating and
208 eluted in broad elution time from 30 to 80 min (Fig. 3 (B)). Interestingly, early peak fractions
209 (F3, F4, and F5) consisted of whey proteins whereas later peak fractions (F7, F8, and F9)
210 contained both caseins and whey proteins. This result suggests that self-aggregation of whey
211 proteins is the major contributor to the formation of large molecular weight aggregates
212 compared to the contribution of casein micelle and whey proteins where the interactions are
213 relatively small. Havea et al. (2001) characterized heat-induced whey protein aggregates. They
214 found that homo- and heteropolymers of β -lactoglobulin (β -LG), α -lactalbumin (α -LA), and
215 bovine serum albumin (BSA) were produced via disulfide bonds during the heating of whey
216 protein concentrate. This report suggested that whey protein aggregates with diverse sizes can
217 be formed by self-aggregation of whey proteins.

218 Gaspard et al. (2017) reported that the stability of heat-induced milk protein aggregates
219 increased as the proportion of casein increased in the aggregates. The presence of κ -casein or
220 sodium caseinate protected the whey protein from heat-induced aggregation, and these effects
221 were closely related to decreased hydrophobic interaction (Guyomarc'h et al., 2009). Kehoe
222 and Foegeding (2011) reported that β -casein acts as a chaperone and controls the size of whey
223 protein self-aggregation upon heating. Competition occurred between β -casein and whey
224 proteins during the aggregation process.

225 Based on the above results, an increase of whey proteins up to CW-2:8 may cause
226 protein instability, especially in long shelf-life UHT-sterilized protein beverages. However,
227 there was no sign of protein stability problems by heating in CW-10:0, CW-8:2, and CW-5:5.

228

229 **Effect of modified CW ratios on growth performance**

230 The effects of CW ratio on body weight, body weight gain, feed intake, and feed

231 efficiency ratio were examined. As shown in Table 2, no significant difference was observed
232 in body weight and body weight gain in all experimental groups. In addition, feed intake was
233 unchanged among the samples containing both casein and whey protein. Only the sole casein
234 fed group showed greater feed intake compared to the other groups. However, the groups
235 that had the higher proportion of whey proteins (CW-5:5 and CW-2:8) showed higher feed
236 efficiency ratios than the other groups.

237 This observation was consistent with a previous study that body weight gain in rats
238 reared with modified CW ratios (CW2:8, CW4:6, CW6:4, and CW8:2) did not show a
239 significant difference (Yajima et al., 1998). However, Eller and Reimer (2010) demonstrated
240 that complete dairy proteins consisting of casein and whey reduced weight gain in high-fat and
241 high-sucrose diet-fed rats compared to casein or whey protein alone. Administration of whey
242 protein showed reduced weight gain compared to a casein control in high-fat fed mice, and this
243 weight gain reduction was associated with changes in gut microbiota (Tranberg et al., 2013).
244 The difference in diet composition (normal vs. high fat) and duration of feeding trial (8 or 14
245 vs. 4 wks) may be responsible for the discrepancy in the results between the present study and
246 previous reports (Eller and Reimer, 2010). Taken together, diets with modified casein to whey
247 protein ratios did not alter body weight and weight gain; however, diets with greater than or
248 equal to 50% of whey protein showed a lower feed intake and higher feed efficiency ratio.

249

250 **Effect of modified CW ratios on protein quality**

251 The protein efficiency ratio (PER), representing the contribution of protein diet in rat
252 growth, has been widely used as a standard method for protein quality assessment. A more
253 precise method than PER has been the net protein ratio (NPR) by considering weight loss of
254 rats from the non-protein diet in weight gain of rats (Gilani, 2012). The amino acid composition

255 and digestibility also affect the nutritional quality of proteins. Thus, the effects of modified
256 casein: whey protein ratios on PER, NPR, and true digestibility (TD) were compared. As shown
257 in Table 3, the PER and NPR were higher for the CW-5:5 and CW-2:8 than for CW-10:0 and
258 CW-8:2. No significant difference was found in nitrogen intake among the groups. Fecal
259 nitrogen was lowered as the portion of whey protein in the diet was increased. Although TD
260 was close to 100% in all groups, CW-2:8 showed significantly higher true digestibility than
261 other samples ($p < 0.05$). Thus, CW-5:5 and CW-2:8 had greater effects on protein utilization
262 than other formulations.

263 It has been reported that whey protein had significantly higher PER, NPR, and TD
264 compared to casein and CW-7:3 (Haraguchi et al., 2010). Unlike the results of our study, CW-
265 7:3 did not show higher PER and NPR than casein. However, they compared only three
266 different diet groups and the effect of different CW ratios on protein digestibility was not
267 further investigated. It is assumed that the portion (30%) of whey protein in the diet was not
268 sufficient to make a difference over the casein group. In accordance with our study, CW-6:4
269 showed higher PER than the casein group in growing rats (Van Dael et al., 2005). This may be
270 due to greater sulfur-containing amino acid content in whey proteins since amino acids such as
271 cysteine and methionine had greater effects in the improvement of PER (Walzem et al., 2002;
272 Potter and Kies, 1990).

273 Phosanam et al. (2021) examined the influence of CW ratio (40:60, 60:40, and 80:20)
274 using an *in vitro* digestion model. The samples with high casein ratios lowered digestibility by
275 extensive gastric coagulation. Huppertz and Chia (2020) reported that gastric coagulation
276 critically influences further digestion by regulating gastric emptying. The formation of casein
277 clots and slower gastric emptying delay the digestion rate of caseins but casein ingestion
278 resulted in a prolonged postprandial increase in plasma amino acids compared with rapidly

279 digested whey proteins with a short plasma amino acid increase (Boirie et al., 1997). Gorissen
280 et al. (2020) demonstrated that greater radio-labeled phenylalanine (Phe) was in systematic
281 circulation when a mixture of casein and whey protein was administered compared with whey
282 protein or casein alone in a human clinical trial. This result suggests that type of protein
283 critically influences protein digestion and the kinetics of amino acid absorption. They also
284 observed that postprandial Phe rise varied depending on protein dosage and age.

285

286 **Effect of modified CW ratios on body composition**

287 Changes in the body composition of the rats fed experimental diets for 4 weeks are
288 shown in Fig. 4. Both total fat and lean mass analyzed by DEXA did not show significant
289 differences in all treatment groups. It is consistent with a previous report that casein and whey
290 protein diet did not change fat mass (Wróblewska et al., 2018). However, according to the
291 results of previous studies, branch chain amino acids (BCAA) especially leucine (Leu) (11%
292 whey vs. 8% casein, w/w), play a key role in muscle protein synthesis (Boirie et al., 1997;
293 Layman, 2003). Whey proteins are considered as better protein sources than caseins for
294 increased muscle mass but their rapid disappearance in plasma possibly limits utilization of
295 BCAA. In accordance with this speculation, too rapid digestion of whey proteins (milk soluble
296 protein isolate) could not meet the postprandial anabolic requirement (Lacroix et al., 2006).

297 Thus, modulation of the optimum CW ratio for better utilization of BCAA provides
298 beneficial effects for increased muscle synthesis. Van Dael et al. (2005) reported that higher
299 PER and improved protein utilization were obtained when the diet composition with CW-40:60
300 was compared with the sole casein diet in growing rats. Although the exact reasons for no
301 difference in muscle mass in the present study are uncertain, decreased feed intake in CW-5:5
302 and CW-2:8 may have counteracted the improved protein utilization in the CW-5:5 and CW-

303 2:8 diet group.

304 Zhang et al. (2007) reported that dietary Leu supplementation effectively improved
305 high fat diet-induced obesity and glucose metabolism whereas increased Leu intake did not
306 show notable effects in normal diet-fed mice. In another study, Leu-fortified whey protein
307 promoted muscle protein synthesis but administration of Leu alone did not show a positive
308 effect on muscle synthesis in aged mice (Dijk et al., 2018). These results suggest important
309 findings for product application. The effect of dietary protein-induced muscle protein synthesis
310 varied depending on the age and nutritional status of the target groups, and therefore
311 modulation of casein:whey protein formulation might have more positive effects on obese and
312 elderly populations than healthy people.

313 BMD decreased as the proportion of whey proteins increased in the diet. A significant
314 difference was found between the CW-10:0 and CW-2:8 groups ($p < 0.05$). However, there
315 was no significant difference among the other groups (Fig. 4 (C)). Based on the product
316 information, the calcium content of MCI and WPI is 1,900 mg/100 g and 46 mg/100 g,
317 respectively. The difference in total calcium content in the protein source probably affects
318 calcium availability. In terms of the qualitative aspect, the type of mineral (organic vs.
319 inorganic) is also important for the absorption and retention efficiency for animals (Liu et al.,
320 2014). Micellar casein contains calcium in the form of colloidal calcium phosphate (organic
321 form) which facilitates better absorption than the inorganic form. The same effect was
322 demonstrated in calcium-fortified milk using mice (Singh et al., 2007). Our findings are
323 consistent with previous results that the casein fed group showed higher total and trabecular
324 BMD compared to the whey protein fed group in piglets (Budek et al., 2007). McKinnon et
325 al. (2010) reported that diets containing goat milk casein (80% and 57%) resulted in increased
326 calcium absorption in growing rats compared to the casein-free diet containing equal protein

327 and calcium content. In summary, there was no significant difference in BMD among CW-
328 10:0, CW-8:2, and CW-5:5 in growing rats while calcium fortification might be beneficial for
329 CW-2:8 for adequate bone growth and development.

330

331 Conclusion

332 Modification of the casein to whey protein ratio affected the extent of protein
333 aggregation and heated CW-2:8 showed significantly increased larger particle ($> 100 \mu\text{m}$) size
334 distribution. The largest protein aggregates were formed by whey protein self-aggregation.
335 There was no significant difference in protein aggregation when the CW ratios changed from
336 10:0 to 5:5. In terms of protein quality, protein utilization and digestibility showed an
337 increasing trend as the proportion of whey proteins increased in the diet. There was no
338 significant difference in BMD between native cow's milk (CW-8:2) and CW-5:5 but CW-2:8
339 resulted in significantly lower BMD. Future detailed studies will be required to investigate the
340 effects of modified casein and whey protein formulations on metabolic health and disease
341 prevention.

342

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478 in mice via multimechanisms. *Diabetes* 56:1647-1654.

479 **Table captions**

480 **Table 1. Composition of experimental diets (g/kg diet)**

481 All experimental diets were based on AIN-93M composition; CW, casein:whey protein.

482

483 **Table 2. Growth performance of the rats fed diets with modified casein to whey protein**
484 **ratio for 4 weeks ($n = 8$ for each group)**

485 CW, casein:whey protein; Data are expressed as mean \pm SEM. The values with different letters
486 in the same row indicate significant differences at $p < 0.05$.

487

488 **Table 3. Effect of modified casein and whey protein ratio on protein quality in rats fed**
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490 CW, casein:whey protein; Data are expressed as mean \pm SEM. The values with different letters
491 in the same row indicate significant differences at $p < 0.05$.

492

493 **Table 1. Composition of experimental diets (g/kg diet)**

Ingredients	AIN-93M	CW-10:0	CW-8:2	CW-5:5	CW-2:8	N-free
Casein	140	117.37	93.90	58.69	23.47	-
Whey protein	-	-	22.35	55.87	89.39	-
Sucrose	100	100	100	100	100	100
Dextrose	155	155	155	155	155	155
Corn starch	465.69	488.32	489.44	491.10	492.83	605.69
Cellulose	50	50	50	50	50	50
Soybean oil	40	40	40	40	40	40
Mineral mix	35	35	35	35	35	35
Vitamin mix	10	410	410	410	410	410
L-Cystein	1.8	1.8	1.8	1.8	1.8	1.8
Choline bitartrate	2.5	2.5	2.5	2.5	2.5	2.5
TBHQ	0.008	0.008	0.008	0.008	0.008	0.008
Total	1000	1000	1000	1000	1000	1000

494 All experimental diets were based on AIN-93M composition; CW, casein:whey protein.

495

496 **Table 2. Growth performance of the rats fed diets with modified casein to whey protein**
497 **ratio for 4 weeks ($n = 8$ for each group)**

	CW-10:0	CW-8:2	CW-5:5	CW-2:8
Body weight (g)	279 ± 9.18 ^a	269 ± 2.66 ^a	275 ± 3.97 ^a	264 ± 3.62 ^a
Body weight gain (g)	170 ± 7.23 ^a	163 ± 2.07 ^a	167 ± 3.33 ^a	154 ± 0.92 ^a
Feed intake (g/day)	20.2 ± 0.50 ^a	19.1 ± 0.16 ^{ab}	18.3 ± 0.31 ^b	18.3 ± 1.79 ^b
Feed efficiency ratio (%)	29.9 ± 0.49 ^b	30.4 ± 0.33 ^b	32.9 ± 0.88 ^a	30.6 ± 0.22 ^{ab}

498 CW, casein:whey protein; Data are expressed as mean ± SEM. The values with different letters
499 in the same row indicate significant differences at $p < 0.05$.

500

501 **Table 3. Effect of modified casein and whey protein ratio on protein quality in rats fed**
 502 **diets with modified casein to whey protein ratio for 4 weeks ($n = 8$ for each group)**

	CW-10:0	CW-8:2	CW-5:5	CW-2:8
Protein efficiency ratio (%)	2.99 ± 0.05 ^b	3.04 ± 0.03 ^b	3.29 ± 0.09 ^a	3.06 ± 0.02 ^{ab}
Net protein ratio (%)	3.31 ± 0.04 ^b	3.37 ± 0.03 ^b	3.57 ± 0.07 ^a	3.40 ± 0.02 ^{ab}
Nitrogen intake (g/rat)	1.02 ± 0.04 ^a	0.98 ± 0.02 ^a	0.97 ± 0.02 ^a	1.03 ± 0.03 ^a
Fecal nitrogen (g/rat)	0.12 ± 0.00 ^a	0.09 ± 0.01 ^b	0.09 ± 0.00 ^b	0.07 ± 0.01 ^c
True digestibility (%)	92.7 ± 0.22 ^b	93.2 ± 0.25 ^b	93.5 ± 0.15 ^b	94.8 ± 0.22 ^a

503 CW, casein:whey protein; Data are expressed as mean ± SEM. The values with different letters
 504 in the same row indicate significant differences at $p < 0.05$.

505

506

507 **Figure captions**

508 **Fig. 1. Changes in solubility of protein dispersion (5%, w/v) before and after heating ($n =$**
509 **3).** CW, casein:whey protein; Protein dispersions with different casein:whey protein ratios were
510 heated in a 95°C water for 30 min. The values with different letters indicate significant
511 differences at $p < 0.05$.

512 **Fig. 2. Changes in volume particle size distribution of protein dispersion (5%, w/v) before**
513 **and after heating ($n = 3$).** CW, casein:whey protein; Protein dispersions with different
514 casein:whey protein ratios were heated in a 95°C water for 30 min.

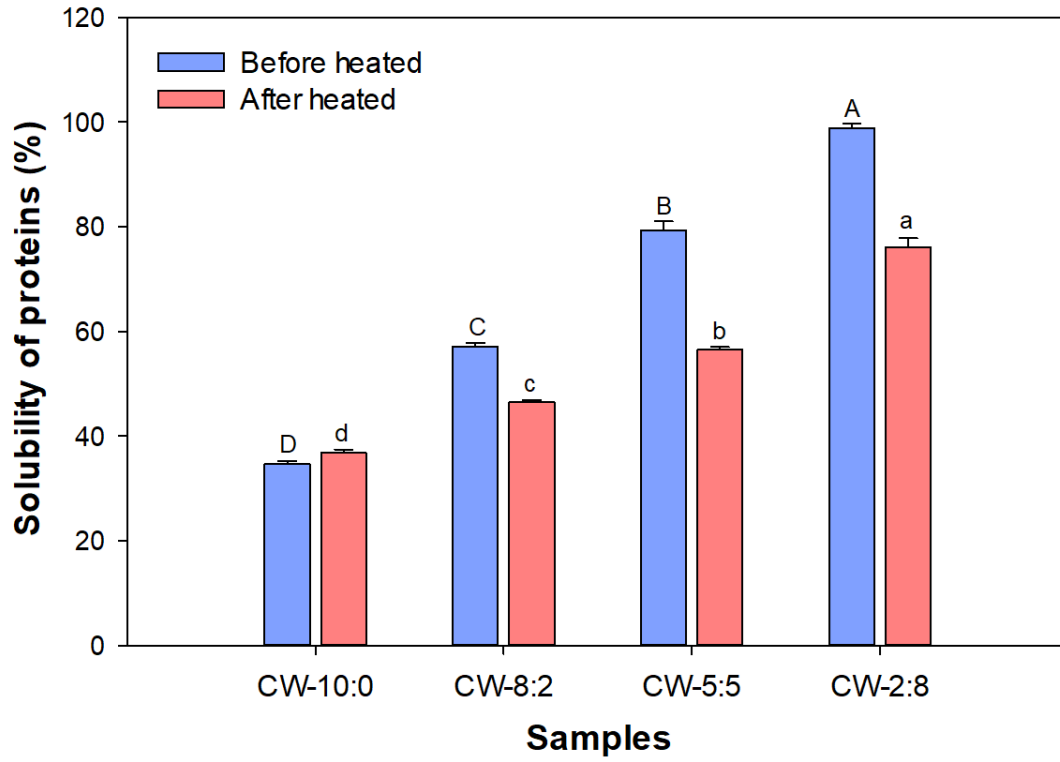
515 **Fig. 3. Protein profile analysis of CW-2:8 dispersion ($n = 3$).** (A) size exclusion
516 chromatogram of unheated CW-2:8 dispersion, (B) size exclusion chromatogram of heated
517 CW-2:8 dispersion, and (C) SDS-PAGE electrophoregram of peak fraction obtained from
518 unheated and heated CW-2:8 dispersion.

519 **Fig. 4. Effect of modified casein:whey protein ratios on body composition ($n = 8$ for each**
520 **group).** (A) fat mass, (B) lean mass, and (C) bone mineral density. Data are expressed as mean
521 \pm SEM. The values with different letters indicate significant differences at $p < 0.05$.

522

523 **Fig. 1**

524



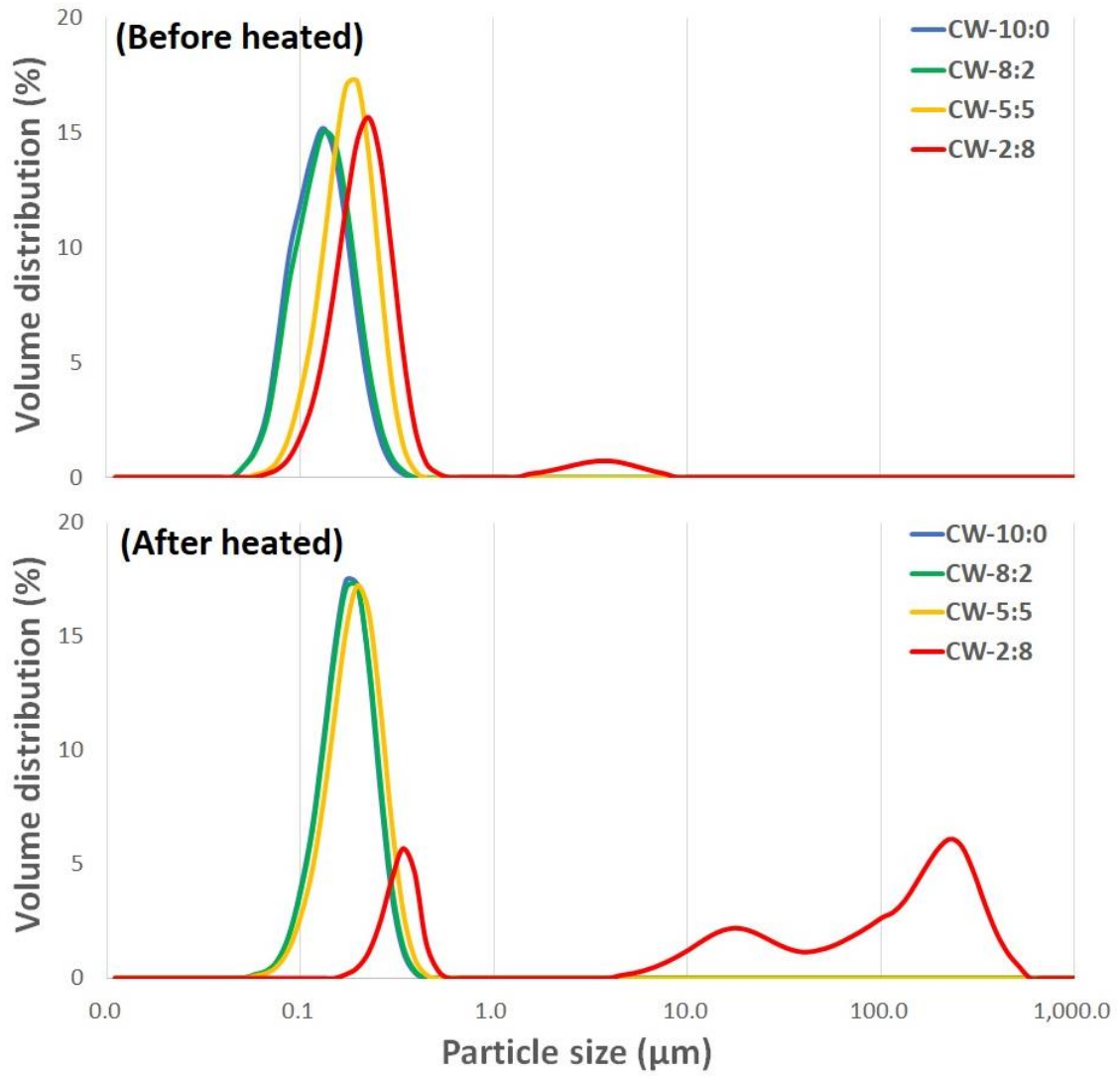
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527 **Fig. 2**

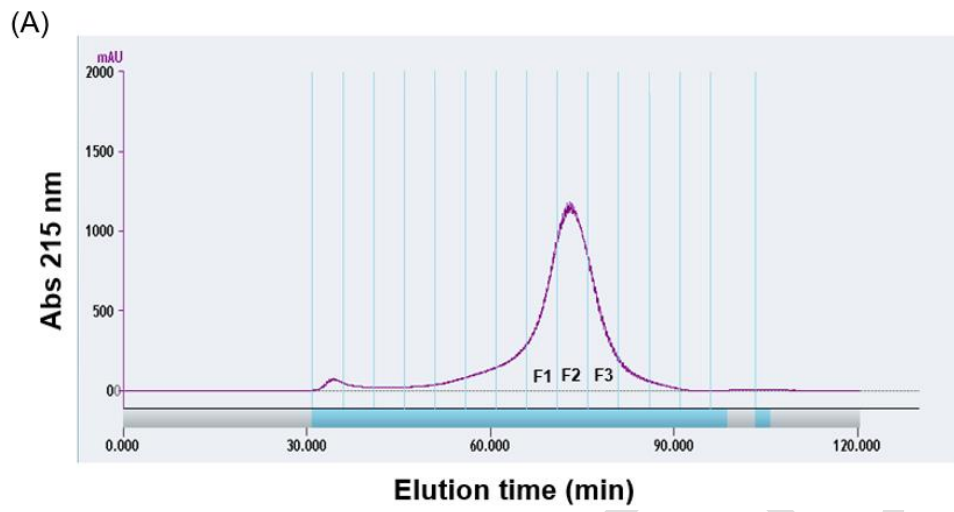
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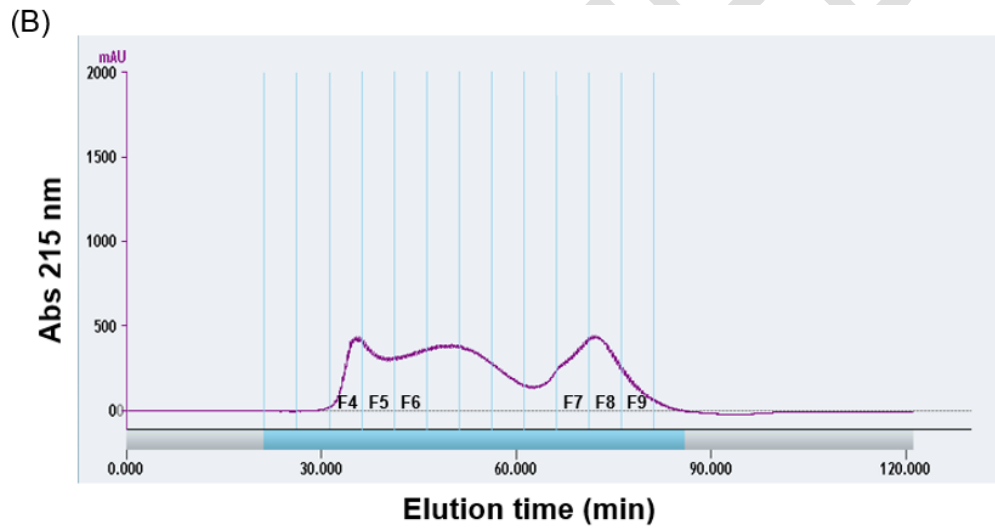
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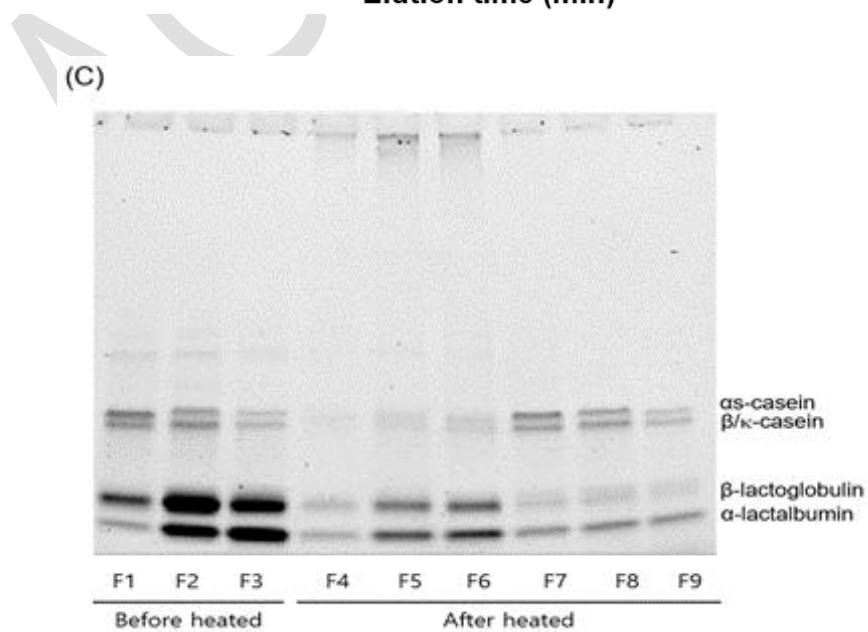
531 **Fig. 3**



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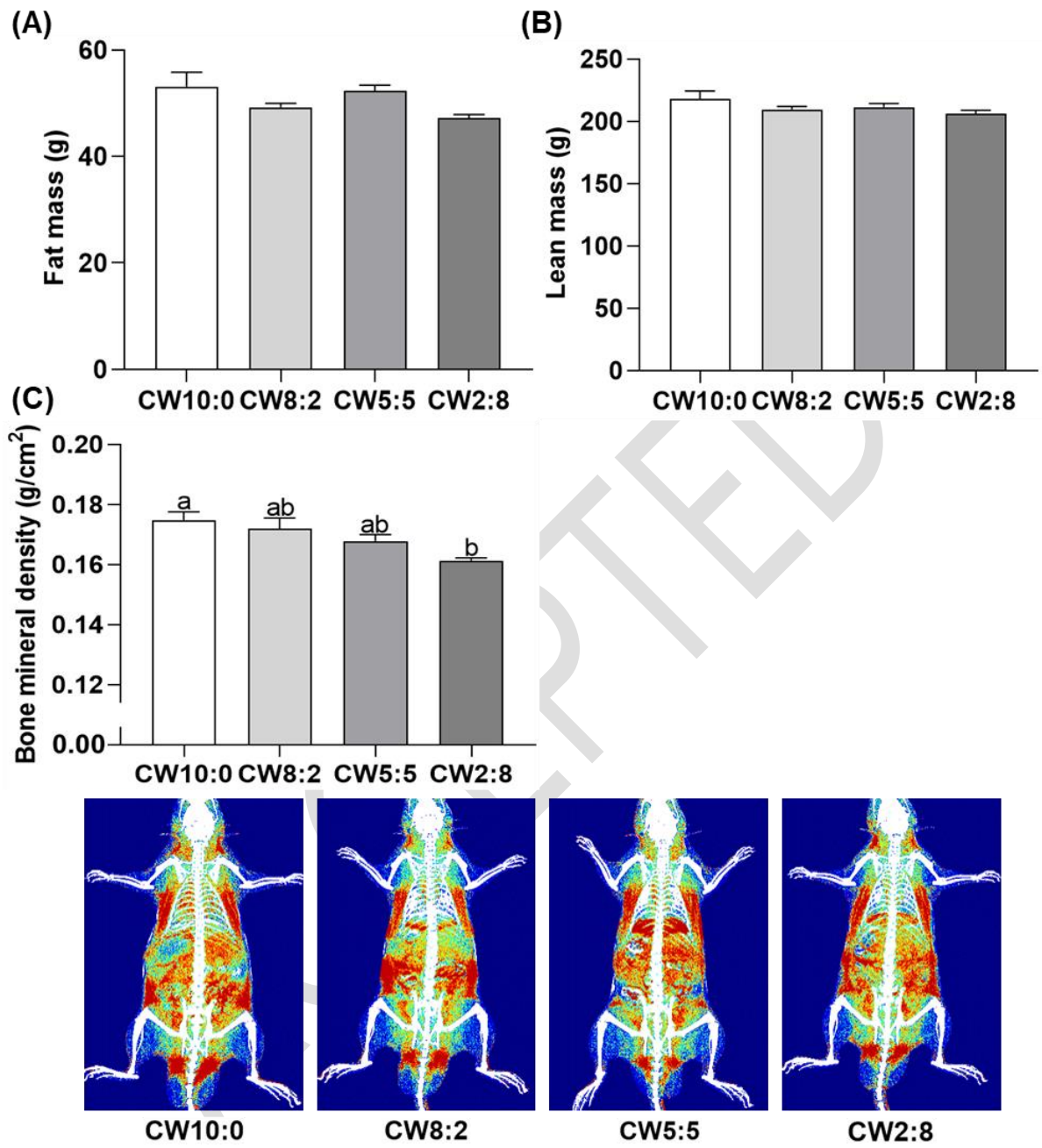
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536 Fig. 4



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