

Antihypertensive Effect of Milk Fermented by *Lactiplantibacillus plantarum* K79 on Spontaneously Hypertensive Rats

Sang-Dong Lim^{1,*}, Kyungwon Lee¹, Taewon Han¹, Hyunjhong Jhun¹, Ah-Ram Han¹,

Yongjin Hwang² and Sangpil Hong¹

¹Korea Food Research Institute, Wanju 55365, Korea

²Novalacto co., Ltd., Daejeon 34016, Korea

Running title: Antihypertensive effect of milk fermented by *Lp. plantarum* K79

*Corresponding author:

Sang-Dong Lim

Korea Food Research Institute, Wanju 55365, Korea

Tel.: +82-63-219-9082

Fax: +82-63-219-9288

E-mail: limsd@kfri.re.kr

*ORCID

Sang-Dong Lim

<https://orcid.org/0000-0002-1500-4413>

Kyungwon Lee <https://orcid.org/0000-0007-9404-4811>

Taewon Han

[https://orcid.org/ 0000-0002-7700-1960](https://orcid.org/0000-0002-7700-1960)

Hyunjhong Jhun

[https://orcid.org/ 0000-0002-3694-9715](https://orcid.org/0000-0002-3694-9715)

Ah-Ram Han

[https://orcid.org/ 0000-0001-7111-7095](https://orcid.org/0000-0001-7111-7095)

Yongjin Hwang

[https://orcid.org/ 0009-0001-3168-8587](https://orcid.org/0009-0001-3168-8587)

Sangpil Hong

[https://orcid.org/ 0000-0002-4060-0129](https://orcid.org/0000-0002-4060-0129)

ACCEPTED

Abstract

The aim of this study is to investigate whether milk fermented by *Lactiplantibacillus plantarum* K79, which exhibits ACE inhibitory activity, has an effect on lowering the blood pressure of hypertensive rats and to investigate biomarker changes in their blood.

Experimental group: normal group (NG, WKY): distilled water, control group (NCG, SHR): distilled water, high treatment group (HTG, SHR): 500 mg/kg/day, medium treatment group (MTG, SHR): 335 mg/kg/day, low treatment group (LTG, SHR): 170 mg/kg/day, positive control group (PCG, SHR): Enalapril, 10 mg/kg/day.

The experimental animals used in this study were divided into groups composed of 8 animals. In terms of weight change, a significant difference was observed between the normal group and the SHR group, but there was no significant difference between the SHR group. After 8 weeks of feeding, Blood pressure was lowered more significantly in the HTG(209.9 ± 13.3 mmHg) than in the NCG(230.8 ± 7.3 mmHg). The treatment group has an effect of lowering blood pressure by significantly suppressing blood pressure-related biomarker protein expression than NG. The results obtained can be used as an antihypertensive material in a variety of food raw materials.

Key words: fermented milk, hypertension, *Lactiplantibacillus plantarum*, spontaneously hypertensive rats, angiotensin converting enzyme

1 **Introduction**

2 Hypertension is a risk factor in the development of cardiovascular diseases such as coronary
3 artery disease, left ventricular hypertrophy, valvular heart disease, atrial fibrillation, stroke, and
4 arrhythmias such as renal failure. Without timely diagnosis and appropriate treatment, it can
5 cause illness or death (Farooq and Ray, 2015; Kjeldsen, 2018). It is estimated that 1.4 billion
6 people worldwide suffer from hypertension, but only 14% have it under control (WHO, 2021).
7 Angiotensin I converting enzyme plays an important physiological role in regulating blood
8 pressure through the renin-angiotensin system (Lee et al., 2004). It can convert angiotensin I
9 to angiotensin II, and can also inactivate the vasodilator bradykinin (Yang et al., 1970; Yang
10 et al., 1971). Therefore, inhibiting the angiotensin-converting enzyme (ACE) reduces the
11 activity of angiotensin II, although it can also lower blood pressure by increasing bradykinin
12 levels (Pyo and Lee, 2007).

13 ACE inhibitory activity is considered an effective way to treat hypertension. Synthetic ACE
14 inhibitors such as Captopril, Alacepril, Lisinopril, and Enalapril are currently being used to
15 treat hypertension, but they have undesirable side effects such as dysgeusia, rash, coughing,
16 hypotension, renal failure, and hyperkalemia (Brown and Vaughan, 1998; Chakraborty and
17 Roy, 2021; Xia et al, 2020). In contrast, naturally occurring ACE inhibitors are considered safe
18 (Chen et al, 2022). To date, ACE inhibitors have been found in fermented milk (Rendon-
19 Rosales et al, 2022), fish surimi (Oh et al, 2020), rabbit meat (Chen et al, 2022), and abalone
20 viscera (Iwamoto et al, 2023), and ACE inhibitory peptides have been isolated from these foods
21 and other sources. ACE inhibitory peptides derived from milk proteins are produced by
22 enzymatic hydrolysis, microbial fermentation or genetic engineering (Castellano et al., 2013).
23 In particular, many lactic acid bacteria produce ACE inhibitory peptides, such as Val-Pro-Pro-
24 and Ile-Pro-Pro, from fermented milk (Hirota et al., 2007; Mizushima et al., 2004). Bioactive

25 peptides derived from fermented milk by *Lactobacillus helveticus* have been reported to
26 decrease the blood pressure of Spontaneously Hypertensive Rats (SHR) (Narva et al. 2004). In
27 addition, angiotensin-converting enzyme inhibitory peptides derived from the enzymatic
28 hydrolysis of milk protein have been observed to decrease blood pressure in hypertensive mice
29 (Ramchandran and Shah, 2011; Wang et al., 2012). The aim of this study was to investigate
30 whether pasteurized fermented milk by *Lp. plantarum* K79, which exhibits ACE inhibitory
31 activity, has an effect on lowering blood pressure in hypertensive rats and to investigate
32 biomarker changes in their blood.

33

34 **Materials and Methods**

35 **Lactic acid bacteria**

36 *Lp. plantarum* K79 isolated from kimchi: This strain was selected after screening it for
37 probiotic properties such as acid and bile tolerance, antibacterial activity, antibiotic
38 tolerance and ACE inhibitory activity. The culture was maintained in an MRS broth
39 (Difco, USA), and the patent accession number is KACC 81222BP.

40

41 **Sample preparation**

42 13.5% skim milk powder and 1% glucose were mixed, sterilized at 90°C for 5 minutes,
43 and then cooled to 37°C. At this time, lyophilized *Lp. Plantarum* K79 strain activated for 1
44 hour was inoculated and cultured with stirring at 37°C until the pH reached 4.2. The culture
45 was heat-treated by stirring at 85°C for 20 minutes, cooled to below 50°C, and then spray-
46 dried and powdered. The powdered raw materials were suspended and used for testing.

47 **Animals and treatment**

48 The experiment included control groups composed of 5-week-old SHR (SLC, Japan) male
49 rats, and normal groups composed of Wistar-Kyoto rats(WKY), which were used after one
50 week of acclimatization. The experimental animals were fed with the AIN-93G diet, and
51 during the acclimatization period the filtered drinking water was changed every day so that
52 it could be consumed freely. During the breeding period, the temperature was $23\pm 1^{\circ}\text{C}$,
53 humidity was $50\pm 5\%$, noise was less than 60 phones, lighting time was 08:00~20:00 (12
54 hours a day), illuminance was 150~300 Lux, ventilation was 10~12 times per hour. This
55 experiment was conducted with the approval of the Korea Food Research Institute (KFRI-
56 M-22024) in compliance with the regulations on animal experimentation ethics. During the
57 breeding period, the experimental animals were fed with regular solid feed (Samtako,
58 Gyunggi, Korea), and their blood pressure was measured at the end of the acclimatization
59 period. Individual identification was indicated.

60 Experimental group: normal group (WKY): distilled water, control group (SHR): distilled
61 water, high treatment group (SHR): 500 mg/kg/day, medium treatment group (SHR): 335
62 mg/kg/day, low treatment group (SHR): 170 mg/kg/day, positive control group (SHR):
63 Enalapril, 10 mg/kg/day. Each group used in the experiment consisted of 8 animals. At this
64 time, the oral administration method, which is the clinical application route of this
65 formulation, was selected and each sample was directly administered into the stomach of
66 each experimental animal.

67

68

69 **Measurement of body weight, feed intake, and blood pressure**

70 All the experimental animals were weighed once per week before the start of
71 administration, after the start of administration, and until the end of the test. For each
72 breeding box, the total amount of feeding and watering on the day and the remaining amount
73 the next day were measured once per week for 4 weeks after the start of administration, and
74 the amount of daily consumption was indicated for each group. The systolic pressure of each
75 rat was measured with a tail cuff blood pressure meter (INDIR, Model LE5002, Spain), and
76 was maintained at 32° C for 30 minutes to detect the pulse of each rat's tail artery before
77 measurement.

79 **Analysis of blood serum and organ weight**

80 At the end of the 10-week experimental period, blood samples were collected from all the
81 rats and immediately placed in sterile tubes. Serum was collected by centrifugation at 2,000
82 x g for 15 min at 4 °C. The serum samples were then analyzed for total cholesterol (TC),
83 triglyceride (TG) concentration, High Density Lipoprotein (HDL) cholesterol, and Low
84 Density Lipoprotein (LDL) cholesterol by GCCL (Korea). Also, the liver, kidneys, lungs,
85 heart and testes of the rats were quickly isolated after sacrifice and weighed.

87 **Blood pressure-related biomarkers**

88 The angiotensinogen protein expression, angiotensin II protein, and renin protein content
89 in the plasma were measured using a Rat Angiotensinogen ELISA kit, a Rat Angiotensin II
90 ELISA kit, and a Rat Renin ELISA kit (Mybiosource, USA), respectively.

91 **Protein expression in the kidney**

92 According to the method of Averill et al. (2003), kidney tissue was left in 4% formalin for 48
93 hours before being transferred to 70% ethanol. Blocks of cardiac tissue were imbedded in
94 paraffin; 5- μ m sections were transferred to slides and the paraffin was removed by sequential
95 washes with xylene, 100% ethanol, 95% ethanol, 75% ethanol, and double-distilled water.
96 Sections of tissue were incubated with 3% hydrogen peroxide for 5 minutes, washed with PBS
97 (pH 7.2), dried, and then incubated with 5% normal goat serum for 1 hour at room temperature.
98 The sections were washed with PBS and incubated overnight at 4°C with an affinity-purified
99 rabbit polyclonal antibody to angiotensin-1 at 1:25 dilution of the antibody in 1% BSA. The
100 Angiotensin-1 antibody was purified. The next day, the tissues were washed with PBS and
101 incubated for 3 hours at 4°C with a biotinylated anti-rabbit antibody at a dilution of 1:400 in
102 1% BSA. The slides were rinsed with PBS, blotted dry, and reacted immunocytochemically by
103 avidin-biotin solution and stained brown with 3,3'-diaminobenzidine (Sigma Chemical Co) in
104 Tris buffered saline (0.05 mol/L, pH 7.6 to 7.7). The reaction was stopped in PBS, and the
105 sections were rinsed in double-distilled water before being counterstained with hematoxylin
106 (Sigma). The tissue sections were dehydrated in ethanol (70% to 100%) and then HistoClear
107 (National Diagnostics). Finally, they were mounted under coverslips with Histomount.

108

109 **Statistical analysis**

110 The results are expressed as the mean \pm standard deviation (SD) per experimental group
111 using SPSS version 23 (IBM, Armonk, NY, USA). The significance of the differences was
112 analyzed by conducting a one-way analysis of variance (ANOVA) using Duncan's multiple
113 range tests. Significance was considered at $p < 0.05$.

114 **Results and Discussion**

115 **Change of body weight, feed intake, and blood pressure**

116 Looking at the change in weight according to the experimental period, there was a significant
117 difference between the normal group (WKY), which was fed with distilled water, and the
118 hypertensive rat (SHR) group within the same period (Fig. 1). This was found to be consistent
119 with Gattone's (1986) report that WKY had a similar weight to SHR at birth, but that its weight
120 became greater than that of SHR over time. However, there was no significant difference in
121 average body weight between the treatment group and the control group, indicating that milk
122 fermented by *Lp. plantarum* K79 used in the experiment did not have a significant effect on
123 the weight of the rats.

124 There was a slight difference in intake, but no significant change was observed, indicating
125 that the fermented milk containing *Lp. plantarum* K79 had nothing to do with the weight gain
126 of the rats. However, at 7 weeks of feeding, the low-treatment group showed a significant
127 increase compared to the normal and medium, high-treatment groups (Table 1).

128 The results of measuring the blood pressure according to the experimental period showed
129 that there was a significant change in blood pressure in the NG, followed by the PCG, treatment
130 group, and NCG, until 10 weeks. When comparing the treatment group and the NCG, the
131 treatment group showed a tendency to lower blood pressure until 6 weeks, but it was found that
132 there was no significance (Table 2). However, after 8 weeks, Blood pressure was lowered more
133 significantly in the HTG(209.9 ± 13.3 mmHg) than in the NCG(230.8 ± 7.3 mmHg). Ishiguro et
134 al (2012) reported that systolic blood pressure (SBP) and diastolic blood pressure (DBP) were
135 reduced by more than 30 mmHg and 20 mmHg, respectively, after oral administration of
136 purified sweet potato protein hydrolyses to SHR. Tsai et al. (2006) found that the SBP of SHR

137 decreased by 19 mmHg after 8 weeks of oral administration of fermented soymilk with lactic
138 acid bacteria. These results were similar to the results obtained in this study. According to Yuan
139 et al. (2022) and Alshuniaber et al. (2021), this drop of blood pressure in rats is related to
140 inflammation and could be obtained by inhibiting the ACE activity of the renin-angiotensin
141 system.

142

143 **Change of blood serum and organ weight**

144 Regarding blood lipids, total cholesterol, HDL cholesterol and LDL cholesterol were all
145 significantly higher in the normal group than in the hypertensive rat group, while triglyceride
146 was significantly lower in the normal group (Table 3). This finding was consistent with the
147 report of Cho et al. (2018), who observed that the SHR group had lower TC and HDL-C levels
148 than the WKY group. Among the blood pressure rat groups, total cholesterol and LDL
149 cholesterol were slightly higher in the medium-dose treatment group, but the levels of HDL
150 cholesterol and triglyceride were insignificant.

151 When examining the weight of each organ of the SHR euthanized 10 weeks after
152 administration of the fermented product, the weight of the liver in the PCG was found to be
153 significantly higher than that in the NG, while the weight of the kidneys was significantly lower
154 in the NG than in the other groups (Table 4). There was no significant change in the weight in
155 the lungs of all groups, and the weight of the heart was significantly higher in the NCG and the
156 treatment group than in the PCG and the NG. The organs of the NG weigh less than those of
157 the other groups, but the testes weigh significantly more. This was consistent with the report
158 by Walter and Hamet (1986) that the heart, kidneys and liver weighed less in WKY than in
159 SHR.

160 **Change of blood pressure-related biomarkers**

161 As is well known, Ang II is produced systematically. Angiotensinogen (AGT), a substrate of
162 RAS, is released by the liver and broken down in the circulation to form Ang I by renin secreted
163 by the proximal glomerular apparatus of the kidneys. Vasodilator I (Ang I) is then readily
164 activated, and Ang II is caused by angiotensin-converting enzyme (ACE), which is expressed
165 at high levels primarily on the surface of endothelial cells in the pulmonary circulation. When
166 the concentration of these biomarkers increases, it affects the increase in blood pressure
167 (Ichihara et al., 2004; Yim and Yoo, 2008). Table 5 shows changes in blood pressure-related
168 biomarkers in the plasma of rats after 10 weeks. It was confirmed that the NG, PCG, and HTG
169 had significantly lower angiotensinogen protein expression than the NCG. Angiotensin-II
170 showed significantly lower expression in the NG, PCG, and treated groups than in the NCG.
171 Renin also was expressed the least in NG compared to NCG, followed by PCG and HTG.
172 Therefore, it is judged that significantly suppressing blood pressure-related biomarker protein
173 expression had the effect of lowering the blood pressure of the treatment group.

174

175 **Angiotensin-I protein expression in the kidney**

176 Human angiotensin is expressed in the tissues of diverse organs including those of the liver,
177 heart, blood vessel walls, brain, and kidneys, and in adipose tissue. Although all organ systems
178 have certain components of the Renin-Angiotensin System (RAS), the kidneys have all the
179 components of the RAS, and also show compartmentalization and intracellular accumulation
180 of tubular and interstitial networks (Kobori et al. 2007)

181 Fig. 2 shows the results of an experiment confirming the expression of angiotensin-I using
182 Immunohistochemistry. Looking at the intrarenal glomeruli, it was found that the protein
183 expression of angiotensin-I in the hypertensive rats fed only with distilled water, i.e. the NCG,

184 was increased to a visually observable level compared to the normal rats, the PCG, and the
185 treatment group. Therefore, it was observed that suppressing the protein expression of
186 angiotensin-I had an antihypertensive effect on the treated group.

187

188 **Conclusion**

189 The aim of this study was to investigate whether pasteurized milk fermented by *Lp.*
190 *plantarum* K79, which exhibited ACE inhibitory activity, has the effect on lowering the blood
191 pressure of hypertensive rats and to investigate biomarker changes in their blood. The reduction
192 in blood pressure was significant up to 10 weeks in the following order: NG, PCG, treatment
193 group, and NCG. After 8 weeks, the blood pressure of HTG was significantly lower than that
194 of NCG. The treatment group has an effect of lowering blood pressure by significantly
195 suppressing blood pressure-related biomarker protein expression than NG. The protein
196 expression of angiotensin-I in the kidney of hypertensive rats fed only with distilled water, i.e.
197 the NCG, was increased to a visually observable level compared to the other groups.

198 Therefore, the results of this study suggest that the fermented product by *Lp. plantarum* K79
199 can be applied as an antihypertensive material.

200

201 **Conflict of Interest**

202 The authors declare no potential conflicts of interest.

203

204

205 **Acknowledgements**

206 This work was supported by the Korea Institute of Planning and Evaluation for Technology
207 in Food, Agriculture, and Forestry (IPET) through the Technology Commercialization Support
208 Program (No. 821028-03) funded by the Ministry of Agriculture, Food and Rural Affairs
209 (MAFRA).

210

211 **Authors Contributions**

212 Conceptualization: Lim SD. Data curation: Hong SP, Jhun HJ. Formal analysis: Han AR,
213 Lee KW, Han TW. Methodology: Jhun HJ. Validation: Hong SP. Software: Han AR. Hong SP.
214 Investigation: Hwang YJ. Writing – original draft: Lim SD. Writing-review & editing: Lim
215 SD.

216

217 **Ethics Approval**

218 The Committee on the Ethics of Animal Experiments of the Korea Food Research Institute
219 has approved the animal experiments undertaken for the purposes of this study (KFRI-M-
220 22024).

221

222 **References**

223 Alshuniaber M, Alhaj O, Abdallah Q, Jahrami H. 2021. Effects of camel milk hydrolysate on
224 blood pressure and biochemical parameters in fructose-induced hypertensive rats. *Nutr. Food*
225 *Sci* 52:292–307.

226 Averill DB, Ishiyama Y, Chappell MC, Ferrario CM. 2003. Cardiac angiotensin-(1-7) in
227 ischemic cardiomyopathy. *Circulation* 108:2141-2146.

228 Brown NJ, Vaughan DE. 1998. Angiotensin-converting enzyme inhibitors. *Circulation*
229 97:1411-1420.

230 Castellano P, Aristoy MC, Sentandreu MA, Vignolo G, Toldra F. 2013. Peptides with
231 angiotensin I converting enzyme (ACE) inhibitory activity generated from porcine skeletal
232 muscle proteins by the action of meat-borne *Lactobacillus*. *J Prot* 89:183–190.

233 Chakraborty R, Roy S. 2021. Angiotensin-converting enzyme inhibitors from plants: A review
234 of their diversity, modes of action, prospects, and concerns in the management of diabetes-
235 centric complications. *J Integr Med* 19: 478–492.

236 Chen J, Yu X, Chen Q, Wu Q, He Q. 2022. Screening and mechanisms of novel angiotensin-I-
237 converting enzyme inhibitory peptides from rabbit meat proteins: A combined in silico and
238 vitro study. *Food Chem* 370: 131070.

239 Cho KH, Yadov D, Kim SJ, Kim JR. 2018. Blood pressure lowering effect of Cuban
240 Policosanol is accompanied by improvement of hepatic inflammation, lipoprotein profile, and
241 HDL quality in Spontaneously Hypertensive Rats. *Molecules* 23:1080

242 Farooq U, Ray SG. 2015. 2014 Guideline for the management of high blood pressure (eighth
243 joint national committee): take-home messages. *Med Clin N Am* 99:733–738.

244 Gattone II VH. 1986. Body weight of the Spontaneously Hypertensive Rat during the suckling
245 and weanling period. *Jpn Heart J* 27:881-884.

246 Hirota T, Ohki K, Kawagishi R, Kajimoto Y, Mizuno S, Nakamura Y, Kitakaze M. 2007. Casein
247 hydrolysate containing the antihypertensive tripeptides Val-Pro-Pro and Ile-Pro-Pro improves

248 vascular endothelial function independent of blood pressure-lowering effects: contribution of
249 the inhibitory action of angiotensin-converting enzyme. *Hypertens Res* 30:489–496.

250 Ichihara A, Kobori H, Nishiyama A, Navar LG. 2004. Renal renin-angiotensin system. *Contrib*
251 *Nephrol* 143:117–130.

252 Ishiguro K, SameshimA y, Kume T, Ikeda KI, Matsumoto J, Yoshimoto M. 2012. Hypotensive
253 effect of a sweetpotato protein digest in spontaneously hypertensive rats and purification of
254 angiotensin I-converting enzyme inhibitory peptides. *Food Chem* 3:774-779.

255 Iwamoto N, Sasaki A, Maizawa T, Hamada-Sato N. 2023. Abalone Viscera fermented with
256 *Aspergillus oryzae* 001 prevents pressure elevation by inhibiting angiotensin converting
257 enzyme. *Nutri* 15:947.

258 Kjeldsen SE. 2018. Hypertension and cardiovascular risk: General aspects. *Pharmacol.*
259 *Res* 129: 95–99.

260 Kobori H, Nangaku M, Navar LG, Nishiyama A. 2007. The intrarenal renin-angiotensin system:
261 from physiology to the pathobiology of hypertension and kidney disease. *Pharmacol Rev*
262 59:251-287.

263 Lee DH, Kima JH, Park JS, Choi YJ, Lee JS. 2004. Isolation and characterization of a novel
264 angiotensin I-converting enzyme inhibitory peptide derived from the edible mushroom
265 *Tricholoma giganteum*. *Peptides* 25:621–627.

266 Mizushima S, Ohshige K, Watanabe J, Kimura M, Kadowaki T, Nakamura Y, Tochikubo O,
267 Ueshima H. 2004. Randomized controlled trial of sour milk on blood pressure in borderline
268 hypertensive men. *Am J Hypertens* 17:701–706.

269 Narva M, Collin M, Jauhiainen T, Vapaatalo H, Korpela R. 2004. Effects of *Lactobacillus*

270 *helveticus* fermented milk and its bioactive peptides on bone parameters in spontaneously
271 hypertensive rats. *Milchwissenschaft* 59:359–363.

272 Oh JY, Je JG, Lee HG, Kim EA, Kang SI, Lee JS, Jeon YJ. 2020. Anti-hypertensive activity of
273 novel peptides identified from olive flounder (*Paralichthys olivaceus*) surimi. *Foods* 9: 647.

274 Pyo YH, Lee TC. 2007. The potential antioxidant capacity and angiotensin I-converting
275 enzyme inhibitory activity of monascus-fermented soybean extracts: evaluation of monascus-
276 fermented soybean extracts as multifunctional food additives. *J Food Sci* 72:S218–S223.

277 Ramchandran L, Shah NPJ. 2011. Yogurt can beneficially affect blood contributors of
278 cardiovascular health status in hypertensive rats. *J Food Sci* 76:H131-H136.

279 Rendón-Rosales MÁ, Torres-Llanez MJ, Mazorra-Manzano MA, González-Córdova AF,
280 Hernández-Mendoza A, Vallejo-Cordoba B. 2022. In vitro and in silico evaluation of
281 multifunctional properties of bioactive synthetic peptides identified in milk fermented
282 with *Lactococcus lactis* NRRL B-50571 and NRRL B-50572. *LWT* 154: 112581.

283 Tsai JS, Pan YSLS, Chen TJ. 2006. Antihypertensive peptide and γ -aminobutyric acid from
284 prozyme 6 facilitated lactic acid bacteria fermentation of soymilk. *Process Biochem* 41: 1282-
285 1288.

286 Walter SV, Hamet P. 1986. Enhanced DNA synthesis in heart and kidney of newborn
287 spontaneously hypertensive rats. *Hypertension* 8:520-525

288 Wang X, Wang L, Cheng X, Zhou J, Tang X, Mao XY. 2012. Hypertension-attenuating effect
289 of whey protein hydrolysate on spontaneously hypertensive rats. *Food Chem* 134:122-126.

290 WHO. Guideline for the Pharmacological Treatment of Hypertension in Adults; WHO:
291 Geneva, Switzerland, 2021; p. 1.

292 Xia Y, Yu J, Xu W, Shuang Q. 2020. Purification and characterization of angiotensin-I-
293 converting enzyme inhibitory peptides isolated from whey proteins of milk fermented
294 with *Lactobacillus plantarum* QS670. J Dairy Sci 103: 4919–4928.

295 Yang HYT, Erdo's EG, Levin Y. 1970. A dipeptidyl carboxypeptidase that converts angiotensin
296 I and inactivates bradykinin. Biochim Biophys Acta 214:374–376.

297 Yang HY, Erdo's EG, Levin Y. 1971. Characterization of a dipeptide hydrolase (kininase II:
298 angiotensin I converting enzyme). J Pharmacol Exp Ther 177:291–300.

299 Yim HE, Yoo KH. 2008. Renin-angiotensin system-considerations for hypertension and kidney.
300 Electrolytes Blood Press 6:42-50.

301 Yuan L, Li Y, Chen M, Xue L, Wang J, Ding Y, Zhang J, Wu S, Ye Q, Zhang S, Yang R, Zhao
302 H, Wu L, Liang T, Xie X, Wu Q. 2022. Antihypertensive Activity of Milk Fermented
303 by *Lactiplantibacillus plantarum* SR37-3 and SR61-2 in L-NAME-Induced Hypertensive Rats.
304 Foods 11:233

Table 1. Change of feed intake in SHR fed with experimental diets(g/rat) for 10 weeks

	1	2	3	4	5	6	7	8	9	10
NG	38.8±2.5 ^{NSa}	36.5±2.3 ^{NSa}	36.4±2.4 ^{NSa}	37.3±2.9 ^{NSa}	38.4±1.9 ^{NSa}	38.2±2.3 ^{NSa}	38.5±2.0 ^{Ba}	38.9±2.1 ^{NSa}	38.3±2.3 ^{NSa}	39.0±3.5 ^{NSa}
NCG	39.0±1.2 ^{abc}	38.6±1.0 ^{abc}	37.5±1.1 ^c	38.0±0.8 ^{bc}	39.6±1.3 ^{ab}	40.1±2.0 ^a	40.7±1.7 ^{ABa}	38.8±1.1 ^{abc}	40.6±1.2 ^a	39.3±0.9 ^{abc}
PCG	40.2±1.4 ^a	37.0±1.8 ^b	38.0±1.6 ^{ab}	37.7±1.0 ^{ab}	38.9±1.7 ^{ab}	38.7±1.1 ^{ab}	39.9±1.9 ^{ABab}	39.1±2.2 ^{ab}	40.1±2.2 ^a	39.0±2.1 ^{ab}
LTG	40.7±1.9 ^{ab}	38.6±2.5 ^b	37.6±2.5 ^b	38.2±1.8 ^b	38.7±1.9 ^b	40.3±2.2 ^{ab}	42.8±2.9 ^{Aa}	39.6±2.1 ^{ab}	40.8±1.9 ^{ab}	39.0±1.8 ^b
MTG	39.6±2.1 ^a	38.6±2.2 ^a	37.6±1.6 ^a	38.2±0.7 ^a	38.2±0.9 ^a	38.5±1.4 ^a	39.2±1.7 ^{Ba}	38.3±2.5 ^a	39.5±2.8 ^a	39.4±2.6 ^a
HTG	40.2±1.4 ^a	37.5±0.8 ^b	37.5±0.9 ^b	38.1±1.6 ^{ab}	38.3±1.0 ^{ab}	38.0±1.4 ^b	37.9±1.0 ^{Bb}	37.8±1.5 ^b	38.8±0.7 ^{ab}	38.1±2.3 ^{ab}

All values are the mean±SD(n=4).

^{NS} Values in the column are not significantly different (p<0.05).

^{A-B} In the same column are significantly different (p<0.05).

^{a-c} In the same row are significantly different(p<0.05).

NG, Normal group (WKY) with distilled water; NCG, Negative control group (SHR) with distilled water; PCG, Positive control group (SHR) with Enalapril; LTG, Low treatment group (SHR); MTG, Medium treatment group (SHR); HTG, High treatment group (SHR)

TC, Total cholesterol; HDL, High-density lipoprotein; LDL, Low-density lipoprotein; TG, Triglyceride

Table 2. Changes in the systolic blood pressure of rats for 10 weeks (Unit : mmHg)

	0	2	4	6	8	10
NG	138.0±16.9 ^a	143.3±12.7 ^a	144.1± 9.2 ^a	143.0±9.7 ^a	153.0±10.0 ^a	152.0±6.1 ^a
NCG	179.8±16.9 ^b	203.6±6.7 ^d	214.0±10.8 ^c	222.5±6.4 ^c	230.8±7.3 ^d	240.6±4.0 ^d
PCG	179.8±19.0 ^b	174.8±11.9 ^b	184.0±20.6 ^b	181.0±13.1 ^b	183.1±23.5 ^b	187.6±12.8 ^b
LTG	181.0± 9.0 ^b	190.1±18.2 ^c	204.0±13.5 ^c	212.3±11.1 ^c	214.6±16.0 ^{cd}	230.0±13.4 ^{cd}
MTG	183.5±17.3 ^b	193.5±7.3 ^{cd}	203.6±12.1 ^c	216.3±9.6 ^c	216.8±8.8 ^{cd}	227.0±15.5 ^{cd}
HTG	182.9±13.1 ^b	192.8±10.1 ^{cd}	211.3±8.3 ^c	212.5±11.3 ^c	209.9±13.3 ^c	222.7±10.1 ^c

All values are the mean±SD(n=8).

^{a-ci} the same column are significantly different(p<0.05).

NG, Normal group (WKY) with distilled water; NCG, Negative control group (SHR) with distilled water; PCG, Positive control group (SHR) with Enalapril; LTG, Low treatment group (SHR); MTG, Medium treatment group (SHR); HTG, High treatment group (SHR)

Table 3. Blood serum levels in rats after 10 weeks

	TC(mg/dL)	HDL(mg/dL)	LDL(mg/dL)	TG(mg/dL)
NG	144.4±4.8 ^a	101.5±7.8 ^a	22.8±2.1 ^a	54.9±12.2 ^b
NCG	90.8±5.1 ^{bc}	59.5±3.8 ^b	13.0±0.5 ^c	119.4±19.1 ^a
PCG	88.1±5.6 ^c	59.1±4.4 ^b	12.9±1.8 ^c	122.0±15.6 ^a
LTG	89.9±5.5 ^{bc}	60.1±4.2 ^b	12.9±0.6 ^c	134.1±25.1 ^a
MTG	94.1±4.3 ^b	60.9±4.1 ^b	15.0±2.7 ^b	104.8±47.3 ^a
HTG	88.1±5.7 ^c	58.4±4.7 ^b	13.4±1.3 ^{bc}	105.5±24.3 ^a

All values are the mean±SD(n=8).

^{a-c} In the same column are significantly different(p<0.05).

NG, Normal group (WKY) with distilled water; NCG, Negative control group (SHR) with distilled water; PCG, Positive control group (SHR) with Enalapril; LTG, Low treatment group (SHR); MTG, Medium treatment group (SHR); HTG, High treatment group (SHR)

TC, Total cholesterol; HDL, High-density lipoprotein; LDL, Low-density lipoprotein; TG, Triglyceride

Table 4. Organ weight in rat after 10 weeks(g/rat)

	Liver	Kidneys	Lungs	Heart	Testes
NG	11.80±1.13 ^b	2.37±0.17 ^b	1.36±0.14 ^a	1.07±0.09 ^c	3.27±0.21 ^a
NCG	13.03±0.29 ^{ab}	2.58±0.08 ^a	1.52±0.20 ^a	1.39±0.04 ^a	2.93±0.13 ^b
PCG ⁾	13.15±0.59 ^a	2.58±0.08 ^a	1.39±0.11 ^a	1.26±0.04 ^b	2.96±0.09 ^b
LTG ⁾	12.97±1.10 ^{ab}	2.59±0.15 ^a	1.43±0.08 ^a	1.40±0.08 ^a	2.90±0.15 ^b
MTG ⁾	12.34±1.02 ^{ab}	2.56±0.16 ^a	1.55±0.23 ^a	1.35±0.06 ^a	2.84±0.09 ^b
HTG ⁾	12.43±0.86 ^{ab}	2.54±0.13 ^a	1.38±0.09 ^a	1.37±0.05 ^a	2.89±0.18 ^b

All values are the mean±SD(n=8).

^{a-c} In the same column are significantly different(p<0.05).

NG, Normal group (WKY) with distilled water; NCG, Negative control group (SHR) with distilled water; PCG, Positive control group (SHR) with Enalapril; LTG, Low treatment group (SHR); MTG, Medium treatment group (SHR); HTG, High treatment group (SHR)

TC, Total cholesterol; HDL, High-density lipoprotein; LDL, Low-density lipoprotein; TG, Triglyceride

Table 5. Change of blood pressure-related biomarkers in the blood plasma of rats after 10 weeks

	Angiotensinogen(ng/mL)	Angiotensin-II(pg/mL)	Renin(pg/mL)
NG	18.3±1.7 ^a	5.8±1.5 ^a	153.6±23.7 ^a
NCG	28.1±2.3 ^c	13.4±1.0 ^c	204.8±15.5 ^d
PCG	21.6±1.9 ^{ab}	8.4±1.9 ^b	175.2±12.0 ^b
LTG	24.6±3.8 ^{bc}	10.0±0.7 ^b	194.7±18.0 ^{cd}
MTG	23.9±3.3 ^{bc}	9.1±1.7 ^b	180.4±11.5 ^{bc}
HTG	21.7±1.7 ^{ab}	8.7±1.7 ^b	177.5±11.4 ^b

All values are the mean±SD(n=8).

^{a-c} In the same column are significantly different(p<0.05).

NG, Normal group (WKY) with distilled water; NCG, Negative control group (SHR) with distilled water; PCG, Positive control group (SHR) with Enalapril; LTG, Low treatment group (SHR); MTG, Medium treatment group (SHR); HTG, High treatment group (SHR)

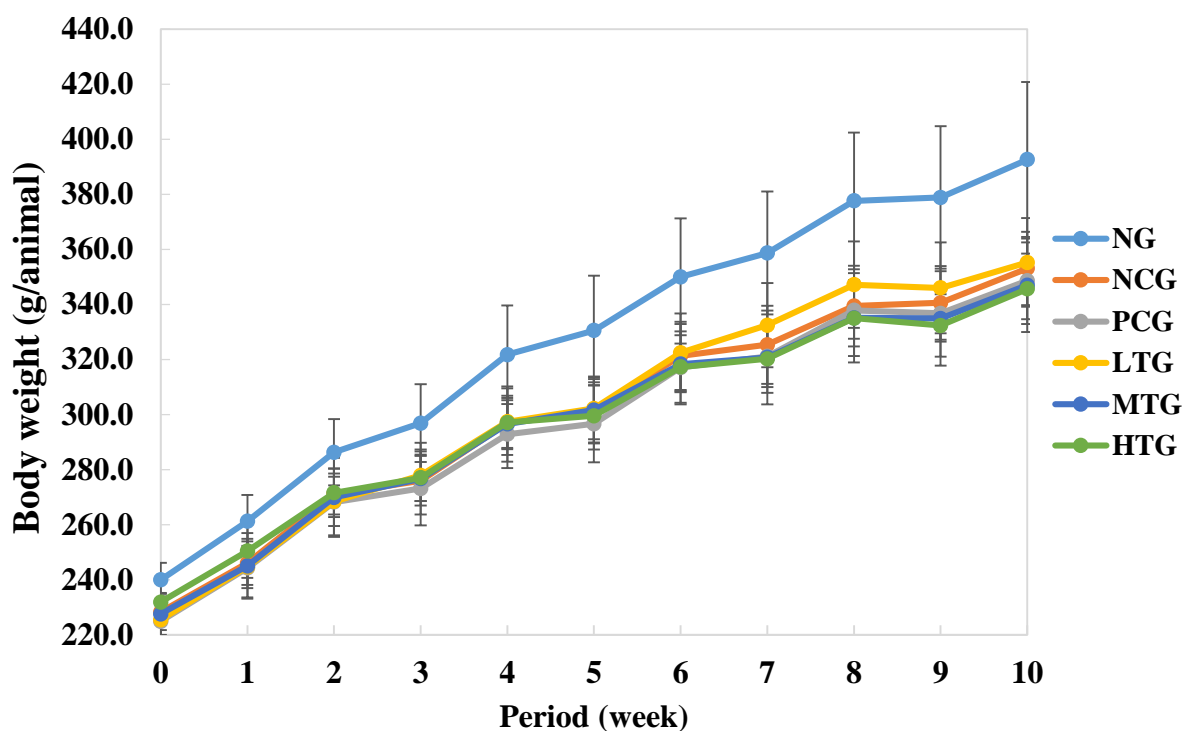


Fig. 1. Change in body weight of SHR fed with experimental diet (g/rat) for 10 weeks

All values are the mean \pm SD (n=8).

^{a-c} In the same column are significantly different(p<0.05).

NG, Normal group (WKY) with distilled water; NCG, Negative control group (SHR) with distilled water; PCG, Positive control group (SHR) with Enalapril; LTG, Low treatment group (SHR); MTG, Medium treatment group (SHR); HTG, High treatment group (SHR)

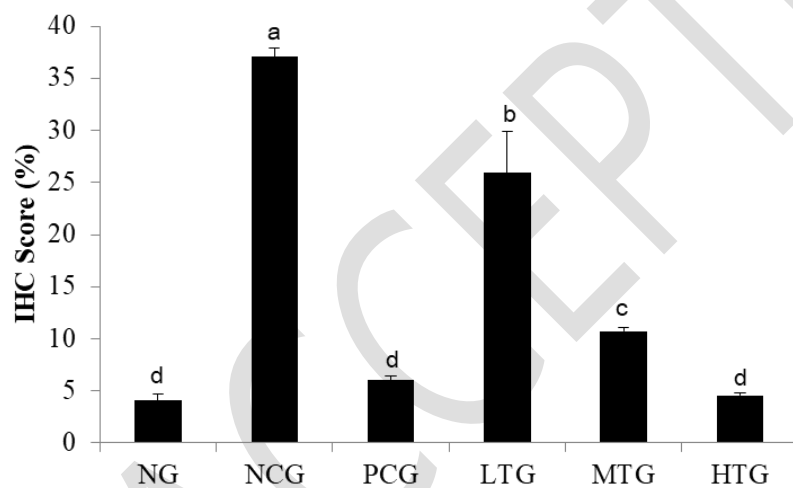
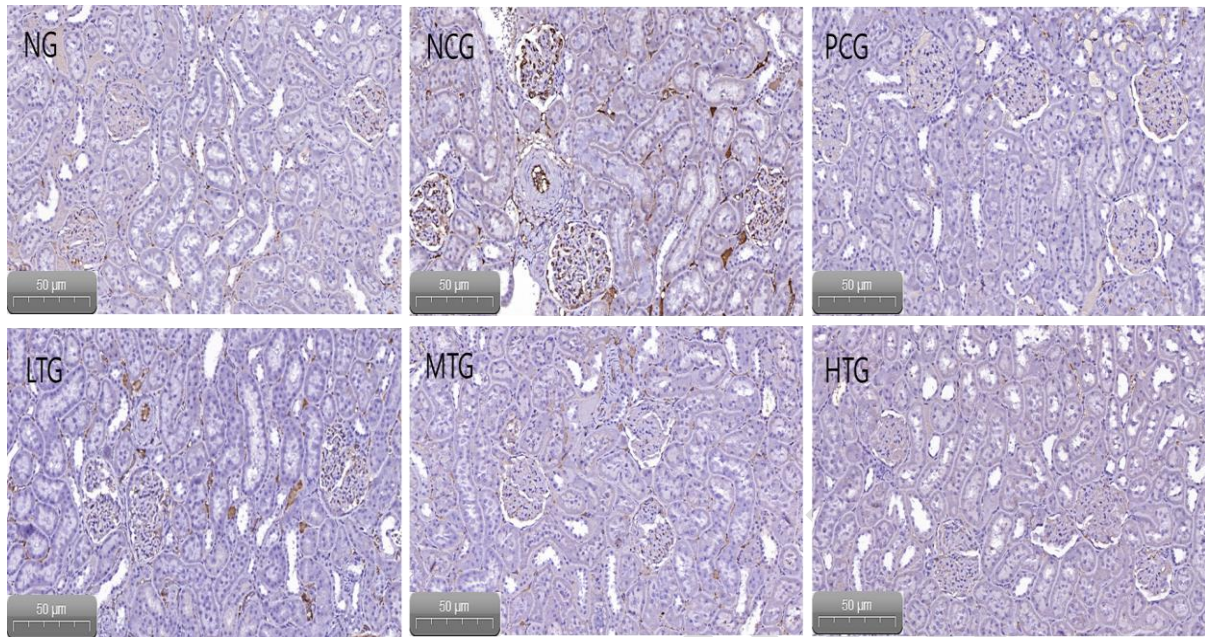


Fig. 2. Experiment to confirm the expression of angiotensin-1 protein staining in rat kidneys.

Data are expressed as mean \pm SD, ^{a-d} In the same column are significantly different ($p < 0.05$).

Tissue sections were selected for the measurement of staining areas using ImageJ. NG, Normal group (WKY) with distilled water; NCG, Negative control group (SHR) with distilled water; PCG, Positive control group (SHR) with Enalapril; LTG, Low treatment group (SHR); MTG, Medium treatment group (SHR); HTG, High treatment group (SHR)