

1  
2  
3  
4

**TITLE PAGE**  
**- Food Science of Animal Resources -**  
 Upload this completed form to website with submission

<b>ARTICLE INFORMATION</b>	<b>Fill in information in each box below</b>
<b>Article Type</b>	Research article
<b>Article Title</b>	A comparison of the physicochemical and storage characteristics of emulsified sausages made from black goat meat and conventional meats
<b>Running Title (within 10 words)</b>	Emulsion-type Sausages by Animal Species
<b>Author</b>	Da-Mi Choi, Sol-Hee Lee <sup>1,*</sup> , Hack-Youn Kim*
<b>Affiliation</b>	Department of Animal Resources Science, Kongju National University, Yesan 32439, Korean Resources Science Research, Kongju National University, Yesan 32439, Korea 1 Department of Animal Science, Chungbuk National University, Cheongju 28644, Republic of Korea
<b>Special remarks – if authors have additional information to inform the editorial office</b>	
<b>ORCID (All authors must have ORCID) <a href="https://orcid.org">https://orcid.org</a></b>	Da-Mi Choi ( <a href="https://orcid.org/0000-0002-0368-3738">https://orcid.org/0000-0002-0368-3738</a> ) Sol-Hee Lee ( <a href="https://orcid.org/0000-0003-1124-7095">https://orcid.org/0000-0003-1124-7095</a> ) Hack-Youn Kim ( <a href="https://orcid.org/0000-0001-5303-4595">https://orcid.org/0000-0001-5303-4595</a> )
<b>Conflicts of interest</b> List any present or potential conflicts of interest for all authors. (This field may be published.)	The authors declare no potential conflict of interest.
<b>Acknowledgements</b> State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available. (This field may be published.)	This research was supported by the Cooperative Research Program for Agriculture Science and Technology Development (Project No. PJ016217), Rural Development Administration, Republic of Korea.
<b>Author contributions</b> (This field may be published.)	Conceptualization: Choi DM. Data curation: Choi DM, Lee SH. Formal analysis: Choi DM, Lee SH. Methodology: Choi DM, Lee SH, Kim HY. Software: Choi DM. Validation: Choi DM, Lee SH, Kim HY. Investigation: Choi DM, Lee SH, Kim HY. Writing - original draft: Choi DM. Writing - review & editing: Choi DM, Lee SH, Kim HY.
<b>Ethics approval (IRB/IACUC)</b> (This field may be published.)	

5  
6  
7

**CORRESPONDING AUTHOR CONTACT INFORMATION**

<b>For the <u>corresponding</u> author (responsible for correspondence, proofreading, and reprints)</b>	<b>Fill in information in each box below</b>
First name, middle initial, last name	Sol-Hee Lee
Email address – this is where your proofs will be sent	leesh73@chungbuk.ac.kr
Secondary Email address	
Postal address	Chungbuk National University, Seowon-gu, Chungju-si 28644, Korea

Cell phone number	+82-10-4152-7391
Office phone number	
Fax number	

8  
9

**CORRESPONDING AUTHOR CONTACT INFORMATION**

<b>For the <u>corresponding</u> author (responsible for correspondence, proofreading, and reprints)</b>	<b>Fill in information in each box below</b>
First name, middle initial, last name	Hack-Youn, Kim
Email address – this is where your proofs will be sent	kimhy@kongju.ac.kr
Secondary Email address	
Postal address	Department of Animal Resources Science, Kongju National University, Chungnam 32439, Korea
Cell phone number	
Office phone number	+82-41-330-1241
Fax number	+82-41-330-1249

10  
11

ACCEPTED

12 **A comparison of the physicochemical and storage characteristics of**  
13 **emulsified sausages made from black goat meat and conventional**  
14 **meats**

15  
16 Abstract

17 This study evaluated the suitability of black goat meat as a raw material for meat  
18 products by comparing the physicochemical and storage characteristics of emulsified  
19 sausages from different livestock species: black goat sausage (GS), beef sausage (BS),  
20 pork sausage (PS), and chicken sausage (CS). GS and PS showed similar proximate  
21 composition, while GS and BS had comparable values for lightness, yellowness, and  
22 hue angle, indicating potential consumer appeal. Water-holding capacity (WHC) and  
23 cooking yield showed no significant differences between GS, BS, and PS, highlighting  
24 black goat's ability to retain moisture. GS and CS showed significantly higher pH value  
25 than that of the other samples ( $p < 0.05$ ). The thiobarbituric acid reactive substance  
26 (TBARS) values, indicating lipid oxidation, were significantly lower in GS and PS ( $p <$   
27  $0.05$ ), showing that GS resists oxidation well, with a strong correlation to fat content ( $R^2$   
28  $= 0.95$ ). By the 3rd and 4th weeks of storage, GS and CS had higher the volatile basic  
29 nitrogen values ( $p < 0.05$ ), correlating with pH ( $R^2 = 0.83$ ), while bacterial counts in  
30 GS, BS, and PS remained below 7 log CFU/mg for up to 5 weeks. GS's high WHC,  
31 cooking yield, and low TBARS values suggest good commercial potential.

32 Keywords: Black goat; Meat products; Livestock species; Meat raw materials;  
33 Quality characteristics

34

## 35 **Introduction**

36 Animal meat is a rich source of essential amino acids (such as histidine, lysine, leucine, and  
37 threonine, which cannot be synthesized by the human body), as well as minerals and vitamins that  
38 are important sources of energy for humans (Karwowska et al., 2021). Although, beef, pork, and  
39 chicken are the predominantly consumed animal meats, the consumption of mutton and goat meat,  
40 which were previously domesticated for milk production, has consistently increased, particularly  
41 in regions such as Asia, the Middle East, and Africa (Pandey and Upadhyay, 2022; Teixeira et al.,  
42 2020). The black goat (*Capra hircus*) is a domesticated goat belonging to the bovine family,  
43 characterized by its small size and black fur (Dong et al., 2015). Goat meat is characterized by  
44 relatively low fat, low calories, and low cholesterol compared to other meats (Lalhriatpuii and  
45 Singh, 2021). Also, goat meat is known to be a relatively acceptable meat with few cultural and  
46 religious restrictions, so it has great potential to be widely consumed in various cultures  
47 (Sujarwanta et al., 2024). In particular, goat meat is gaining attention as a suitable meat for health-  
48 conscious consumers, as it is a high-protein food that provides a rich source of essential amino  
49 acids and various micronutrients, such as iron, zinc, and vitamins (Cordeiro et al., 2022). However,  
50 goat meat has a high proportion of unsaturated fatty acids, which makes it more susceptible to  
51 spoilage when exposed to oxygen, heat, or light (Forte et al., 2024).

52 Meat products are susceptible to protein degradation and lipid oxidation caused by microbial  
53 contamination resulting from factors such as high fat and moisture content, inadequate handling  
54 during processing, and insufficient heat treatment (Boeira et al., 2020). Destruction of the  
55 secondary and tertiary structure of proteins caused by various factors such as heat, oxygen, and  
56 pH can lead to protein denaturation, thereby deteriorating food quality (Barbhuiya et al., 2021;  
57 Wang et al., 2021). Furthermore, fat generates hydroperoxides through oxygen and heat, which  
58 are then decomposed into secondary byproducts such as aldehydes and ketones, significantly  
59 reducing the shelf life of meat products (Ahmad et al., 2023). Therefore, the differential chemical  
60 composition and fatty acid profile of the species-specific meat may cause differences in the shelf  
61 life (Dave and Ghaly, 2011; KARAKÖ K et al., 2010).

62 Emulsified sausages refer to ground meat products in which the raw meat is finely mixed with  
63 salt, fat, ice water, and other ingredients, followed by emulsification and heating (Lee et al., 2020).  
64 In emulsification, salt-soluble protein; a crucial factor, forms a film on the outer surface of the  
65 emulsified fat globules, creating a gel matrix with an even dispersion of fat and myofibrillar tissue  
66 (Jung et al., 2022). Thus, the non-water-miscible solid fat gets dispersed into small globules,  
67 forming a uniform oil-in-water emulsion with the protein dissolved in the liquid phase (Zhang et  
68 al., 2022). Fat not only contributes to the flavor and texture of meat products, but also influences  
69 their taste and quality (Domínguez et al., 2019). However, uneven emulsion structures and fats  
70 are composed of unstable structures that readily react with oxygen, making them susceptible to  
71 oxidation leading to quality deterioration and reduced shelf life (Wongnen et al., 2022).

72 Recently, the food industry has been increasingly proposing challenges for the development  
73 of products and technologies aimed at increasing the production and acceptance of goat-derived  
74 products (Guerra et al., 2011). Therefore, with the anticipated increase in consumption of  
75 alternative livestock such as sheep and goat meat, as well as meat products, there is a need to  
76 develop measures that facilitate their acceptance and consumption (Mazhangara et al., 2019;  
77 Teixeira et al., 2020). Accordingly, research on various products utilizing goat meat, such as  
78 sausages (Park et al., 2020), patties (Khan et al., 2020), nuggets (Banerjee et al., 2020), fermented  
79 sausages (Ko et al., 2021), and mortadella (Guerra et al., 2011) is actively being conducted.

80 In this study, to address the challenges associated with goat meat, emulsified sausages made  
81 from goat meat, as well as commonly used meats such as beef, pork, and chicken (all using the  
82 same cuts), were compared. The purpose of this study is to demonstrate that goat meat can be  
83 used as a raw material for meat products without significant differences compared to other  
84 livestock species. To this end, we performed an analysis of the quality and storage characteristics  
85 of emulsified sausages among different livestock species.

86

87

## 88 **Materials and Methods**

### 89 **Sample preparation**

90 The black goat; *M. biceps femoris* (Gaon, Gang-jin, Korea), beef; *M. biceps femoris* (Daon,  
91 Yesan, Korea), pork; *M. biceps femoris* (I-homemeat, Seoul, Korea), and chicken drumsticks  
92 (Hamoni-mart, Yesan, Korea) used in this study were purchased 24 h after slaughter and utilized  
93 for the experiments. The raw meat (with excessive connective tissue removed) and pork back fat  
94 were ground separately using a grinder (PA-82, Mainca, Barcelona, Spain) equipped with a 3 mm  
95 plate. The sausage manufacturing process consisted of the following steps. First, after adding  
96 sugar (1%) and salt (1.2%) to the raw meat (60%), the mixture was mixed for 1 min using a bowl  
97 cutter (K-30, Talsa, Valencia, Spain) to extract salt-soluble proteins. Next, the back fat (20%) and  
98 half of the ice water (20%) were added to the mixture and mixed for 1 min. Finally, the remaining  
99 half of the ice water was added, and the mixture was mixed for 1 min to produce the final emulsion.  
100 It was ensured that the temperature of the emulsion did not exceed 10°C during all the steps. The  
101 prepared emulsion was filled into natural casings using a filling machine and then heated at 80°C  
102 for 40 min using a chamber, followed by cooling at 20°C for 30 min. The manufactured sausages  
103 were stored for 5 weeks at 4°C in vacuum packs and were used in the experiment. The sausages  
104 were cooked all at once, and the samples were separated by week. For each respective week, the  
105 samples were brought to room temperature at room temperature for 30 minutes before being used  
106 in the experiments. And a total of three batches were prepared for each species, and 700 g per  
107 batch (each 100g × 7) was produced for each species and used for the experiment.

108

### 109 **Measurement of proximate composition**

110 Association of Official Analytical Chemists (AOAC) analysis was used to measure and  
111 compare the proximate compositions of the samples (950.46, 942.05, 991.36, 920.153). The  
112 proximate composition used a sample immediately after manufacturing.

113

114

115 **Measurement of color**

116 Color was measured at the center of the sample cross-section after cooking. The lightness,  
117 redness, and yellowness were measured using a colorimeter (CR-10, Minolta, Tokyo, Japan).  
118 The colorimeter was equipped with a pulsed xenon lamp, a 2° standard observer, a light source  
119 D65, and an 8 mm aperture. The hue angle was calculated using the following equation:

120 
$$\text{Hue angle} = \tan^{-1} \times \frac{\text{CIE } b^*}{\text{CIE } a^*}$$

121

122 **Measurement of water holding capacity (WHC)**

123 Water holding capacity was measured using the method of Lee et al. (2020). Five grams of the  
124 sample wrapped in filter paper (Whatman No. 1, GE Healthcare, Chicago, IL, USA) was placed  
125 in a 50 mL conical tube and centrifuged at 4°C and 246 × g for 10 min using a centrifuge (Supra  
126 R22, Hanil Science, Gimpo, Korea). The water holding capacity was determined by measuring  
127 the weight of the sample before and after centrifugation using the following formula:

128 
$$\text{WHC} = \frac{A - B}{A} \times 100$$

129 
$$A = \frac{\text{Sample weight before centrifugation (g)} \times \text{moisture content (\%)}}{100}$$

130 
$$B = \text{Sample weight before centrifugation (g)}$$

131 
$$- \text{Sample weight after centrifugation (g)}$$

132

133 **Measurement of cooking yield**

134 The cooking yield was measured using same weight samples as much as possible to maintain  
135 uniformity in the degree of heat exposure to the samples. After cooking at 80°C for 40 min using  
136 a chamber (10.10ESI/SK, Alto Shaam, Menomonee Falls, WI, USA), it was allowed to cool at  
137 20°C for 30 min (Lee et al., 2023). The sample weight was measured before and after cooking,  
138 and the cooking yield was calculated by incorporating these measurements into the following  
139 formula:

140 
$$\text{Cooking yield (\%)} = \frac{\text{After cooking (g)}}{\text{Before cooking (g)}} \times 100$$

141

#### 142 **Measurement of pH**

143 The pH values of the cooked samples were measured using a glass electrode pH meter  
144 (Model S220; Mettler-Toledo, Schwerzenbach, Switzerland). After mixing the sample and  
145 distilled water in a ratio of 1:4, it was homogenized for 30 seconds under the condition of  
146 10,000 rpm using an ultraturrax (HMZ-20DN, Poonglim Tech, Seongnam, Korea). Before  
147 measurement, the pH meter was calibrated with buffer solutions of pH 4.01, 7.00, and 10.00,  
148 respectively (Suntex Instruments Co., Ltd., Taipei, Taiwan).

149

#### 150 **Measurement of thiobarbituric acid reactive substances (TBARS)**

151 TBARS levels were measured using the distillation method (Lee et al., 2024). First, 5 g from  
152 each cooked sample was homogenized with 50 mL of distilled water and 200  $\mu$ L of 0.3%  
153 butylated hydroxytoluene and transferred to a distillation flask. The homogenate was dis-tilled  
154 with 47.5 mL distilled water, 2.5 mL 4 N HCl, and 1 mL antifoaming agent, and 20 mL of the  
155 distillate was collected. Next, 5 mL of 0.02 M 2-thiobarbituric acid in 90% acetic acid was added  
156 to each screw cap tube containing 5 mL of the distillate and mixed. The screw cap tubes were  
157 heated at 100°C for 35 min using a water bath (JSWB-30T, JSR, Gongju, Korea) and then cooled  
158 with running water for 10 min. The absorbance was measured at 538 nm using a multimode  
159 microplate reader (SpectraMax iD3; Molecular Devices, San Jose, CA, USA). The amount of  
160 malondialdehyde (MDA) was calculated using a standard curve of 1,1,3,3-tetraethoxypropane,  
161 and the TBARS value was reported as mg MDA per kg of sample.

162

#### 163 **Measurement of volatile basic nitrogen (VBN)**

164 VBN was measured using the method of Kim and Kim (2024). The VBN content of the cooked  
165 samples was measured using the microdiffusion method. After mixing 10 g of the sample with 30  
166 mL of deionized water, the mixture was homogenized at 5,614  $\times$  g for 1 min using a homogenizer



167 (AM-5, Nihonseiki Kaisha Ltd., Tokyo, Japan). The homogenate was placed in a cylinder and up  
168 to 100 mL of distilled water was added. Thereafter, the homogenate was filtered through a filter  
169 paper (Whatman No. 1, GE Healthcare, Chicago, IL, USA). Next, 1 mL of the filtrate was put  
170 into the outer chamber of the Conway dish, and 1 mL of 0.01 N H<sub>3</sub>BO<sub>3</sub> and 100 μL of the Conway  
171 indicator were added to the inner chamber. Then, after adding 1 mL of 50% K<sub>2</sub>CO<sub>3</sub> to the outer  
172 chamber, the Vaseline-coated lid was closed and the mixture in the inner chamber was reacted at  
173 40°C for 90 min. The amount of VBN was titrated by mixing 0.02 N H<sub>2</sub>SO<sub>4</sub> until the reacted  
174 solution in the inner chamber changed from green to red. The VBN content was expressed using  
175 the following formula:

$$176 \quad \text{VBN (mg/ 100g)} = \frac{B1 - B2}{A} \times 0.14 \times a \times b \times 100$$

177 Where 'A' is the sample weight (g); 'B1' is the titration amount of the sample (mL); 'B2' is  
178 the titration of blank (mL); 'a' is the standardization index of 0.02 N sulfuric acid; and 'b' is the  
179 dilution factor.

180

### 181 **Measurement of total bacterial counts (TBC)**

182 The total number of microorganisms on the basis of the storage period was measured using  
183 the following method. After mixing 25 g of the cooked sample with 225 mL 0.1% buffered  
184 peptone water (BPW), the mixture was homogenized for 1 min using a stomacher (WH4000-  
185 2751-9, 3M; Saint Paul, MN, USA). Thereafter, 1 mL of the filtrate was collected and diluted in  
186 9 mL of 0.1% BPW and the process was repeated as many times as necessary. The diluted filtrate  
187 was plated on tryptic soy agar and cultured in an incubator (WSC-2610, ATTO, Tokyo, Japan) at  
188 37 °C for 24 h, and the number of colonies produced was measured and expressed as log colony  
189 forming unit (log CFU / g).

190

### 191 **Statistical analysis**

192 In this study, analyses of proximate composition, color, WHC, and cooking yield (4 treatments  
193 \* 5 replication), and pH, TBARS, VBN, and TBC (4 treatments \* 5 storage periods \* 5 replication)

194 were performed. One-way Analysis of Variance, following the general linear model (GLM)  
195 procedure of SAS software (Version 9.4 Windows, SAS Institute Inc., Cary, NC, USA), was  
196 conducted to perform the analysis of variance of the experimental results. Additionally, the  
197 significance of the data was analyzed using Tukey's studentized range test ( $p < 0.05$ ). The fixed  
198 effects for analysis of physicochemical and storage properties included the treatment types (Goat,  
199 Beef, Pork, and Chicken), and storage periods (0, 1, 3, 5 weeks). The data in this experiment was  
200 presented as mean  $\pm$  standard deviation. The correlation between meat, moisture, protein, fat, pH,  
201 TBARS, VBN, and TBC was analyzed using Python (google, colaboratory). This was performed  
202 using libraries such as Pandas for loading and data preparation, Matplotlib for data visualization,  
203 and Seaborn for heatmap and data summary.

204

## 205 **Results and Discussion**

### 206 **Proximate composition**

207 The proximate compositions of the emulsified sausages according to animal species are shown  
208 in Table 1 (Table 3). The GS and PS samples exhibited significantly higher moisture content than  
209 the other samples ( $p < 0.05$ ). In contrast, the BS and CS samples exhibited significantly higher  
210 fat content than that by the other samples ( $p < 0.05$ ). Beef is the meat of livestock with well-  
211 developed intramuscular fat; thus, the higher fat content in the BS samples may be due to the  
212 inherently higher fat content compared to that from the other livestock (Mohammed et al., 2020).  
213 On the other hand, in the case of goats, most of the fat is accumulated in the internal organs,  
214 resulting in relatively lowfat content in the carcass (van Wyk et al., 2022). Zhang et al. (2022)  
215 reported a negative correlation between fat and moisture content, which is consistent with the  
216 findings of this study. The protein content was significantly lower in the CS samples than that in  
217 the other samples ( $p < 0.05$ ). Chen et al. (2016) reported that the protein content in chicken  
218 drumsticks is approximately 19%, which is lower than that of black goats (approximately 21%),  
219 pork (approximately 21%), and beef (approximately 22%) (Cheng et al., 2020; Choi et al., 2023;  
220 Wójciak et al., 2021). The variation in the protein content of the sausages according to animal

221 species is presumed to be derived from differences in the proximate composition of the raw meat.  
222 The GS sample exhibited a lower crude ash content than that by the other samples. The typical  
223 ash content of goat meat ranges from approximately 0.93–1.63%, and a similar value of  
224 approximately 1.62% was observed in the present study (Lalhriatpuii and Singh, 2021). Based on  
225 the general compositional analysis, variations in the proximate composition were observed  
226 depending on the animal species, suggesting potential differences in shelf life and quality.  
227 Therefore, biochemical characteristics and storage stability analyses of emulsified sausages from  
228 different animal species were performed.

229

### 230 **Color, WHC, and cooking yield**

231 The color, WHC, and cooking yield of emulsified sausages according to animal species are  
232 shown in Table 2 (Table 3). Lightness was significantly different among the samples ( $p < 0.05$ ).  
233 The redness was significantly higher in the GS and BS samples than that in the other samples ( $p$   
234  $< 0.05$ ). This is because chicken meat contains a high level of white muscle fibers; 'type IIB',  
235 whereas goat and beef contain a significant amount of red muscle fibers (Cheng et al., 2022).  
236 Livestock species exhibit varying levels of myoglobin content in the muscles. Ruedt et al. (2023)  
237 reported myoglobin content of 2.6–2.9 mg/g for goats, 4–10 mg/g for cattle, 0.3–3 mg/g for pigs,  
238 and 0.1–0.6 mg/g for chickens depending on the species. Similarly, in this study, higher redness  
239 was observed in the order  $BS > GS > PS > CS$ , suggesting that the myoglobin content inherent to  
240 the original raw meat influenced the results. Yellowness was significantly higher in the CS sample  
241 than that in the other samples ( $p < 0.05$ ). This may be due to the feed that chickens typically  
242 consume. The color of chicken meat, particularly the yellow color, is influenced by carotenoids  
243 found in the feed (such as corn and alfalfa), and these carotenoids accumulate in the meat and fat,  
244 giving them a yellow tint (Wei et al., 2023). Generally, meat yellowness positively correlates with  
245 lightness and negatively correlates with redness (Luciano et al., 2009; Wang et al., 2021).  
246 Therefore, in this study, the high lightness and low redness of the CS samples influenced the  
247 yellowness. The hue angle value was the highest in the CS sample followed by PS, GS, and the

248 least in BS samples ( $p < 0.05$ ). The hue angle changes from  $0^\circ$  (red) to  $90^\circ$  as it approaches yellow  
249 color (Bernardez-Morales et al., 2023). Therefore, it was established that the hue angle value of  
250 CS, which exhibited higher yellowness compared to the other samples, was high, whereas the hue  
251 angle values of the GS and BS samples, characterized by higher redness, were low. Ultimately,  
252 GS showed values numerically close to those of BS in redness, yellowness, and hue angle. Goat  
253 meat provides color values within an acceptable range for meat products. However, it cannot be  
254 ruled out that the correlation between color and other factors may vary depending on the rearing  
255 environment and feed composition. Therefore, it is suggested that future analyses should be  
256 conducted under experimental conditions that eliminate the effects of feed.

257 The WHC of the emulsified sausages did not differ significantly among the GS, BS, and PS  
258 samples; however, the CS sample exhibited the lowest WHC ( $p < 0.05$ ). Xu et al. (2020) reported  
259 that as the proportion of fast-glycolytic fibers such as MyHC-IIB increases, the protein solubility  
260 decreases, leading to a decrease in the meat WHC. Therefore, the WHC of chickens, which  
261 contain a high proportion of type IIB muscle fibers, was low. The WHC measurement results  
262 showed that the GS group exhibited values similar to those of the BS and PS groups, suggesting  
263 that the use of goat meat can yield similar results in meat product manufacturing.

264 The CS sample exhibited the highest cooking yield, whereas the GS, BS, and PS samples  
265 demonstrated cooking yields similar to results of WHC measurement. The GS sample showed a  
266 high cooking yield, which was attributed to its high content of connective tissues (Bakhsh et al.,  
267 2019). Black goat meat and chicken legs contain more connective tissues than other animal  
268 species, and the connective tissues, which are composed of proteins, can interact with water to  
269 contribute to the formation of free water (Lee et al., 2021; Voytsekhivska et al., 2020; Wang et  
270 al., 2022). However, free water is very unstably bound by surface tension, so it is easily released  
271 to the surface of the meat by small impact (Geng et al., 2022). As a result, the cooking yield of  
272 the CS sample was high, while the WHC was low. GS and CS samples showed significantly  
273 higher cooking yield values than those in BS and PS samples. However, as the amount of  
274 connective tissue increases, there is a possibility that the texture of meat products may become

275 tougher or less desirable. Therefore, it is believed that the impact of a high proportion of  
276 connective tissue on consumer acceptability should also be taken into consideration.

277

## 278 **pH**

279 The pH not only affects quality characteristics such as the WHC and cooking yield of meat  
280 products, but it also has a close relationship with microbial growth indicators (Clinquart et al.,  
281 2022; Nisae et al., 2020). Low pH (acidic conditions) stabilizes the oxidation state of  $\text{Fe}^{2+}$ , thereby  
282 inhibiting the rate at which myoglobin is oxidized to metmyoglobin (Hoa et al., 2021). Conversely,  
283 in high pH (alkaline conditions), the oxidation of myoglobin is facilitated, leading to an increase  
284 in the formation of metmyoglobin, and excessive alkalinity can also decrease the structural  
285 stability of myoglobin, resulting in color changes or deterioration (Hoa et al., 2021). Figure 1  
286 shows the pH of emulsified sausages based on animal species and storage period. The pH of all  
287 samples, except for the PS sample, showed an increasing trend with increasing storage period.  
288 This may be due to the accumulation of alkaline substances produced as microorganisms grow  
289 and the microbial decomposition of amino acids into alkaline compounds like ammonia, both of  
290 which increase the pH (Anal, 2019; Zhang et al., 2023). Throughout all storage periods, the pH  
291 values were significantly higher in GS and CS samples than that in the other samples ( $p < 0.05$ ).  
292 The normal pH of raw black goat meat is 5.5–6.2 (Gawat et al., 2022), and according to Zhang et  
293 al. (2022), the pH of raw chicken leg meat is approximately 6.94. This is at a higher level  
294 compared to the normal pH range of other meats (5.4–5.7), and may have affected the final  
295 sausage pH. Therefore, it can be presumed that the pH of the meat itself can affect the quality and  
296 storage characteristics of the meat products, and thus, an analysis of the quality and storage  
297 characteristics of emulsified sausages according to the livestock species is deemed necessary.

298

## 299 **TBARS**

300 Figure 2 shows the TBARS results of the emulsified sausages according to the livestock  
301 species. Both the BS and CS samples showed significantly higher TBARS values than those of

302 the other samples at all time points ( $p < 0.05$ ). Polyunsaturated fatty acids (PUFAs) contain  
303 multiple double bonds in their structure, making them highly sensitive to oxidation, and a direct  
304 correlation between PUFA content and TBARS values has been reported (Jerónimo et al., 2020;  
305 Santos et al., 2021). Meat products exhibit varying levels of PUFAs depending on the species,  
306 with black goat containing approximately 7.52%, chicken 23.29%, beef containing 3.73%, and  
307 pork containing 9.14% PUFAs (Choi et al., 2023; Muzolf-Panek and Kaczmarek, 2021). In the  
308 case of goat and pork sausages, the proportion of PUFA is somewhat higher compared to beef;  
309 however, the total fat content in the meat itself is low, which is judged to have contributed to this  
310 result. Furthermore, the proximate composition analysis of the emulsified sausages revealed that  
311 the fat content was significantly higher in BS and CS samples than in the other samples. The  
312 correlation coefficient indicated a significant positive correlation between the fat content of the  
313 emulsified sausages and TBARS (Figure 5;  $R^2=0.95$ ). In this study, we determined that these  
314 results were due to differences in fatty acid composition and fat content between livestock species.  
315 Similarly, in a study on lipid oxidation, Pérez-Andrés et al. (2020) reported that TBARS values  
316 in beef and chicken were higher than those in sheep and pork. The TBARS values showed an  
317 increasing trend with increasing storage period for all livestock species. Lipid oxidation is a major  
318 cause of quality deterioration in meat products and requires careful control because it causes  
319 undesirable changes in odor, taste, texture, and color (Barbhuiya et al., 2021). In this study,  
320 sausages manufactured using black goat meat showed TBARS values similar to those of  
321 commonly manufactured pork sausages at the 5th week of storage.

322

### 323 **VBN**

324 The VBN values of the emulsified sausages according to the livestock species are shown in  
325 Figure 3. The initial VBN values at weeks 0 and 1 were not significantly different between the  
326 livestock species. However, in the 3rd and 4th weeks of storage, the GS and CS samples showed  
327 higher VBN levels than those in the BS and PS samples. Each livestock species has a different  
328 amino acid composition, and amino acids such as arginine and histidine are known to have a

329 significant effect on the VBN content of meat (Hwang et al., 2022). Compared to goats and  
330 chickens, cows and pigs have lower arginine contents. (Guo et al., 2019; Khalid et al., 2022;  
331 Mohammed et al., 2020; Yim et al., 2019). Additionally, a high pH promotes the growth of  
332 microorganisms, which increases protein decomposition and ultimately increases the VBN value  
333 (Kim et al., 2020). In this study, the pH of emulsified sausages showed a positive correlation with  
334 VBN (Figure 5;  $R^2=0.83$ ), and the pH measurement results of emulsified sausages showed that  
335 the GS and CS group had higher pH values than those in the other groups. Therefore, the  
336 differences in VBN values among the livestock species in this study were due to differences in  
337 pH and amino acid composition. As the storage period increased for all livestock species, the  
338 VBN values increased. This is attributed to the formation and accumulation of alkaline  
339 compounds, such as  $\text{NH}_3$  and amines, produced by proteins degraded by endogenous proteases  
340 (Song et al., 2023). The destruction of nutrients due to protein decomposition in meat products  
341 can cause loss of flavor and discoloration, which can negatively impact sensory characteristics  
342 (Pellissery et al., 2020). The results of VBN analysis showed that emphasis should be laid on the  
343 aspect of protein deterioration before commercialization of black goat meat and chicken leg meat,  
344 which exhibited higher VBN levels than the other groups.

345

#### 346 **TBC**

347 Figure 4 shows the TBC of emulsified sausages according to the livestock species. There was  
348 no significant difference in the number of colonies between livestock species until the 3rd week,  
349 but in the 5th week, the CS sample showed significantly higher TBC than in the other samples ( $p$   
350  $< 0.05$ ). In this study, the pH of the CS group was the highest at all storage periods; the high pH  
351 of meat creates a favorable environment for the growth of microorganisms, which can increase  
352 the number of microorganisms (Pellissery et al., 2020). The correlation coefficient showed that  
353 the pH of emulsified sausages was positively correlated with TBC (Figure 5;  $R^2=0.87$ ), and it was  
354 determined that the CS sample with a high pH showed the highest TBC at all storage periods.  
355 Mohammed et al. (2020) supported this finding by reporting that the microbial count

356 (*Staphylococcus aureus*, *Escherichia coli*, *Salmonella*) was higher in chicken meat than in beef,  
357 lamb, and camel meat. The TBC of all the samples tended to increase as the storage period  
358 increased, and the GS, BS, and PS samples maintained less than 7 log CFU/mg until the 5th week  
359 of storage. Microbial growth serves as a standard for determining whether meat is spoiled, and if  
360 the TBC exceeds 7 log CFU/mg, meat and meat products are considered spoiled (Hwang et al.,  
361 2020). Sausages manufactured using black goat meat showed a similar level of microbial growth  
362 as beef and pork sausages up to the 5th week of storage, suggesting that black goat meat is suitable  
363 for meat products.

364

### 365 **Conclusion**

366 In this study, the physicochemical and storage characteristics of emulsified sausages  
367 manufactured from black goat, beef, pork, and chicken leg meat were analyzed.

368 Proximate composition measurements showed that the GS and PS samples had high moisture  
369 content, and the BS and CS samples had high fat content. The highest protein content was  
370 observed in the following order: BS > PS > GS > CS. As the proximate composition was different  
371 for each livestock species, it was determined that there may be differences in storage and quality  
372 depending on the composition. The GS showed redness, yellowness, and hue angle values similar  
373 to those of the BS. In addition, black goat sausage showed a higher WHC value than the other  
374 sausages and similarly showed a high cooking yield. Black goat meat and chicken sausage had  
375 higher pH values than the other samples, and the pH of the raw meat, depending on the livestock  
376 species, affected the emulsified sausage. Beef and chicken sausages showed higher TBARS  
377 values than the other samples during all storage periods. This was influenced by fat content, which  
378 showed a high positive correlation with TBARS ( $R^2=0.95$ ). VBN and TBC analyses showed that  
379 black goat and chicken sausages showed higher protein deterioration and faster microbial growth  
380 than the other samples. These results were related to pH, which positively correlated with VBN  
381 ( $R^2=0.83$ ) and TBC ( $R^2=0.87$ ).

382 The results of this study suggest that the commercialization of black goat meat holds



383 significant potential. However, special care is needed to prevent protein deterioration, and further  
384 research should be conducted.

385 Research on the sensory characteristics of black goat sausage, particularly its flavor and aroma,  
386 plays a crucial role in product development, and higher sensory satisfaction can enhance its  
387 market acceptance. Black goat has a distinctive odor that may cause aversion in some consumers,  
388 but as studies on reducing this characteristic smell progress, consumer accessibility to black goat  
389 products is likely to expand. In fact, our previous study (Choi et al., 2024) demonstrated that  
390 various treatments aimed at odor reduction positively impacted the acceptability of black goat  
391 meat, and such research can contribute to improving consumer perception and generating positive  
392 demand in the market.

393

#### 394 **Acknowledgments**

395 This research was supported by the Cooperative Research Program for Agriculture  
396 Science and Technology Development (Project No. PJ016217), Rural Development  
397 Administration, Republic of Korea.

398

#### 399 **References**

- 400 Ahmad A, Mahmood N, Hussain M, Aiman U, Al-Mijalli SH, Raza MA, Al Jbawi E. 2023.  
401 Improvement in oxidative stability and quality characteristics of functional chicken meat  
402 product supplemented with aqueous coriander extract. *Int. J. Food Prop.* 26:855-865.
- 403 Anal AK. 2019. Quality ingredients and safety concerns for traditional fermented foods and  
404 beverages from Asia: A review. *Fermentation* 5:8.
- 405 AOAC, AOAC Official Methods of Analysis, Official Method 920.153 Ash in Meat by Ashing  
406 Method, twentieth ed., Association of Official Analytical Chemists, Rockville, Maryland,  
407 USA, 2016.

408 AOAC, AOAC Official Methods of Analysis, Official Method 942.05 Protein in Meat by  
409 Combustion Method, twentieth ed., Association of Official Analytical Chemists, Rockville,  
410 Maryland, USA, 2016.

411 AOAC, AOAC Official Methods of Analysis, Official Method 950.46 Moisture in Meat by Air  
412 Drying Method, twentieth ed., Association of Official Analytical Chemists, Rockville,  
413 Maryland, USA, 2016.

414 AOAC, AOAC Official Methods of Analysis, Official Method 991.36 Fat in Meat by Solvent  
415 Extraction Method, twentieth ed., Association of Official Analytical Chemists, Rockville,  
416 Maryland, USA, 2016.

417 Bakhsh A, Hwang YH, Joo ST. 2019. Effect of slaughter age on muscle fiber composition,  
418 intramuscular connective tissue, and tenderness of goat meat during post-mortem time. *Foods*  
419 8:571.

420 Banerjee DK, Das AK, Banerjee R, Pateiro M, Nanda PK, Gadekar YP, Biswas S, McClements  
421 DJ, Lorenzo JM. 2020. Application of enoki mushroom (*Flammulina Velutipes*) stem wastes  
422 as functional ingredients in goat meat nuggets. *Foods* 9:432.

423 Barbhuiya RI, Singha P, Singh SK. 2021. A comprehensive review on impact of non-thermal  
424 processing on the structural changes of food components. *Food Res. Int.* 149:110647.

425 Bernardez-Morales GM, Nichols BW, Douglas SL, Belk AD, Brandebourg TD, Reyes TM,  
426 Sawyer JT. 2023. Extended Storage of Beef Steaks Using Thermoforming Vacuum Packaging.  
427 *Foods* 12:2922.

428 Boeira CP, Piovesan N, Flores DCB, Soquetta MB, Lucas BN, Heck RT, Alves JDS, Campagnol  
429 PCB, Santos DD, Flores EMM, Rosa CSD, Terra NN. 2020. Phytochemical characterization  
430 and antimicrobial activity of *Cymbopogon citratus* extract for application as natural  
431 antioxidant in fresh sausage. *Food Chem.* 319:126553.

432 Chen Y, Qiao Y, Xiao Y, Chen H, Zhao L, Huang M, Zhou G. 2016. Differences in  
433 physicochemical and nutritional properties of breast and thigh meat from crossbred chickens,  
434 commercial broilers, and spent hens. *Asian-Australas. J. Anim. Sci.* 29:855.

435 Cheng H, Song S, Jung EY, Jeong JY, Joo ST, Kim GD. 2020. Comparison of beef quality  
436 influenced by freeze-thawing among different beef cuts having different muscle fiber  
437 characteristics. *Meat Sci.* 169:108206.

438 Cheng H, Song S, Park TS, Kim GD. 2023. Proteolysis and changes in meat quality of chicken  
439 pectoralis major and iliotibialis muscles in relation to muscle fiber type distribution. *Poult. Sci.*  
440 101:102185.

441 Choi DM, Kang KM, Kang SM, Kim HY. 2023. Physicochemical Properties of Black Korean  
442 Goat Meat with Various Slaughter Ages. *Animals* 13:692.

443 Choi DM, Kim HY, Lee SH. 2024. Study on Ways to Improve the Quality of Black Goat Meat  
444 Jerky and Reduce Goaty Flavor through Various Spices. *Food Sci. Anim. Resour.* 44: 635.

445 Clinquart A, Ellies-Oury MP, Hocquette JF, Guillier L, Santé-Lhoutellier V, Prache S. 2022. On-  
446 farm and processing factors affecting bovine carcass and meat quality. *Animal* 16:100426.

447 Cordeiro ARRDA, Bezerra TKA, Madruga MS. 2022. Valuation of goat and sheep by-products:  
448 Challenges and opportunities for their use. *Animals* 12:3277.

449 Dave D, Ghaly AE. 2011. Meat spoilage mechanisms and preservation techniques: a critical  
450 review. *Am. J. Agric. Biol. Sci.* 6:486-510.

451 Domínguez R, Pateiro M, Gagaoua M, Barba FJ, Zhang W, Lorenzo JM. 2019. A comprehensive  
452 review on lipid oxidation in meat and meat products. *Antioxidants* 8:429.

453 Dong Y, Zhang X, Xie M, Arefnezhad B, Wang Z, Wang W, Feng S, Huang G, Guan R, Shen W,  
454 Bunch R, McCulloch R, Li Q, Li B, Zhang G, Xu X, Kijas JW, Salekdeh GH, Wang W, Jiang  
455 Y. 2015. Reference genome of wild goat (*capra aegagrus*) and sequencing of goat breeds  
456 provide insight into genic basis of goat domestication. *BMC genomics* 16:1-11.

457 Forte L, De Palo P, Natrella G, Aloia A, Maggiolino A. 2024. Effects of dry and wet ageing on  
458 culled goat meat quality. *Ital J Anim Sci.* 23:693-706.

459 Gawat M, Kaur L, Singh J, Boland M. 2022. Physicochemical and quality characteristics of New  
460 Zealand goat meat and its ultrastructural features. *Food Res. Int.* 161:111736.

461 Geng M, Wang Z, Qin L, Taha A, Du L, Xu X, Pan S, Hu H, Hu H. 2022. Effect of ultrasound  
462 and coagulant types on properties of  $\beta$ -carotene bulk emulsion gels stabilized by soy protein.  
463 Food Hydrocoll. 123:107146.

464 Guerra ICD, Félex SSS, Meireles BRLM, Dalmás PS, Moreira RT, Honório VG, Morgano MA,  
465 Milani RF, Benevides SD, Queiroga RCRE, Madruga MS. 2011. Evaluation of goat  
466 mortadella prepared with different levels of fat and goat meat from discarded animals. Small  
467 Rumin. Res. 98:59-63.

468 Guo Q, Kong X, Hu C, Zhou B, Wang C, Shen QW. 2019, Fatty acid content, flavor compounds,  
469 and sensory quality of pork loin as affected by dietary supplementation with L-arginine and  
470 glutamic acid. J. food Sci. 84:3445-3453.

471 Hoa VB, Cho SH, Seong PN, Kang SM, Kim YS, Moon SS, Choi YM, Kim JH, Seol KH. 2021.  
472 The significant influences of pH, temperature and fatty acids on meat myoglobin oxidation: a  
473 model study. J. Food Sci. Technol. 58:3972-3980.

474 Hwang SH, Lee J, Nam TG, Koo M, Cho YS. 2022. Changes in physicochemical properties and  
475 bacterial communities in aged Korean native cattle beef during cold storage. Food Sci. Nutr.  
476 10:2590-2600.

477 Hwang SI, Hong GP. 2020. Effects of high pressure in combination with the type of aging on  
478 the eating quality and biochemical changes in pork loin. Meat Sci. 162:108028.

479 Jerónimo E, Soldado D, Sengo S, Francisco A, Fernandes F, Portugal AP, Alves SP, Santos-Silva  
480 J, Bessa RJ. 2020. Increasing the  $\alpha$ -tocopherol content and lipid oxidative stability of meat  
481 through dietary *Cistus ladanifer* L. in lamb fed increasing levels of polyunsaturated fatty acid  
482 rich vegetable oils. Meat Sci. 164:108092.

483 Jung DY, Lee HJ, Shin DJ, Kim CH, Jo C. 2022. Mechanism of improving emulsion stability of  
484 emulsion-type sausage with oyster mushroom (*Pleurotus ostreatus*) powder as a phosphate  
485 replacement. Meat Sci. 194:108993.

486 KARAKÖK SG, Ozogul Y, Saler, M, Ozogul, F. 2010. Proximate analysis. Fatty acid profiles  
487 and mineral contents of meats: A comparative study. J. Muscle Foods 21:210-223.

488 Karwowska M, Stadnik J, Stasiak DM, Wójciak K, Lorenzo JM. 2021. Strategies to improve the  
489 nutritional value of meat products: Incorporation of bioactive compounds, reduction or  
490 elimination of harmful components and alternative technologies. *Int. J. Food Sci. Technol.*  
491 56:6142-6156.

492 Khalid W, Arshad MS, Nayik GA, Alfarraj S, Ansari MJ, Guiné RP. 2022. Impact of Gamma  
493 Irradiation and Kale Leaf Powder on Amino Acid and Fatty Acid Profiles of Chicken Meat  
494 under Different Storage Intervals. *Molecules* 27:8201.

495 Khan IA, Xu W, Wang D, Yun A, Khan A, Zongshuai Z, Ijaz MU, Yiqun C, Hussain, M, Huang  
496 M. 2020. Antioxidant potential of chrysanthemum morifolium flower extract on lipid and  
497 protein oxidation in goat meat patties during refrigerated storage. *J. food Sci.* 85:618-627.

498 Kim HJ, Kim HJ, Jeon J, Nam KC, Shim KS, Jung JH, Kim KS, Choi Y, Kim, SH, Jang A. 2020.  
499 Comparison of the quality characteristics of chicken breast meat from conventional and animal  
500 welfare farms under refrigerated storage. *Poult. Sci.* 99:1788-1796.

501 Kim SG, Kim HY. 2024. Proteolysis of dry-cured ham according to the type and temperature of  
502 starter mixed with *Debaryomyces hansenii* and *Penicillium nalgiovense*. *Food Biosci.*  
503 58:103787.

504 Ko JH, Seo JK, Lee JG, Yang HS. 2021. Physicochemical properties, volatile compounds and  
505 sensory attributes of dry-fermented Sausage manufactured with goat meat. *J. Agric. Life Sci.*  
506 55:97-107.

507 Lalhriatpuii M, Singh AK. 2021. Goat meat: No less source of protein in comparison to other  
508 meat for human consumption. *Goat Sci. Environ., Health Econ. IntechOpen*, 1-33.

509 Lee JA, Kang KM, Kim HY. 2024. Changes in Physicochemical Characteristics of Goat Meat  
510 Emulsion-type Sausage According to the Ratio of Fat and Water Contents. *Food Sci. Anim.*  
511 *Resour.*

512 Lee SH, Choe J, Kim JC, Kim HY. 2020. Effect of seawater on the technological properties of  
513 chicken emulsion sausage in a model system. *Food Sci. Anim. Resour.* 40:377.

514 Lee SH, Joe SD, Kim GW, Kim HY. 2020. Physicochemical properties of sausage manufactured  
515 with carp (*Carassius carassius*) muscle and pork. Food Sci. Anim. Resour. 62:903.

516 Lee SH, Kim HY. 2021. Comparison of quality and sensory characteristics of spent hen and  
517 broiler in South Korea. Animals 11:2565.

518 Lee SH, Kim HY. 2023. Analysis of physicochemical properties of dry-cured beef made from  
519 Hanwoo and Holstein meat distributed in South Korea. Heliyon 9.

520 Luciano G, Monahan FJ, Vasta V, Pennisi P, Bella M, Priolo A. 2009. Lipid and colour stability  
521 of meat from lambs fed fresh herbage or concentrate. Meat Sci. 82:193-199.

522 Mazhangara IR, Chivandi E, Mupangwa JF, Muchenje V. 2019. The potential of goat meat in the  
523 red meat industry. Sustainability 11:3671.

524 Mohammed HHH, Jin G, Ma M, Khalifa I, Shukat R, Elkhedir AE, Zeng Q, Noman AE. 2020.  
525 Comparative characterization of proximate nutritional compositions, microbial quality and  
526 safety of camel meat in relation to mutton, beef, and chicken. LWT 118:108714.

527 Muzolf-Panek M, Kaczmarek A. 2021. Chemometric analysis of fatty acid composition of raw  
528 chicken, beef, and pork meat with plant extract addition during refrigerated storage. Molecules  
529 26:4952.

530 Nisar MF, Arshad MS, Yasin M, Khan MK, Afzaal M, Sattar S, Suleria HAR. 2020. Evaluation  
531 of gamma irradiation and moringa leaf powder on quality characteristics of meat balls under  
532 different packaging materials. J. Food Process. Preserv. 44:e14748.

533 Pandey HO, Upadhyay D. 2022. Global livestock production systems: Classification, status, and  
534 future trends. Emerging Issues in Climate Smart Livestock Production 47-70.

535 Park JY, Lee SY, Choi YS, Nam KC. 2020. Quality Characteristics of Low-fat Black Goat  
536 Sausage Using Loquat Leaf. J. Agric. Life Sci. 54:59–65.

537 Pellissery AJ, Vinayamohan PG, Amalaradjou MAR, Venkitanarayanan K. 2020. Spoilage  
538 bacteria and meat quality. In Meat quality analysis, Academic Press, 307-334.

539 Pérez-Andrés JM, Crobotova J, Harrison SM, Brunton NP, Cullen PJ, Rustad T, Tiwari BK. 2020.  
540 Effect of cold plasma on meat cholesterol and lipid oxidation. Foods 9:1786.

541 Ruedt C, Gibis M, Weiss J. 2023. Meat color and iridescence: Origin, analysis, and approaches  
542 to modulation. *Compr. Rev. Food Sci. Food Saf.* 22:3366-3394.

543 Santos MD, Matos G, Casal S, Delgadillo I, Saraiva JA. 2021. Quality evolution of raw meat  
544 under hyperbaric storage—Fatty acids, volatile organic compounds and lipid oxidation profiles.  
545 *Food Biosci.* 42:101108.

546 Song S, Cheng H, Park J, Kim GD. 2023. Relationship between peptides and the change in quality  
547 characteristics of beef strip loin (*M. longissimus lumborum*) and tenderloin (*M. psoas major*).  
548 *Food Chem.* 137036.

549 Sujarwanta RO, Afidah U, Suryanto E, Rusman, Triyannanto E, Hoffman LC. 2024. Goat and  
550 Sheep Meat Production in Indonesia. *Sustainability* 16:4448.

551 Teixeira A, Silva S, Guedes C, Rodrigues S. 2020. Sheep and goat meat processed products  
552 quality: A review. *Foods*, 9:960.

553 van Wyk GL, Hoffman LC, Strydom PE, Frylinck L. 2022. Differences in meat quality of six  
554 muscles obtained from southern african large-frame indigenous veld goat and boer goat  
555 wethers and bucks. *Animals* 12:382.

556 Voytsekhivska L, Franko O, Okhrimenko Y, Verbytskyi S, Gavrilenko A. 2020. Quality  
557 parameters of collagen containing raw materials and courses of their use. *J. Eng. Sci.* 4:183-  
558 190.

559 Wang J, Yang P, Han D, Huang F, Li X, Song Y, Wang H, Liu J, Zheng J, Zhang C. 2022. Role  
560 of Intramuscular Connective Tissue in Water Holding Capacity of Porcine Muscles. *Foods*  
561 11:3835.

562 Wang K, Peng X, Lv F, Zheng M, Long D, Mao H, Si H, Zhang P. 2021. Microbiome-metabolites  
563 analysis reveals unhealthy alterations in the gut microbiota but improved meat quality with a  
564 high-rice diet challenge in a small ruminant model. *Animals* 11:2306.

565 Wang Z, Tu J, Zhou H, Lu A, Xu B. 2021. A comprehensive insight into the effects of microbial  
566 spoilage, myoglobin autoxidation, lipid oxidation, and protein oxidation on the discoloration  
567 of rabbit meat during retail display. *Meat Sci.* 172:108359.

568 Wei Y, Qin K, Qin X, Song F, Xu X. 2023. Effects of different types of xanthophyll extracted  
569 from marigold on pigmentation of yellow-feathered chickens. *Anim. Biosci.* 36:1853.

570 Wójciak KM, Halagarda M, Rohn S, Kęska P, Latoch A, Stadnik J. 2021. Selected nutrients  
571 determining the quality of different cuts of organic and conventional pork. *Eur. Food Res.*  
572 *Technol.* 247:1389-1400.

573 Wongnen C, Ruzzama N, Chaijan, M, Cheong LZ, Panpipat W. 2022. *Glochidion wallichianum*  
574 leaf extract as a natural antioxidant in sausage model system. *Foods* 11:1547.

575 Xu D, Wang Y, Jiao N, Qiu K, Zhang X, Wang L, Wang L, Yin J. 2020. The coordination of  
576 dietary valine and isoleucine on water holding capacity, pH value and protein solubility of  
577 fresh meat in finishing pigs. *Meat Sci.* 163:108074.

578 Yim DG, Choi YS, Nam KC. 2019. Sea tangle (*Laminaria japonica*) supplementation on meat  
579 quality of Korean native black goat. *J. Anim. Sci. Technol.* 61:352.

580 Zhang B, Liu Y, Peng H, Lin Y, Cai K. 2023. Effects of ginger essential oil on physicochemical  
581 and structural properties of agar sodium alginate bilayer film and its application to beef  
582 refrigeration. *Meat Sci.* 198:109051.

583 Zhang L, Chen Q, Liu Q, Xia X, Wang Y, Kong B. 2022. Effect of different types of smoking  
584 materials on the flavor, heterocyclic aromatic amines, and sensory property of smoked chicken  
585 drumsticks. *Food Chem.* 367:130680.

586 Zhang Y, Wang X, Chen H, Ren F, Liu Z, Wang P, Liu X. 2022. Application of gel-in-oil-in-  
587 water double emulsions as a pork oil replacer in emulsified sausage. *J. Food Process. Preserv.*  
588 46:e16333.

589



590 Table 1. Proximate composition of emulsion-type sausage (immediately after cooking) with  
591 various livestock type.

Trait (%)	GS	BS	PS	CS
Moisture	61.61±0.13 <sup>a</sup>	54.42±0.40 <sup>b</sup>	61.69±0.31 <sup>a</sup>	55.97±1.31 <sup>b</sup>
Crude protein	13.77±0.23 <sup>c</sup>	17.39±0.17 <sup>a</sup>	16.10±0.17 <sup>b</sup>	11.84±0.90 <sup>d</sup>
Crude fat	20.46±0.23 <sup>b</sup>	24.69±0.49 <sup>a</sup>	20.20±0.80 <sup>b</sup>	25.74±0.17 <sup>a</sup>
Crude ash	1.62±0.02 <sup>c</sup>	1.67±0.02 <sup>bc</sup>	1.71±0.02 <sup>b</sup>	1.90±0.03 <sup>a</sup>

592 <sup>a-c</sup>Means lacking a common superscript are significantly different (p < 0.05).

593 GS, goat sausage; BS, beef sausage; PS, pork sausage; CS, chicken sausage.

ACCEPTED

594 Table 2. Color, water holding capacity, and cooking yield of emulsion-type sausage (immediately  
 595 after cooking) according to various livestock type and storage periods.

Trait	GS	BS	PS	CS
Lightness	61.70±0.10 <sup>c</sup>	57.83±0.06 <sup>d</sup>	74.33±0.06 <sup>b</sup>	77.13±0.12 <sup>a</sup>
Redness	7.07±0.15 <sup>a</sup>	7.27±0.06 <sup>a</sup>	4.23±0.06 <sup>b</sup>	3.43±0.06 <sup>c</sup>
Yellowness	15.47±0.23 <sup>c</sup>	15.03±0.06 <sup>d</sup>	16.40±0.10 <sup>b</sup>	18.27±0.06 <sup>a</sup>
Hue angle	65.44±0.62 <sup>c</sup>	64.20±0.15 <sup>d</sup>	75.53±0.21 <sup>b</sup>	79.36±0.16 <sup>a</sup>
WHC (%)	94.62±0.96 <sup>a</sup>	93.52±0.37 <sup>a</sup>	94.03±0.91 <sup>a</sup>	90.04±1.61 <sup>b</sup>
Cooking yield (%)	82.61±0.81 <sup>b</sup>	68.87±0.20 <sup>d</sup>	77.20±0.17 <sup>c</sup>	92.38±0.48 <sup>a</sup>

596 <sup>a-d</sup>Means lacking a common superscript are significantly different ( $p < 0.05$ ).  
 597 GS, goat sausage; BS, beef sausage; PS, pork sausage; CS, chicken sausage.

598 Table 3. Root mean square error (RMSE) for the proximate composition, color, WHC, and  
 599 cooking yield of goat sausage, beef sausage, pork sausage, and chicken sausage.

Trait	RMSE			
	GS	BS	PS	CS
Moisture	7.85	7.38	7.85	7.48
Crude protein	3.71	4.17	4.01	3.44
Crude fat	4.52	4.97	4.49	5.07
Crude ash	1.27	1.29	1.31	1.38
Lightness	7.85	7.60	8.62	8.78
Redness	2.66	2.70	2.06	1.85
Yellowness	3.93	3.88	4.05	4.27
Hue angle	8.09	8.01	8.69	8.91
WHC (%)	9.73	9.67	9.70	9.49
Cooking yield (%)	9.09	8.30	8.79	9.61

600 GS, goat sausage; BS, beef sausage; PS, pork sausage; CS, chicken sausage.

601

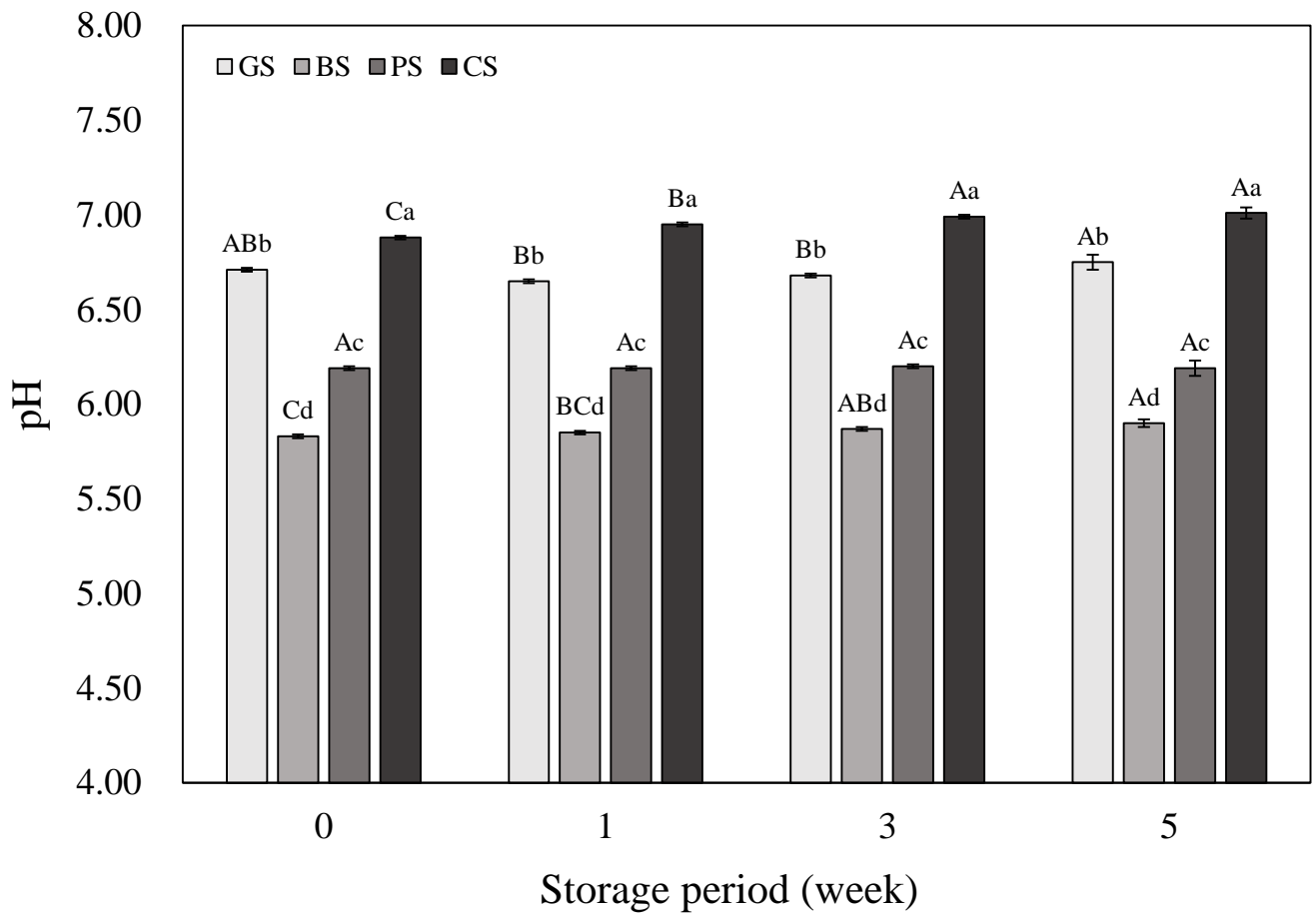


Fig. 1. pH of emulsion-type sausage according to various livestock type and storage periods. GS: goat sausage; BS: beef sausage; PS: pork sausage; CS: chicken sausage <sup>a-d</sup> Means lacking a common superscript are significantly different ( $p < 0.05$ ). <sup>A-C</sup> Means lacking a common superscript are significantly different ( $p < 0.05$ ).

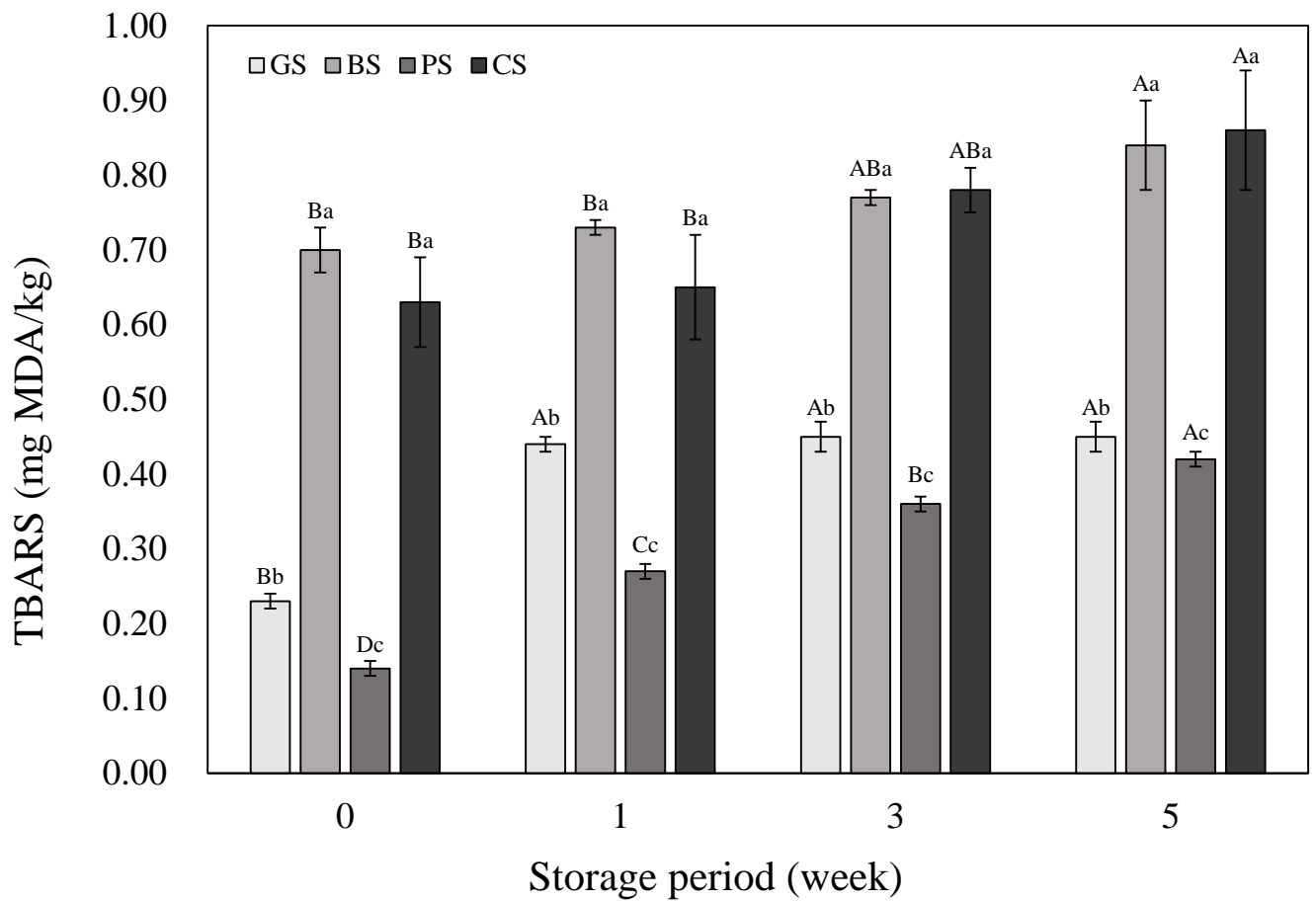


Fig. 2. Thiobarbituric acid reactive substances of emulsion-type sausage according to various livestock type and storage periods. GS: goat sausage; BS: beef sausage; PS: pork sausage; CS: chicken sausage. <sup>a-c</sup> Means lacking a common superscript are significantly different ( $p < 0.05$ ). <sup>A-D</sup> Means lacking a common superscript are significantly different ( $p < 0.05$ ).

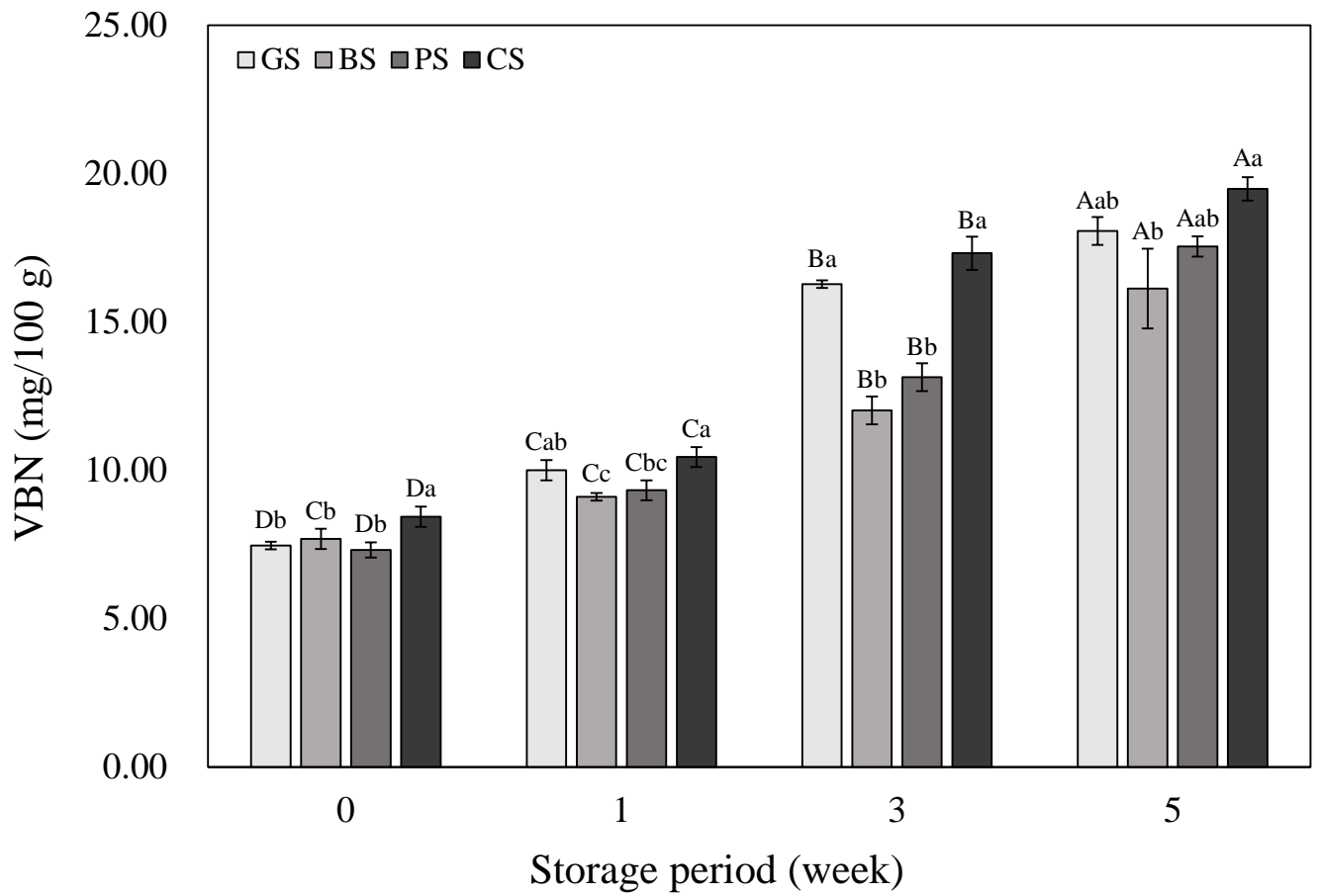


Fig. 3. Volatile basic nitrogen of emulsion-type sausage according to various livestock type and storage periods. GS: goat sausage; BS: beef sausage; PS: pork sausage; CS: chicken sausage. <sup>a-c</sup> Means lacking a common superscript are significantly different ( $p < 0.05$ ). <sup>A-D</sup> Means lacking a common superscript are significantly different ( $p < 0.05$ ).

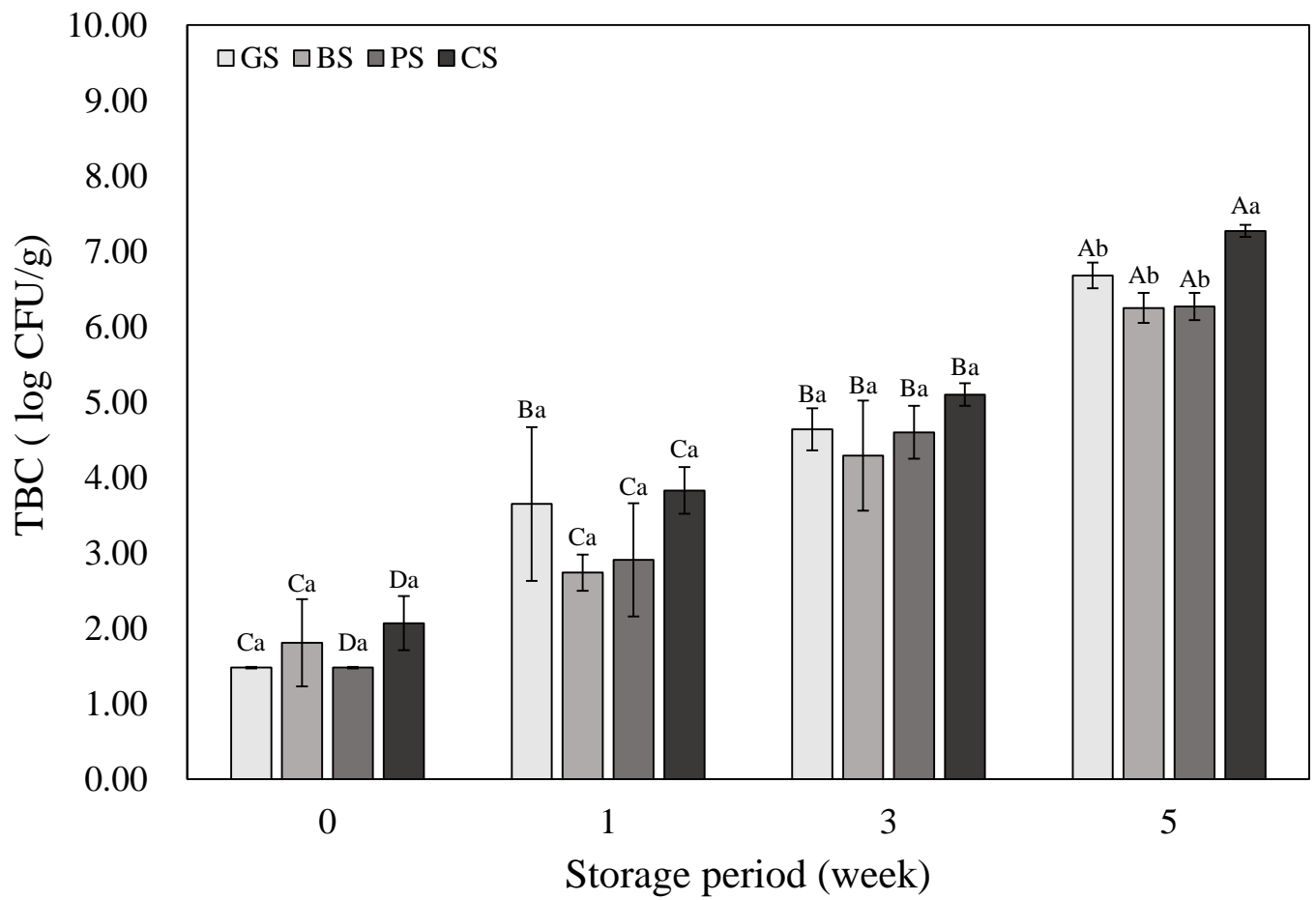


Fig. 4. Total bacterial counts of emulsion-type sausage according to various livestock type and storage periods. GS: goat sausage; BS: beef sausage; PS: pork sausage; CS: chicken sausage. <sup>a-b</sup> Means lacking a common superscript are significantly different ( $p < 0.05$ ). <sup>A-C</sup> Means lacking a common superscript are significantly different ( $p < 0.05$ ).

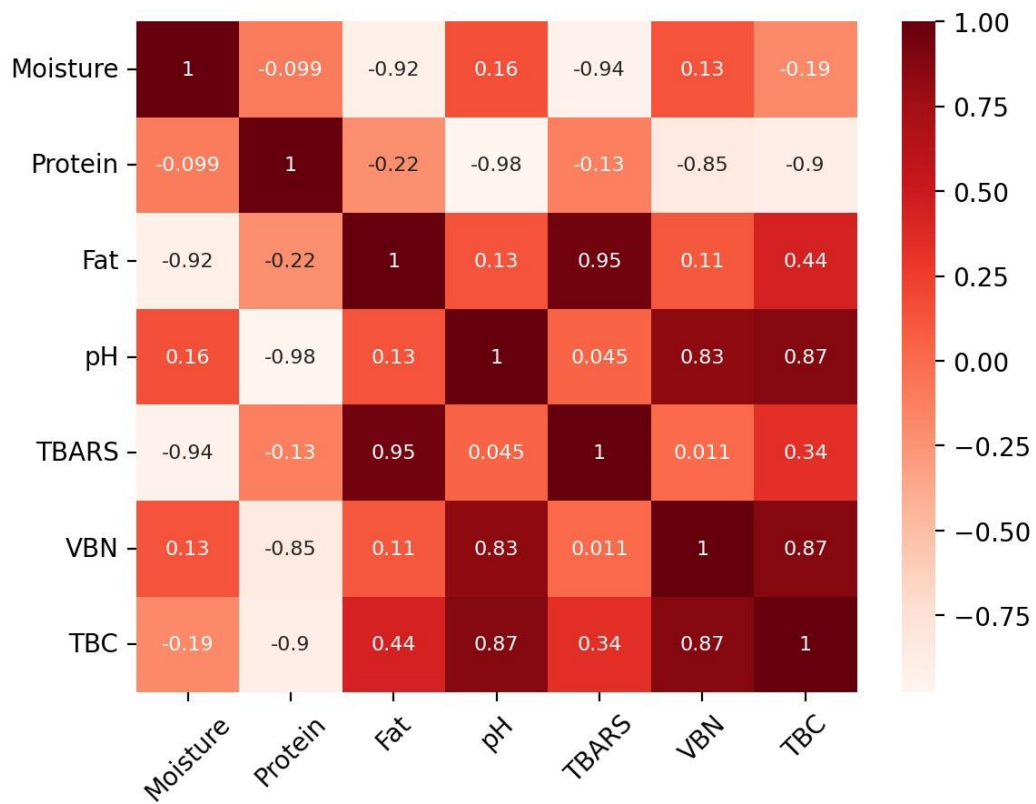


Fig. 5. Heatmap of the correlation analysis between livestock types and storage-related parameters.