# **TITLE PAGE**

# - Food Science of Animal Resources - Upload this completed form to website with submission

ARTICLE INFORMATION	Fill in information in each box below				
Article Type	Research article				
Article Title	Effects of Temperature on the Microbial Growth and Quality of Unsealed Dry Pet Food during Storage				
Running Title (within 10 words)	Storage Temperature Effects on Dry Pet Food				
Author	Dongbin Park <sup>1</sup> , Hyun Jung Lee <sup>1</sup> , Anand Kumar Sethukali <sup>1,2</sup> , Dong-Gyun Yim <sup>3</sup> , Sungkwon Park <sup>4</sup> , and Cheorun Jo <sup>1,5</sup> *				
Affiliation	<ul> <li>Department of Agricultural Biotechnology, Center for Food and Bioconvergence, and Research Institute of Agriculture and Life Science, Seoul National University, Seoul 08826, Korea</li> <li>Department of Animal Science, Faculty of Agriculture, University of Jaffna, Kilinochchi 44000, Sri Lanka</li> <li>Department of Animal Science, Kyungpook National University, Sangju 37224, Korea</li> <li>Department of Food Science and Biotechnology, College of Life Science, Sejong University, Seoul 05006, Korea</li> <li>Institute of Green Bio Science and Technology, Seoul National University, Pyeongchang 25354, Korea</li> </ul>				
Special remarks – if authors have additional information to inform the editorial office					
ORCID (All authors must have ORCID) https://orcid.org	Dongbin Park (https://orcid.org/0000-0003-4979-6049) Hyun Jung Lee (https://orcid.org/0000-0002-6891-8008) Anand Kumar Sethukali (https://orcid.org/0000-0003-0817-6396) Dong-Gyun Yim (https://orcid.org/0000-0003-0368-2847) Sungkwon Park (https://orcid.org/0000-0002-7684-9719) Cheorun Jo (https://orcid.org/0000-0003-2109-3798)				
Conflicts of interest List any present or potential conflict s of interest for all authors. (This field may be published.)	The authors declare no potential conflict of interest.				
Acknowledgements State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available. (This field may be published.)	This study was supported by the "Cooperative Research Program for Agriculture Science and Technology Development (Project No. PJ016891)" Rural Development Administration, Republic of Korea.				
Author contributions (This field may be published.)	Conceptualization: Yim DG, Park S, Jo C. Data curation: Park D, Lee HJ. Formal analysis: Park D, Anand Kumar S. Validation: Park D, Lee HJ. Investigation: Yim DG, Park S. Writing - original draft: Park D, Lee HJ. Writing - review & editing: Park D, Lee HJ, Anand Kumar S, Yim DG, Park S, Jo C.				
Ethics approval (IRB/IACUC) (This field may be published.)	This article does not require IRB/IACUC approval because there are no human and animal participants.				

## **CORRESPONDING AUTHOR CONTACT INFORMATION**

For the <u>corresponding</u> author (responsible for correspondence, proofreading, and reprints)	Fill in information in each box below
First name, middle initial, last name	Cheorun Jo

Email address – this is where your proofs will be sent	cheorun@snu.ac.kr		
Secondary Email address			
Postal address	Department of Agricultural Biotechnology, Seoul National University, Seoul 08826, Republic of Korea		
Cell phone number	+82-10-3727-6923		
Office phone number	+82-2-880-4804		
Fax number	+82-2-880-2271		



**ABSTRACT** 

Dry pet food is a convenient option for pet owners, but its storage conditions can impact its

microbial growth and quality. In this study, we examined the effects of storage temperature on the

quality and microbial growth of dry pet food. The pet food was stored at 25°C (D25) and 35°C

(D35), respectively, and samples were analyzed on days 0, 30, 90, and 120 for microbial growth

and quality attributes such as pH, color, lipid oxidation, and volatile basic nitrogen. While no initial

microbial growth was observed due to the low water content and water activity, quality attributes

showed changes over time. However, the changes were negligible, indicating that dry pet food

remains feedable for up to 120 days even at a storage temperature of 35°C. These findings suggest

that dry pet food can be safely stored at room temperature or up to 35°C without compromising its

quality.

Keywords: Dry pet food, Storage temperature, Microbial growth, Quality

## Introduction

In recent years, there is an increasing interest in pet food with the growing awareness of the pet industry (Leiva et al., 2019; Prata, 2022). There are several types of pet food, including dry pet food, semi-moist pet food, wet pet food, etc. (Carrión, 2023). Among them, dry pet food is the most convenient type for pet owners because its low moisture content prevents mold or bacteria growth, facilitating storage (Morelli et al., 2021). In addition, dry pet food has the advantage of being inexpensive because it is easy to mass-produce.

Generally, dry pet food is stored at room temperature, and it is recommended to be stored in a dry and cool place. However, buying dry pet food in bulk can lead to longer consumption periods, exposing it to various factors that may affect its quality. Once the package is opened, it cannot remain fresh due to exposure to air. For example, the humid and hot summer can make the food damp and moldy. According to a survey by Morelli et al. (2021), 23.6% of dry pet food consumers responded that stored feed could be unintentionally exposed to high temperatures. High-temperature storage promotes the oxidation of fat, which can lead to the formation of rancid flavor in dry pet food (Robert et al., 1980; Onilude et al., 2010).

Dry pet food typically contains grains such as corn, soy, oats, and barley, which have been exposed to soil microbes during growth and harvest (Scudamore and Livesey, 1998). The manufacturing process can kill microbes through high-temperature pellet extrusion (Gazzotti et al., 2015). However, post-manufacture packaging and distribution can allow for the multiplication of residual microbes (Scudamore and Livesey, 1998). It is leading to spoilage under proper conditions. Warm conditions, like in summer, accelerate microbial growth.

Consuming contaminated feed can risk food poisoning to pets. Cases of acute or chronic aflatoxicosis caused by contaminated feed have been consistently reported to harm the health of pets (Crump et al., 2002).

The recommended storage condition of dry pet food is only stated as a cool, dry, well-ventilated place away from direct sunlight. General room temperature is 25°C, however, in Korea, the year consists of four different seasons (spring, summer, autumn, and winter) with a large variation in temperature and humidity between them. And especially in summer, it gets quite humid and hot. The temperature often rises above 35°C. Such differences can vary the quality of food stored at room temperature, like dry pet food, so its changes should be studied carefully considering the actual conditions in Korea. In addition, most of the dry pet food is stored opened at home. Therefore, in order to determine the safety and stability of the opened feed, it is necessary to check the changes in microorganisms and the quality of the feed according to the storage conditions. For these reasons, this study attempted to study changes over time in microorganisms and quality while exposing the dry pet food to the air at different storage temperatures.

## Materials and methods

## Sample preparation and storage conditions

The extruded kibble-type of dry pet food (10 mm in diameter) was provided by ATbio Co., Ltd. (Namyangju-si, Korea). The ingredients we used in the dry pet food included duck powder, chicken powder, salmon powder, salmon oil, soybean meal, tapioca starch, beet pulp, parsley, yucca extract, flaxseed, fructooligosaccharides, astaxanthin, vitamins (A, B12, D, and E), and minerals (calcium, iron, and zinc). After mixing and grinding the ingredients, the mixture was extruded by heating it to above 70°C for more than 30 seconds at a motor speed of 60Hz. Table 1 shows the the proximate composition of the tested dry pet food.

The samples (approximately 100 g) were packaged in plastic film bags without sealing to simulate the actual storage condition of dry pet food. The packaged samples were divided into two different groups [25°C and 35°C (D25 and D35, respectively)], considering the weather

conditions in Korea, and stored for 120 days. During the storage, RH was monitored as 40%. Then, the samples were collected at 30-day intervals to analyze microorganisms and quality during storage days. Microorganisms, water content, water activity, color, and pH were analyzed right after the samples were collected each storage day. The samples for 2-thiobarbituric acid reactive substance (TBARS) and volatile basic nitrogen (VBN) analysis were stored at -80°C and were tested together at a later time.

#### Water content

The water content measurements were performed according to the analytical methods of the AOAC International (AOAC, 2019). The minced sample (3 g) was spread on a dish and weighed. The dish containing the sample was dried in a drying oven at 105°C for 16 hrs (DS-520L, Daewon Science, Gyeonggi-do, Korea). The water content was obtained by calculating the weight loss of the sample after drying.

Water content (%)

$$= \left(\frac{\text{Weight of sample before drying } - \text{ Weight of sample after drying}}{\text{Weight of sample before drying}}\right)$$

$$\times 100$$

# pН

The pH was measured according to the method described in Jung et al. (2022). The sample (1 g) and 9 mL of distilled deionized water (DDW) was homogenized with the mechanical homogenizer (T25 basic, IKA Works, Inc., Staufen, Germany) at 9,600 rpm for 30 seconds. After centrifuging the homogenate at 2,265 ×g (Continent 512R, Hanil Co., Ltd., Incheon, Korea), the supernatant was filtered through a filter paper (Whatman No.1, Whatman PLC., Kent, UK). Then the pH was measured using a pH meter (Seven2GO, Mettler-Toledo Inc.,

Schwerzenbach, Switzerland) after calibration in standard solutions with pH of 4.01, 7.00, and 9.21, respectively.

#### Water activity

To measure the water activity of the dry pet food, we used a water activity meter (HygroPalm HP23-AW-A, Rotronic, Bassersdorf, Switzerland). The sample cup was filled with 3 g of intact samples without grinding. Then, it was mounted on the machine, adjusting the mode of the machine to the measurement, and the measurement start button was pressed. When the notification sound that it has stabilized, the value was read and recorded.

# Microbial analysis

For microbial analysis, total aerobic bacteria and yeasts and molds were analyzed. First, 25 g of the intact samples and 225 mL of 0.85% sodium chloride were transferred to a sterile Whirl-Pak bags (Nasco, Fort Atkinson, WI, USA). Then, they were blended using a stomacher (BagMixer400P, Interscience, St. Nom, France) for 2 min. The resulting solution was serially diluted, and each dilution was spread on plate count agar (PCA; Difco, Becton Dickinson Co., Sparks, MD, USA) and potato dextrose agar (PDA; Difco, Becton Dickinson Co.). Before enumeration, the PCA and PDA plates were incubated at 37 °C for 48 hours and 25 °C for 120 hours, respectively. Colonies found in the PCA plates were to be counted as total aerobic bacteria (TAB), and those found in PDA plates were to be counted as yeasts and molds (YM). The number of microorganisms counted was to be expressed as CFU/g.

#### Color

A colorimeter (CM-5, Konica Minolta Co., Ltd., Osaka, Japan) with an 8 mm measuring port was used to measure the color of the dry pet food sample. The colorimeter was calibrated using

standard black and white calibration plates (CM-5, Konica Minolta Co., Ltd.). A sample of 10 g was placed on the measuring port and the thickness was kept constant at 20 mm. The color of the sample was expressed as CIE color  $L^*$ ,  $a^*$ , and  $b^*$  which mean lightness, redness, and yellowness, respectively. The mean of five measurements was recorded for different parts of each sample.

## 2-Thiobarbituric acid reactive substances (TBARS)

The TBARS value was determined using the methods described by Shin et al. (2022). First of all, 15 mL of DDW and 50 μL of 7.2% 2,6-Di-terti-butyl-4-methyl-phenol in ethanol were added to 5 g of the minced sample. Then, they were homogenized at 9,600 rpm (T25 basic, IKA Works, Inc.) for 30 seconds. After centrifuging the homogenate at 2,265 ×g (Continent 512R, Hanil Co., Ltd.), the supernatant was filtered through filter paper (Whatman No.4, Whatman PLC.). The filtrated solution (1 mL) was then transferred to a new tube, and 2 mL of 20 mM thiobarbituric acid in 15% trichloroacetic acid was added. After heating them in 90°C of water bath for 30 minutes, the samples were cooled, vortexed, and centrifuged at 2,265 ×g for 15 minutes (Continent 512R, Hanil Co., Ltd.). The absorbance of the supernatant solution was evaluated at 532 nm using a spectrophotometer (M23, Molecular Devices, California, USA). TBARS value were expressed as mg of malondialdehyde (MDA) per kg of the dry pet food. The calculation was carried out using the coefficients obtained from the standard curve.

# Volatile basic nitrogen (VBN)

The VBN value was determined using the Conway's micro-diffusion method (Conway, 1948). Dry pet food (3 g) and DDW (27 mL) were homogenized at 9,600 rpm for 30 seconds (T25 basic, IKA Works, Inc.). After centrifuging the homogenate at 2,265 ×g (Continent 512R, Hanil Co., Ltd.), the supernatant was filtered through filter paper (Whatman No.1, Whatman PLC.). The filtrated solution (1 mL) was added to the outer part of a Conway micro-diffusion

cell. To the center part of the cell, 0.01 N boric acid (1 mL) and 100 μL of indicator was added. The indicator is 1:1 mixture of 0.066% methyl red and 0.066% bromocresol green in ethanol. Then, after adding 1 mL of 50 % potassium carbonate to the another side of the outer cell, the cell was immediately covered and incubated for 2 hours at 37°C. After incubation, the inner cell was titrated with 0.01 N hydrochloric acid (HCl). The VBN value was calculated as follows.

VBN (mg/100g) = 
$$0.14 \times (V_1 - V_0) \times d \times 100$$

Where:  $V_0$  is the volume of 0.01 N HCl (mL) added in the blank,  $V_1$  is the volume of 0.01 N HCl (mL) added in the sample, and d is the dilution factor.

## Statistical analysis

This study was conducted with three replications (n=3). Statistical analysis was performed using SAS software (SAS, Release 9.4; SAS Institute Inc., Cary, NC). All data were assessed using analysis of variance (one-way ANOVA). To compare the mean value of each day, a significance test was performed at the p<0.05 level using the Tukey's multiple comparison test.

#### **Results and Discussion**

## Water content

During 120 days of storage period, slightly different changes in the water content were observed between D25 and D35 (Fig. 1). D25 decreased its water content until day 90 and increased thereafter, while that in D35 was tended to decrease, except for day 90 (p<0.05). Due to the different changes, D35 showed lower water content than D25 on day 120 (17.1 and 17.5%, respectively). This result shows that the water content tended to decrease at the higher temperature. Similar to our study, the study of Paraginski et al. (2014) showed the lowest water content when corn was stored at the highest temperature. However, in our study, the differences

between D25 and D35 were actually very small, at 0.4%. Therefore, changes in the water content of the dry pet food might be negligible regardless of storage temperatures.

#### рH

The pH change can represent food deterioration (Roy and Rhim, 2021). When dry pet food was stored under different storage temperature, their pH value was decreased in both groups with different manners (Table 2). The faster decrease was observed in D35 compared to D25. A prior study has also confirmed that the higher the storage temperature, the higher the decrease in pH value (Jung et al., 2015). The pH of both D25 and D35 showed a decreasing trend (p<0.05) with different rates of decease, but the changes in pH were negligibly small during the 4 months of storage (Table 2). Reduction of pH during food storage is mainly due to microbial fermentation (Tang et al., 2016). However, we did not detect any microorganisms grown in the dry pet food. Dry pet food is formulated to have low moisture content, inhibiting microbial metabolism and enzymatic reactions that could contribute to pH changes (Rezaei et al., 2007). Instead, we found that high temperatures can change the structure of proteins in food, which can lead to a drop in the pH of food (Masson & Lushchekina, 2022). Therefore, we speculate that the reduction in pH may be due to protein denaturation. It has been reported that pH change can affect taste, texture, color, and nutritional value of food (Andrés-Bello et al., 2013). For example, the pH of fried rice dropped significantly from 8.19 to 5.80 after 5 weeks at 25°C, but only decreased from 8.19 to 6.81 at 4°C, and these differences in storage temperatures were shown to affect sensory preference. (Jung et al., 2015). However, in this study, the pH changes after 4 months in D25 and D35 are from 6.26 to 6.22 and from 6.26 to 6.20, respectively. Therefore, the pH reduction under both conditions is practically insignificant.

#### Water activity

Water activity represents the availability of water for biochemical reactions. It is expressed as the ratio of the vapor pressure in a substance to the vapor pressure of pure water (Mathlouthi, 2001). The effect of water activity on food quality is important. For example, when the water activity is 0.80 or higher, fungi easily occur in food (Leistner, 2000). On the other hand, as the water activity value decrease below the monolayer of water (about 0.30), the lipid oxidation rate increases, which negatively affects the quality of food (Nelson & Labuza, 1992).

In the dry pet food, their water activity was 0.23 on day 0 (Fig. 2). However, after 120 days of storage, it was significantly increased in D25 (0.28), whereas D35 (0.11) decreased (p<0.05). This is because the higher temperatures boost the kinetic energy of water molecules, making it easier for them to transition from liquid to a vapor state and reducing water activity (Labuza et al., 1985). But anyway, throughout the experimental period, the water activity range of the dry pet food indicated that the microorganisms could not survive, rather it was close to the value that could easily cause lipid oxidation (Gumus and Decker, 2021).

# Microbial growth

Regardless of different storage conditions, both TAB and YM were not detected in the dry pet food until 120 days of storage. For the growth of microorganisms, it requires conditions such as temperature, moisture, pH, and nutritional sources, and it is difficult to grow well if any of them is lacking (Dixon, 2012). For example, water that is not attached to food molecules supports the growth of bacteria, yeasts, and molds. Water activity refers to this unbound water (Ghaly et al., 2010). It has been reported that most bacteria cannot be grown if water activity is below 0.91 (Sperber, 1983; Beuchat, 1983), and most molds cannot survive if it is below 0.80 (Leistner, 2000). In this study, the water activity was ranged from 0.11 to 0.28, which are lower than possible level for the growth of TAB and YM. Therefore, even at 35°C, a better temperature

for most bacteria to grow, microorganisms could not survive because of their extremely low water activity.

On the other hand, these results could also be attributed to enough sterilization during the sample production process. Tran (2008) notes that the pet food industry primarily manufactures dry pet food using an extrusion process, which effectively sterilizes the food through high-temperature and high-pressure treatment. But the main reason why microorganisms were not detected even after exposing the sample to the outside air is thought to be due to the low water activity at which microorganisms cannot survive.

## Color

In general, the color of food is one of the sensory characteristics and is the standard for guessing the product value, deterioration, freshness, and storage period of food (Lee and Shin, 2019). Table 3 shows the change of color of dry pet food during 120 days at 25°C and 35°C. After 120 days of storage, under both D25 and D35 conditions,  $L^*$  values were decreased significantly, while  $a^*$  values and  $b^*$  values were increased significantly. By day 120, storage temperature notably affected the  $L^*$  values and  $b^*$  values, with no significant impact on the  $a^*$ values. From the  $90^{th}$  day of storage, the  $L^*$  value significantly differed between temperatures, with D35 being lower than D25. In the case of the  $a^*$  value, D35 was significantly higher on day 30, but by day 90, D25 had caught up, showing no significant difference from D35. For the  $b^*$ value, D35 showed a significant increase compared to D25 on day 30, and then fluctuated within a similar value. The increase in redness ( $a^*$  value) and yellowness ( $b^*$  value) is likely due to the Maillard reaction of reducing sugars and amino acids (Li et al., 2020). According to a study by Stapelfeldt et al. (1997), the Hunter b-value of the milk powder increased over the storage period due to the browning caused by the Maillard reaction. In addition, Mouhoubi-Tafinine et al. (2018) found a significant increase in the hydroxymethyl furfural content, the product of the

Maillard reaction, only at storage temperatures above  $30^{\circ}$ C. The decrease in brightness ( $L^*$  value) would be attributed to the pigment produced by the Maillard reaction, which is associated with an increase in redness and yellowness (Cai et al., 2016; Tan et al., 2021). When these results are put together, it can be judged that the color of the dry pet food sample has changed significantly due to the high storage temperature and the lapse of the storage period. This also mean that long storage and high temperature storage induce deterioration of feed.

On the other hand, assessing pet food quality based on color is not effective because each pet food product has unique characteristics and ingredients. Moreover, dogs, the primary consumers of pet food, rely more on smell than color (Landsberg et al., 2011). They have a limited color perception compared to humans, only able to see yellow and blue, due to having fewer types of cone cells in their eyes (Byosiere et al., 2018; Barber et al., 2020). Therefore, from a dog's perspective as a consumer, the quality of the feed cannot be determined by identifying the difference in color.

#### **TBARS**

Malondialdehyde (MDA) is produced due to lipid oxidation and color reaction with thiobarbituric acid is carried out. The more severe the degree of lipid oxidation, they produce a red-pink color, and the higher the TBARS value (Ohkawa et al., 1978). Fig. 3 shows the change of TBARS value of dry pet food during 120 days at 25°C and 35°C. The initial value was 3.77 mg MDA/kg. On the 30<sup>th</sup> day of storage, D35 (4.46 mg MDA/kg) had the higher value than D25 (3.62 mg MDA/kg). Consistent with our predictions, the temperature-dependent relationship with chemical reactions resulted in enhanced lipid oxidation at higher temperatures. (Silbey et al., 2022). In a similar case, Liu et al., (2019) had reported that higher temperatures further promote lipid oxidation in peanuts. Unusually, however, there was no difference due to storage temperature from day 90. This may be attributed to the antioxidants included in the dry pet food.

Some ingredients, such as salmon oil, flaxseed, parsley, vitamin E, and astaxanthin, possess antioxidative activities (King et al., 1992; Oliveira et al., 2018). They may inhibit the further progression of lipid oxidation during storage. In addition, after 120 days, the final values for D35 and D25 were 3.28 and 3.27 mg MDA/kg, respectively, showing a reduction of MDA content from the initial value. This decrease is seen as the loss of TBARS component due to Maillard reaction during storage. Because MDA tends to react with compounds produced by the Maillard reaction (Gomez-Sanchez et al., 1992). There are also previous studies showing that Maillard reaction products have antioxidant properties (Pischetsrieder et al., 1998; Yilmaz and Toledo, 2005). For these reasons, TBARS values can decrease if a Maillard reaction occurred during storage (Hernández et al., 2014). From the present study, we could conclude that 4 months of storage period did not affect the promotion of the lipid oxidation of the dry pet food.

## **VBN**

VBN is generated by the degradation of proteins and amines (Bekhit et al., 2021). Its formation pathway is mainly related to the activity of endogenous enzymes and contaminated bacteria (Kathuria et al., 2022). Fig. 4 shows the change of VBN of dry pet food during 120 days at 25°C and 35°C. The initial value was 7.9 mg/100 g. From the 90th day of storage, both D25 and D35 showed an increase of 1.2 mg/100 g and the same value was maintained at 120 days. Since no microorganisms have been found in this study, it is difficult to say that it is an increase caused by bacteria. But this could be attributed to protein degradation caused by endogenous enzyme remaining in the dry pet food (Holman et al., 2021). Anyway, it was a very small increase and showed a stable level that did not reach the standard of 20 mg/100 g for decay. Therefore, it can be concluded that the dry pet food sample was not decomposed overall regardless of the storage temperature and period.

## Conclusion

For 120 days, no microorganisms were detected from the dry pet food at both 25°C and 35°C storage temperatures. Due to very low water activity, the quality characteristics such as pH, TBARS, and VBN changed very slightly. Therefore, we conclude that dry pet food can be store at room temperature or up to 35°C for 4 months without compromising its quality.

## **Conflicts of interest**

The authors declare no potential conflicts of interest.

# Acknowledgements

This study was supported by the "Cooperative Research Program for Agriculture Science and Technology Development (Project No. PJ016891)" Rural Development Administration, Republic of Korea.

## **Author Contributions**

Conceptualization: Yim DG, Park S, Jo C. Data curation: Park D, Lee HJ. Formal analysis: Park D, Anand Kumar S. Validation: Park D, Lee HJ. Investigation: Yim DG, Park S. Writing - original draft: Park D, Lee HJ. Writing - review & editing: Park D, Lee HJ, Anand Kumar S, Yim DG, Park S, Jo C.

## **Ethics Approval**

This article does not require IRB/IACUC approval because there are no human and animal participants.

## References

- Andrés-Bello A, Barreto-Palacios V, García-Segovia P, Mir-Bel J, Martínez-Monzó J. 2013. Effect of pH on color and texture of food products. Food Eng Rev 5:158-170.
- Barber ALA, Mills DS, Montealegre-ZF, Ratcliffe VF, Guo K, Wilkinson A. 2020. Functional performance of the visual system in dogs and humans: A comparative perspective. Comparat Cogn Behav Rev 15:1-44.
- Bekhit AEDA, Holman BWB, Giteru SG, Hopkins DL. 2021. Total volatile basic nitrogen (TVB-N) and its role in meat spoilage: A review. Trends Food Sci Technol 109:280-302.
- Beuchat LR. 1983. Influence of water quality on growth, metabolic activities and survival of yeasts and molds. J Food Protect 46:135–141.
- Byosiere SE, Chouinard PA, Howell TJ, Bennett PC. 2018. What do dogs (*Canis familiaris*) see?

  A review of vision in dogs and implications for cognition research. Psychon Bull Rev
  25:1798-1813.
- Cai L, Li D, Dong Z, Cao A, Lin H, Li J. 2016. Change regularity of the characteristics of Maillard reaction products derived from xylose and Chinese shrimp waste hydrolysates. LWT 65:908-916.
- Carrión PA. 2023. Pet food. In Food safety management. 2<sup>nd</sup> ed. Veslemøy A, Huub L, Yasmine M (ed). Academic Press, Cambridge, MA, USA. pp 363-384.
- Conway EJ. 1948. Microdiffusion analysis and volumetric error. Nature 161:583-583.
- Crump JA, Griffin PM, Angulo FJ. 2002. Bacterial contamination of animal feed and its relationship to human foodborne illness. Clin Infect Dis 35:859-865.
- Dixon GR. 2012. Climate change–impact on crop growth and food production, and plant pathogens. Can J Plant Pathol 34:362-379.

- Gazzotti T, Biagi G, Pagliuca G, Pinna C, Scardilli M, Grandi M, Zaghini G. 2015. Occurrence of mycotoxins in extruded commercial dog food. Anim Feed Sci Technol 202:81-89.
- Ghaly AE, Dave D, Budge S, Brooks MS. 2010. Fish spoilage mechanisms and preservation techniques. Am J Appl Sci 7:859-877.
- Gomez-Sanchez A, Hermos in I, Maya I. 1992. Influence of malondial dehyde on the Maillard degradation of Amadori compounds. Carbohydr Res 229:307-322.
- Gumus CE, Decker EA. 2021. Oxidation in low moisture foods as a function of surface lipids and fat content. Foods 10:860.
- Hernández A, García BG, Jordán MJ, Hernández MD. 2014. Natural antioxidants in extruded fish feed: Protection at different storage temperatures. Anim Feed Sci Technol 195:112-119.
- Holman BW, Bekhit AEDA, Waller M, Bailes KL, Kerr MJ, Hopkins DL. 2021. The association between total volatile basic nitrogen (TVB-N) concentration and other biomarkers of quality and spoilage for vacuum packaged beef. Meat Sci 179:108551.
- Jung DY, Lee HJ, Shin DJ, Kim CH, Jo C. 2022. Mechanism of improving emulsion stability of emulsion-type sausage with oyster mushroom (*Pleurotus ostreatus*) powder as a phosphate replacement. Meat Sci 194:108993.
- Jung JH, Lim JH, Jeong MJ, Jeong IH, Kim BM. 2015. Changes in quality of fried rice with red snow crab meat depending on the storage period and temperature. Korean J Food Cook Sci 31:387-394.
- Kathuria D, Dhiman AK, Attri S. 2022. Sous vide, a culinary technique for improving quality of food products: A review. Trends Food Sci Technol 119:57-68.
- King MF, Boyd LC, Sheldon BW. 1992. Antioxidant properties of individual phospholipids in a salmon oil model system. J Am Oil Chem Soc 69:545-551.
- Labuza TP, Kaanane A, Chen JY. 1985. Effect of temperature on the moisture sorption isotherms and water activity shift of two dehydrated foods. J Food Sci 50:385-392.

- Landsberg G, Hunthausen W, Ackerman L. 2011. Behavior problems of the dog and cat. 3<sup>rd</sup> ed. Elsevier Health Sciences, Amsterdam, Netherlands. pp 13-22.
- AOAC. 2019. Official methods of analysis of AOAC International. 21th ed. AOAC International, Gaithersburg, MD, USA.
- Lee EJ, Shin HS. 2019. Development of a freshness indicator for monitoring the quality of beef during storage. Food Sci Biotechnol 28:1899-1906.
- Leistner L. 2000. Basic aspects of food preservation by hurdle technology. Int J Food Microbiol 55:181-186.
- Leiva A, Molina A, Redondo-Solano M, Artavia G, Rojas-Bogantes L, Granados-Chinchilla F. 2019. Pet Food Quality Assurance and Safety and Quality Assurance Survey within the Costa Rican Pet Food Industry. Animals 9:980.
- Li X, Gao K, Jinfeng B, Wu X, Li X, Guo C. 2020. Investigation of the effects of apple polyphenols on the chromatic values of weakly acidic lysine-fructose maillard system solutions. LWT 125:109237.
- Liu K, Liu Y, Chen F. 2019. Effect of storage temperature on lipid oxidation and changes in nutrient contents in peanuts. Food Sci Nutr 7:2280-2290.
- Masson P, Lushchekina S. 2022. Conformational stability and denaturation processes of proteins investigated by electrophoresis under extreme conditions. Molecules, 27:6861.
- Mathlouthi M. 2001. Water content, water activity, water structure and the stability of foodstuffs. Food Control 12:409-417.
- Morelli G, Stefanutti D, Ricci R. 2021. A Survey among Dog and Cat Owners on Pet Food Storage and Preservation in the Households. Animals 11:273.
- Mouhoubi-Tafinine Z, Ouchemoukh S, Bey BM, Louaileche H, Tamendjari A. 2018. Effect of storage on hydroxymethylfurfural (HMF) and color of some Algerian honey. Int Food Res J 25:1044-1050.

- Nelson KA, Labuza TP. 1992. Relationship between water and lipid oxidation rates: Water activity and glass transition theory. In Lipid Oxidation in Food. American Chemical Society, Washington, DC, USA. pp 98-103.
- Ohkawa H, Ohishi N, Yagi K. 1979. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal Biochem 95:351-358.
- Oliveira VS, Ferreira FS, Cople MCR, Labre TDS, Augusta IM, Gamallo OD, Saldanha T. 2018.

  Use of natural antioxidants in the inhibition of cholesterol oxidation: A review. Compr Rev
  Food Sci Food Saf 17:1465-1483.
- Onilude AA, Igbinadolor RO, Wakil SM. 2010. Effect of varying relative humidity on the rancidity of cashew (*Anacardium occidentale L*.) kernel oil by lipolytic organisms. Afr J Biotechnol 9:4890–4896.
- Paraginski RT, Vanier NL, Berrios JDJ, Oliveira M, Elias MC. 2014. Physicochemical and pasting properties of maize as affected by storage temperature. J Stored Prod Res 59:209-214.
- Pischetsrieder M, Rinaldi F, Gross U, Severin T. 1998. Assessment of the antioxidative and prooxidative activities of two aminoreductones formed during the Maillard reaction: effects on the oxidation of  $\beta$ -carotene, N  $\alpha$ -acetylhistidine, and cis-alkenes. J Agric Food Chem 46:2945-2950.
- Prata JC. 2022. Survey of Pet Owner Attitudes on Diet Choices and Feeding Practices for Their Pets in Portugal. Animals 12:2775.
- Robert R, Stewart C, Bewley JD. 1980. Lipid peroxidation associated with accelerated aging of soybean axes. Plant Physiol 65:245–248.
- Roy S, Rhim JW. 2021. Anthocyanin food colorant and its application in pH-responsive color change indicator films. Crit Rev Food Sci Nutr 61:2297-2325.

- Scudamore KA, Livesey CT. 1998. Occurrence and significance of mycotoxins in forage crops and silage: a review. J Sci Food Agric 77:1-17.
- Shin DJ, Yim DG, Kwon JA, Kim SS, Lee HJ, Jo C. 2022. Effect of cutting time and cooking temperature on physicochemical properties of chicken breast meat emulsion sausage with olive oil. Poultry Sci 101:101554.
- Silbey RJ, Alberty RA, Papadantonakis GA, Bawendi MG. 2022. Physical chemistry. 5<sup>th</sup> ed. Hoboken, NJ, USA.
- Sperber WH. 1983. Influence of Water Activity on Foodborne Bacteria A Review1. J Food Prot 46:142-150.
- Stapelfeldt H, Nielsen BR, Skibsted LH. 1997. Effect of heat treatment, water activity and storage temperature on the oxidative stability of whole milk powder. Int Dairy J 7:331-339.
- Tan JE, Liu T, Yao Y, Wu N, Du H, Xu M, Liao M, Zhao Y, Tu Y. 2021. Changes in physicochemical and antioxidant properties of egg white during the Maillard reaction induced by alkali. LWT 143:111151.
- Tang J, Wang X, Hu Y, Zhang Y, Li Y. 2016. Lactic acid fermentation from food waste with indigenous microbiota: Effects of pH, temperature and high OLR. Waste Manag 52:278-285.
- Tran QD, Hendriks WH, van der Poel AF. 2008. Effects of extrusion processing on nutrients in dry pet food. J Sci Food Agric 88:1487-1493.
- Yilmaz Y, Toledo R. 2005. Antioxidant activity of water-soluble Maillard reaction products. Food Chem 93:273-278.

## Figure legends

- Fig. 1. Changes in water content during storage days of the dry pet food sample stored at different temperatures. A-B Values with different letters between different storage conditions have significant differences (p<0.05). a-d Values with different letters between different storage periods have significant differences (p<0.05). D25, dry pet food stored indoors at an average temperature of 25°C; D35, dry pet food stored in a warm chamber set at 35°C.
- Fig. 2. Changes in water activity during storage days of the dry pet food sample stored at different temperatures. <sup>A-B</sup>Values with different letters between different storage conditions have significant differences (p<0.05). <sup>a-b</sup>Values with different letters between different storage periods have significant differences (p<0.05). D25, dry pet food stored indoors at an average temperature of 25°C; D35, dry pet food stored in a warm chamber set at 35°C.
- Fig. 3. Changes in 2-thiobarbituric acid reactive substances (TBARS) value during storage days of the dry pet food sample stored at different temperatures. A-B Values with different letters between different storage conditions have significant differences (p<0.05). a-b Values with different letters between different storage periods have significant differences (p<0.05). D25, dry pet food stored indoors at an average temperature of 25°C; D35, dry pet food stored in a warm chamber set at 35°C.
- Fig. 4. Changes in volatile basic nitrogen (VBN) value during storage days of the dry pet food sample stored at different temperatures. <sup>a-b</sup>Values with different letters between different storage periods have significant differences (p<0.05). D25, dry pet food stored indoors at an average temperature of 25°C; D35, dry pet food stored in a warm chamber set at 35°C.

Table 1. Proximate composition of the dry pet food sample on day 0

Items	Percentage (%)		
Moisture	18.41		
Crude protein	29.87		
Crude fat	19.33		
Crude fiber	1.70		
Crude ash	6.69		
Calcium	1.59		
Phosphorous	0.95		



Table 2. Changes in pH of the dry pet food stored at different temperatures

Tuestaseat	Storage (day)				SEM <sup>1</sup>
Treatment	0	30	90	120	SEWI
D25	6.26 <sup>a</sup>	6.25 <sup>ab</sup>	6.22 <sup>Abc</sup>	6.22°	0.006
D35	6.26 <sup>a</sup>	6.25 <sup>a</sup>	$6.20^{\mathrm{Bb}}$	$6.20^{b}$	0.006
$SEM^2$	0.011	0.003	0.002	0.004	

<sup>&</sup>lt;sup>1</sup>Standard error of the mean (n = 12), <sup>2</sup> (n = 6).

D25, dry pet food stored indoors at an average temperature of 25°C; D35, dry pet food stored in a warm chamber set at 35°C.

 $<sup>^{</sup>A-B}$ Different letters within the same column indicate significant differences (p<0.05).

<sup>&</sup>lt;sup>a-c</sup>Different letters within the same row indicate significant differences (p<0.05).

Table 3. Changes in color of the dry pet food stored at different temperatures

Item	Treatment -	Storage (day)				CEM1
		0	30	90	120	SEM <sup>1</sup>
CIE L*	D25	38.78 <sup>a</sup>	37.75 <sup>ab</sup>	37.38 <sup>Ab</sup>	37.31 <sup>Ab</sup>	0.314
	D35	$38.78^{a}$	38.03 <sup>a</sup>	$35.21^{Bb}$	$35.64^{\mathrm{Bb}}$	0.360
	$SEM^2$	0.507	0.204	0.293	0.269	
CIE a*	D25	8.34 <sup>b</sup>	$8.62^{\mathrm{Bb}}$	$9.56^{a}$	$9.32^{a}$	0.129
	D35	8.34 <sup>b</sup>	$9.39^{Aa}$	$9.32^{a}$	$9.38^{a}$	0.140
	$SEM^2$	0.234	0.066	0.097	0.059	
CIE b∗	D25	19.94 <sup>b</sup>	$20.79^{\mathrm{Bb}}$	23.54 <sup>Aa</sup>	$23.15^{Aa}$	0.232
	D35	19.94 <sup>b</sup>	23.62 <sup>Aa</sup>	21.57 <sup>Ba</sup>	22.01 <sup>Ba</sup>	0313
	$SEM^2$	0.333	0.197	0.303	0.250	

<sup>&</sup>lt;sup>1</sup>Standard error of the mean (n = 20), <sup>2</sup> (n = 10).

D25, dry pet food stored indoors at an average temperature of 25°C; D35, dry pet food stored in a warm chamber set at 35°C.

A-B Different letters within the same column indicate significant differences (p<0.05).

<sup>&</sup>lt;sup>a-b</sup>Different letters within the same row indicate significant differences (p<0.05).

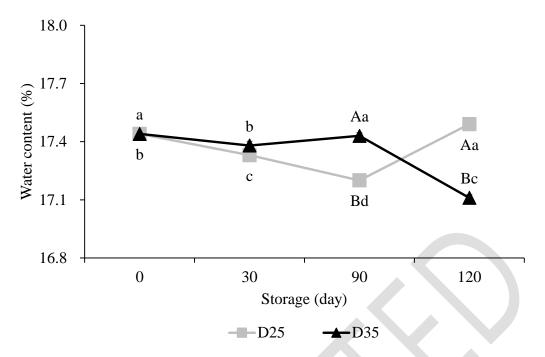


Fig. 1.

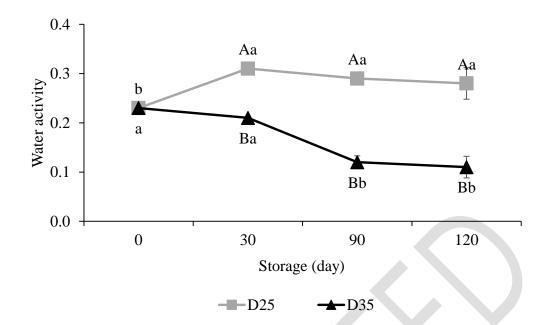


Fig. 2.

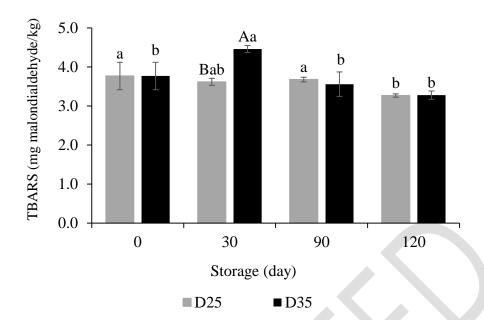


Fig. 3

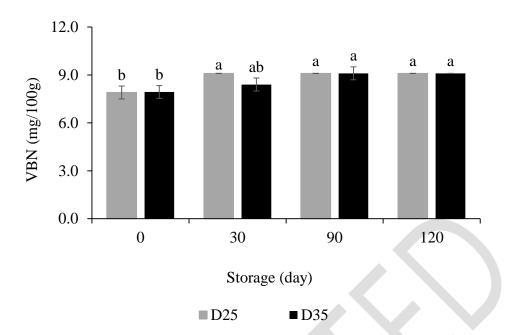


Fig. 4.