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8 **Current status of non-thermal sterilization by pet food raw ingredients**

9

10

Abstract

11 Recently, as the concept of pet food that satisfies both nutritional needs and the five senses has evolved,
12 so too has the demand for effective pet food non-thermal sterilization methods. Prominent non-thermal
13 technologies include high-pressure processing (HPP), plasma, and radiation, which are favored for their
14 ability to preserve nutrients, avoid residues, and minimize compositional changes, thereby maintaining
15 quality and sensory properties. However, to assess their effectiveness on pet food, it is essential to
16 optimize operational parameters such as pressure levels, plasma intensity, radiation dosage, and
17 temperature. Further studies are needed to evaluate microbial sterilization efficacy and sensory
18 attributes. This exploration is expected to lay the groundwork for preventing zoonotic diseases and
19 improving the production of high-quality pet food.

20

21 **Keywords** pet food, non-thermal sterilization, high-pressure processing, plasma, radiation

22 Introduction

23 In recent years, the pet market, both domestically and internationally, has been expanding alongside
24 an increase in pet-owning households. This growth is influenced by social phenomena such as aging,
25 non-marriage, and urbanization, which have led to a rise in dual-income, no-kids (DINKs) households,
26 nuclear families, and single-person households (Choe et al., 2023). As of 2022, one- and two-person
27 households, which include DINKs, nuclear families, and single-person households, comprised 65% of
28 the total household composition, significantly impacting pet ownership growth (Ministry of the Interior
29 and Safety, 2023). According to the Ministry of Agriculture: Food and Rural Affairs "Announcement of
30 2022 public awareness survey on animal protection" (2023), there were 6.02 million households with
31 pets, representing 25.4% of all households. The pet market is anticipated to continue its growth
32 trajectory, with projections showing an increase to 3.24 billion dollars in 2023, a 36.36% rise from 2.37
33 billion dollars in 2020 (Korea Rural Economic Institute, 2018).

34 The global pet market is broadly categorized into pet food, pet healthcare, pet services, and pet
35 technology sectors. Among these, pet food dominated, accounting for approximately 70.94% (98.07
36 million dollars) of the total global pet market value of 138.24 million dollars as of 2020, marking it as
37 the fastest-growing category (Gromek and Perek-Bialas, 2022). In the Korean market, pet food
38 represents 31.5% of the total pet-related industry and is growing at a rate of 5.4% annually (Korea
39 Consumer Agency, 2021). Additionally, pet food constitutes 50.7% of the total pet care expenses,
40 highlighting the expected growth in this sector (KB Management Research Institute, 2021).

41 The current trend in pet food is "pet humanization" which involves the anthropomorphization of pets.
42 As part of this trend, pet foods such as "human grade"—made from ingredients safe for human
43 consumption—and "raw food"—which consider the natural eating habits of pets—are being researched
44 and developed (Ye et al., 2022). Additionally, many pet owners view their pets as family members,
45 evidenced by neologisms such as "pet fam" and "pet me" in Korea, and "fur babies" and "fur family" in
46 English-speaking countries (Rauktis et al., 2017 ; Seo, 2024). Consequently, pet food is developing
47 from "feed" meant for herbivorous livestock to "food" intended for companion animals (Park et al.,

48 2022). Therefore, for the pet food market to continue its expansion, it is crucial to manage protein-
49 oriented animal feeds, distinct from traditional grain-oriented livestock feeds, and to establish effective
50 non-thermal sterilization conditions for the distribution of pet food. Research on pet food forms, such
51 as semi-wet, wet, and freeze-dried, which have better palatability than dry food, has significantly
52 increased (Geary et al., 2023). However, semi-wet and wet pet foods, due to their higher moisture
53 content, are more prone to microbial growth than dry pet foods (Watson et al., 2023). Moreover, health-
54 conscious consumers (pet owners) prefer pet food without synthetic agents, which, while more natural,
55 can deteriorate more quickly and require effective sterilization methods.

56 There are two main types of sterilization used to extend the shelf life of food: heat treatments such as
57 warming, microwaves, and infrared radiation, and non-thermal treatments such as high-pressure
58 processing (HPP), plasma, and radiation (Wang et al., 2023). Among these, heat treatments are the most
59 common sterilization method, where food in various containers, such as pouches, cans, trays, and bottles,
60 is heated directly or indirectly until the target sterilization temperature is reached (Barbosa-Cánovas et
61 al., 2014). This process is crucial as it inhibits microbial activity, enhances flavor, and induces physical
62 changes in food (Miri et al., 2008). However, heat treatment, typically at 121–140°C, can reduce quality
63 characteristics such as flavor and color and destroy some nutritional components (Lazárková et al.,
64 2011). Among nutritional components, proteins such as myoglobin are heat denatured at 30–35°C, and
65 collagen begins denaturing at 58–65°C, disrupting the polypeptide structure (Vinnikova et al., 2019).
66 Myoglobin, a plasma protein, changes color with heating—red at 60°C, pink at 60–70°C, and grayish-
67 brown above 70–80°C—due to oxidation, significantly affecting the palatability of meat products (Han
68 et al., 2024). Furthermore, the denaturation of collagen, a connective tissue protein, triggers protein
69 hydrolysis, reducing its hardness; excessive heat denaturation can increase its toughness (Yu et al.,
70 2017). Given the challenges associated with heat treatment, particularly for meat and fish raw
71 ingredients rich in proteins, which are sensitive to heat, there is a transition to non-thermal sterilization
72 technologies. These methods are being proposed as alternatives to traditional sterilization, especially
73 for pet foods containing substantial amounts of animal protein (Lim and Ha, 2020; Shin, 2020).

74 Non-thermal sterilization is a technique that effectively minimizes the alteration of raw ingredients
75 and inhibits microorganisms, incorporating physical treatments such as HPP, radiation, electric fields,
76 and plasma, as well as chemical treatments using chemicals or cell wall-degrading enzymes (Song,
77 2020). With increasing consumer demand for high-quality foods that are healthful, safe from
78 contamination, and free from synthetic products, non-thermal sterilization treatments are gaining
79 attention as methods to ensure effective sterilization. Currently, non-thermal sterilization is primarily
80 applied to raw food ingredients such as cereals, fruits and vegetables, seafood, and meat products, where
81 traditional heating methods may reduce efficacy (Chauhan et al., 2018). This technique halts the
82 germination of grains, slows the ripening of fruits, inhibits the growth of parasites and microorganisms
83 in food, and ultimately extends shelf life (Jan et al., 2017). For meat and fish, which are primary
84 ingredients in pet food, microbial control is achieved through high-pressure washing, vacuum and
85 modified atmosphere packaging, low-temperature systems, and disinfectant water sprays (Sohaib et al.,
86 2016). However, meat and fish raw ingredients are more prone to external contamination and spoilage
87 than vegetable ingredients, necessitating effective sterilization methods to eradicate harmful
88 microorganisms completely (Molins et al., 2001). In recent years, non-thermal sterilization methods
89 like HPP, plasma, and radiation have been explored for various meat and fish raw ingredients used in
90 raw pet food (Lee et al., 2023 ; Neshovska et al., 2023 ; Serra-Castelló et al., 2023 ; Yadav et al., 2020).

91 The Association of American Feed Control Officials (AAFCO) stipulates that dry pet food should
92 contain 18% to 22.5% crude protein and 5.5% to 8.5% crude fat (Dodd et al., 2021). Consequently, the
93 main ingredients of domestically and internationally distributed pet food typically include meat
94 products such as chicken, beef, pork, and duck, and fish products such as mackerel, tuna, and salmon,
95 along with by-products from these sources (Montegiove et al., 2021). Unlike advanced countries in the
96 pet food industry like the United States and Europe, Korea lacks a standard for the non-thermal
97 sterilization of pet food, necessitating a comprehensive review of existing research to establish such a
98 standard.

99

100 High-pressure processing

101 **Food high-pressure processing technology**

102 HPP, also known as high hydrostatic pressure or ultra-high-pressure (UHP), is a technology that can
103 effectively sterilize harmful microorganisms while maintaining the quality characteristics of food,
104 such as flavor, color, and nutritional content (Seo et al., 2014). This method applies Le Chatelier's
105 chemical equilibrium principle, where the volume and number of molecules in food decrease under
106 pressure, leading to microbial cell membrane disintegration, protein denaturation, and sterilization
107 (Renaud et al., 2022). Additionally, cold isostatic pressure involves pressurizing food with a liquid
108 medium at low or room temperature (Dalai and Sahu, 2010).

109 During HPP, covalent bonds are less affected by pressure, while weaker electrostatic and
110 hydrophobic bonds undergo structural deformation due to pressure (Campus, 2010). This deformation
111 destroys the cellular structure of microorganisms and inactivates enzymes, but does not affect small
112 molecular size vitamins and flavor compounds, thus, preserving nutritional substances (Albert et al.,
113 2021). Moreover, HPP has the advantage of minimizing food deformation since there is no heat
114 involved, providing uniform and instantaneous treatment to food through a liquid medium at pressures
115 ranging from 100 to 600 MPa, resulting in minimal deformation in the size and shape of food (Li and
116 Farid, 2016). Consequently, HPP can be applied not only to raw food products but also to packaged
117 finished products, making it a highly adaptable technology in the food industry (Koutsoumanis et al.,
118 2022).

119 HPP has applications across various fields including food, pharmaceuticals, and medicine,
120 predominantly within the food industry. This application in the food sector began when Hite (1899)
121 observed a reduction in microorganism levels in milk and meat subjected to high-pressures of
122 500~600 MPa (Torres and Velazquez, 2005). Throughout the 20th century, research into HPP
123 technology expanded in countries like the United States, the United Kingdom, and Japan, leading to
124 the development of HPP equipment suitable for sterilizing food by eliminating harmful
125 microorganisms. Figure 1 presents a schematic diagram of the structure and operational process of a

126 currently commercialized HPP device. The container, liquid medium, and pressure range of the device
127 have been standardized, facilitating the wider use of HPP in food processing.

128 As a result, a variety of high-pressure processed and pasteurized foods, including meat products,
129 seafood, juices, jams, and purees, have been commercialized. The primary microorganisms affected
130 by HPP include *Listeria* spp., and *Campylobacter* spp., making HPP particularly advantageous for
131 sterilizing meat and fish raw ingredients, which are highly susceptible to microbial contamination
132 (Campus, 2010). However, it has been reported that foods rich in nutrients, such as carbohydrates,
133 proteins, fats, and salts, or those with low water activity, can exhibit microbial protective effects
134 against HPP (Govaris and Pexara, 2021). Therefore, microbial sensitivity to pressure can vary
135 depending on the state of the food, highlighting the need to establish specific sterilization conditions
136 for each type of food.

137 Countries like the United States, the United Kingdom, and Canada, which are advanced in the
138 development of HPP, are actively implementing HPP technology in the food industry and have
139 established policies for foods processed by HPP. The U.S. Food Safety and Inspection Service (FSIS),
140 the U.K. Food Standards Agency (FSA), Canada's Health Canada, and the Food Standards Agency of
141 Australia and New Zealand (FSANA) have all set policies for the application of HPP in foods (Huang
142 et al., 2017). In contrast, South Korea lacks specific policies and standards for foods processed by
143 high-pressure, underscoring the necessity to establish precise pressure levels and durations to
144 effectively sterilize microorganisms during HPP application.

145

146 **Setting the pet food high-pressure processing standards**

147 Table 1 reviews prior literature on the application of HPP sterilization to meat and fish raw
148 ingredients, which are the primary animal proteins used in pet food. This table illustrates that HPP,
149 predominantly utilized for sterilizing meat and fish products, can achieve the five-log reduction in
150 microorganisms mandated by international regulatory bodies by processing at 400 to 600 MPa at
151 temperatures of 4°C or 25°C in under 10 min (Bolumar et al., 2021). Key control points for effective
152 sterilization include processing temperature, pressure, and time. Notably, lower temperatures, such as

153 4°C, have proven to be more effective in eliminating microorganisms than temperatures around 25°C,
154 with the effectiveness of the microbial disinfection increasing with longer processing times.
155 Additionally, the processing pressure impacts the physical state of proteins in the sample and
156 myoglobin oxidation (Pou, 2021).

157 Approximately 30% of processed foods treated with high-pressure sterilization are meat products.
158 The sterilization of meat products using high-pressure is already widespread, as the quality and
159 sensory properties of meat products are relatively stable under high-pressure conditions compared to
160 fresh meat (Bolumar et al., 2021). However, unlike processed meat products, raw meat is influenced
161 by various factors such as the characteristics of the meat, and the temperature, pressure, and duration
162 of processing. For example, chicken or fish meat, which is softer than pork or beef, tends to undergo
163 protein gelation when pressurized at 200 to 300 MPa (Chen et al., 2018). Additionally, the color of
164 meat changes more sensitively in red meats rich in myoglobin. Pressurizing above 300 MPa disrupts
165 the physical structure of myoglobin, accelerating oxidation, which results in increased brightness and
166 reduced redness of the meat; however, these color changes induced by HPP treatment tend to lessen
167 over extended storage periods (Bak et al., 2017).

168 Therefore, it is necessary to compare the sterilization efficiency and quality between raw ingredients
169 and finished products and to investigate the suitable HPP conditions for microorganisms with high
170 contamination potential depending on the type of product (dry, wet, semi-wet, freeze-dried, etc.).

171

172 Plasma

173 **Food plasma sterilization technology**

174 Plasma is a fourth state of matter in which ions and electrons, separated by high energy, are either in
175 an energy equilibrium or non-equilibrium state, as depicted in Figure 2. It consists of ions, electrons,
176 free radicals, and ultraviolet radiation, serving as the source for phenomena such as lightning, aurora
177 borealis, neon signs, and fluorescent lights (Jeon et al., 2020). Plasma can be categorized into hot
178 plasma (equilibrium plasma; arc discharges) and cold plasma (non-equilibrium plasma; corona and
179 dielectric barrier discharges (DBDs)), according to the equilibrium state of ions and electrons, as

180 illustrated in Figure 3. Cold plasma is also referred to as non-equilibrium plasma because the electron
181 temperature is higher than the overall gas temperature (Fiebrandt et al., 2018).

182 Furthermore, low-temperature plasmas are produced and sustained in an ionized gas state by arc
183 discharge, corona discharge, and DBD at low-pressure (depressurization) or at normal pressure
184 (atmospheric pressure) (Birania et al., 2022). Arc discharge is characterized by generating high-
185 temperature plasma in a very confined area at normal pressure, while corona discharge also generates
186 plasma at normal pressure, resulting in a limited processing area, albeit with comparatively less heat
187 generation than arc discharge (Dalvi-Isfahan et al., 2023). Conversely, dielectric barrier plasma is
188 created by attaching one or two dielectric plates to two parallel metal plates connected to direct
189 current or alternating current electrodes, producing less heat and covering a larger processing area
190 (Nasiru et al., 2021).

191 Among the low-temperature plasma technologies, normal pressure plasma is predominantly utilized
192 in conjunction with low-temperature sterilization. However, controlling the plasma production rate is
193 challenging, and temperature increases can occur due to excessive energy input (Domonkos et al.,
194 2021). In contrast, low-pressure plasma is ideal for non-thermal sterilization as it allows for a
195 controlled rate of plasma generation and minimizes heat production (Fiebrandt et al., 2018).
196 Alongside low-pressure plasma, plasma-activated water (PAW) is also being explored as a non-
197 thermal sterilization method for meat, fruits, and vegetables (Sammanee et al., 2022).

198 Microbial sterilization by plasma has been explored since the mid-1990s, though its practical
199 applications in the food industry commenced in the early 2000s (Li and Farid, 2016). Plasma
200 sterilization is particularly advantageous because it does not involve chemical treatments or leave
201 residues. Its uses in the food industry have expanded to include sterilizing microorganisms, removing
202 contaminants, extending shelf life, inactivating enzymes, eliminating toxins, enhancing food
203 packaging, and treating wastewater (Hati et al., 2018).

204 The mechanism of microbial sterilization by plasma primarily involves DNA modification through
205 ultraviolet rays emitted by the plasma (UV inactivation), the release of volatile substances from
206 chemical bond sterilization (photodesorption), and the adsorption reaction of reactive compounds

207 (etching) (Mravlje et al., 2021). This method is effective against harmful pathogens such as
208 *Escherichia coli*, *Salmonella* spp., *Listeria* spp., noroviruses, and hepatitis viruses. These pathogens
209 are commonly found in environments associated with animal slaughter, making them critical targets
210 for sterilization in the food processing industry.

211 Plasma technology is frequently applied to meat products such as chicken, pork, and beef. It
212 efficiently sterilizes various harmful substances and microorganisms while minimally impacting
213 quality characteristics like color and texture, making it a valuable technology for ensuring safety in
214 the food supply (Misra and Jo, 2017).

215

216 **Setting the pet food plasma sterilization standards**

217 Table 2 is a prior study of plasma sterilization technologies applied to meat and fish raw ingredients,
218 which are the animal proteins most commonly used in pet food. The table shows that plasma,
219 primarily utilized for sterilizing meat and fish, operates under normal pressure or low-pressure at
220 temperatures below 44°C. This setup minimizes protein modification and is achieved through the use
221 of dielectric barrier plasma or PAW (Sen and Mutlu, 2013).

222 Non-thermal plasma sterilization applied to chicken has been mainly studied for chicken breasts and
223 thighs, using dielectric plasma (Lee et al., 2020 ; Zhao et al., 2022) or immersion in PAW (Kang et
224 al., 2022 ; Qian et al., 2021 ; Sammanee et al., 2022). However, chicken with skin has a non-uniform
225 surface, which reduces the microbial inactivation effect compared to chicken with a uniform surface
226 (Noriega et al., 2011). Effective microbial inactivation of chicken skin can be achieved with a
227 radiation gap of 1.0 cm, a voltage of 7.0 kV, and a frequency of 38.5 kHz for 3 min, or a 30 s
228 exposure at 8.0 kV. Conversely, skinless chicken breasts showed negative results for *Listeria* at a
229 radiation distance of 1.0 cm, voltages of 8.0–11.0, and frequencies of 38.5–30.0 kHz, suggesting that
230 higher voltage and frequency are effective in sterilizing *Listeria* in chicken.

231 Other factors affecting the effectiveness of plasma sterilization include exposure time, flow rate,
232 moisture, temperature, and gas composition. Regarding atmospheric composition, nitrogen, argon,

233 and helium are the most commonly used gases (Ulbin-Figlewicz et al., 2015). The effects of vacuum
234 dielectric plasma sterilization of pork loin with these different gas compositions showed that helium
235 had the highest microbial inactivation rate, with yeast and mold showing higher disinfection effects
236 than total microorganisms and thermophilic bacteria. Plasma with different gas compositions did not
237 significantly affect the pH of the sample, indicating that it does not cause significant changes to the
238 original product. Despite these advantages, dielectric barrier plasma has the drawback of requiring
239 large and expensive equipment for large-scale treatment. Consequently, PAW is being researched as a
240 more feasible alternative (Du et al., 2022).

241 PAW is produced by passing an electrode through the liquid in a water tank by using a plasma
242 generator, and this PAW is treated by spraying it onto the surface of the sample or immersing the
243 sample in it (Hadinoto et al., 2023 ; Lotfy and Khalil, 2022 ; Qian et al., 2022). This type of PAW is
244 typically produced using hydrogen peroxide, perchloric acid, sodium hypochlorite, and acetic acid.
245 The typical application conditions for PAW involve a 10 min treatment at temperatures around 25°C.
246 Studies have shown that hydrogen peroxide plasma treatment of pork and chicken is highly effective
247 against pathogens such as *Campylobacter* and *Salmonella Enteritidis*, but not against *Pseudomonas*
248 (Sammanee et al., 2022). The pH and chromaticity measurements indicate variations between deep
249 and surface treatments, suggesting that the use of PAW should be carefully managed to avoid
250 adversely affecting the palatability of the food. When applied to mackerel, PAW also resulted in a
251 decrease in pH without significant changes in chromaticity.

252 Unlike dielectric barrier plasma, PAW can be used in large quantities either through immersion or
253 spraying. However, it is important to assess the effects of different types of PAW on the palatability
254 of food to ensure suitability. Therefore, while plasma sterilization of pet food ingredients and finished
255 products with uniform surfaces is typically superior in maintaining quality and sensory properties
256 when treated with dielectric barrier plasma at around 25°C, this method is more costly compared to
257 treatments using PAW. Consequently, comparative studies between different plasma treatment
258 methods for pet food ingredients and finished products are essential to determine the most cost-
259 effective and quality-preserving sterilization technique.

260

261 Radiation

262 **Food radiation sterilization technology**

263 Radiation is categorized into natural and artificial types, as illustrated in Figure 4. Artificial radiation
264 includes alpha (α), beta (β), and gamma (γ) rays emitted from radioactive isotopes, mechanically
265 generated X-rays, electron beams generated by electron accelerators, and neutron rays produced by
266 nuclear reactors (Shahi et al., 2021). Ionizing radiation, which has the property of ionizing the atoms
267 or molecules of a substance through which it passes, results in the formation of ions. This category
268 includes alpha rays, gamma rays, electron beams, ultraviolet rays, and neutrons (Akhila et al., 2021).

269 Food irradiation has been consistently raising safety concerns, and in response, relevant international
270 organizations such as FAO (Food and Agriculture Organization), IAEA (International Atomic Energy
271 Agency), and WHO (World Health Organization) have demonstrated the safety of irradiating food
272 with less than 10 kGy of gamma rays, X-rays, electron beams, and ultraviolet rays (UV)
273 (Venugopalan and Suprasanna, 2022). This is because radiation leaves no chemical residues and is
274 largely unaffected by environmental conditions (Jildeh et al., 2021). Gamma rays and X-rays, known
275 for their strong penetrative power, allow for the treatment of opaque samples or packaged finished
276 products, which helps prevent secondary contamination that could occur during repackaging after
277 sterilization (Mshelia et al., 2023).

278 Gamma rays are emitted radially around the radiation nucleus, while X-rays and electron beams
279 involve the use of electricity to generate artificially accelerated electrons, focusing on sterilizing
280 concentrated areas. In contrast, UV and electron beams, which have weaker penetrating power than
281 gamma and X-ray rays, can be precisely controlled via a power source, offering advantages in terms
282 of process control, rapidity, accuracy, and efficiency. Among the ultraviolet spectrum, UV is
283 categorized into UVA (315–400 nm), UVB (280–315 nm), and UVC (200–280 nm), with UVC being
284 the type used for microbial sterilization (Shahi et al., 2021).

285 These radiation types are employed in various applications, such as inhibiting the germination and
286 rooting of agricultural products, sterilizing pathogens and parasites, controlling the ripening of
287 agricultural products, and extending their shelf life.

288 The use of radiation in the food industry began in 1895 with Roentgen's discovery of X-rays and the
289 subsequent recognition of their potential to sterilize microorganisms in food. In 1921, radiation was
290 first utilized in the United States to sterilize trichinosis in pork (Jildeh et al., 2021). Initially, the high
291 cost of radiation equipment limited its practical use. However, by 1950, the mass production of
292 radioactive materials facilitated comprehensive research (Demir et al., 2019). Radiation sterilizes food
293 either directly by damaging microbial nucleic acids, which inhibits nucleic acid synthesis and
294 microbial division, or indirectly. Indirect sterilization occurs when radiation generates decomposition
295 products such as free radicals in water molecules; these highly reactive radicals combine with oxygen
296 to form oxidizing agents, which damage microbial cell membranes and thus sterilize the
297 microorganisms (Li and Farid, 2016).

298 Given the varying susceptibility of microorganisms to radiation, its application is categorized into
299 complete sterilization, pathogenic microorganism sterilization, and partial sterilization (Rosario et al.,
300 2021). Complete sterilization involves high-dose radiation (30–50 kGy) targeting endospore-
301 producing microorganisms such as *Bacillus* spp. and *Clostridium* spp. This level of radiation is
302 employed for foods requiring high safety standards such as canned food, hospital food, space food,
303 sports food, and aseptic feeds for laboratory animals. Pathogenic microorganism sterilization, which
304 uses a radiation dose of 1 to 10 kGy, targets harmful microorganisms in food. Partial sterilization aims
305 to reduce the microbial load in food with radiation doses ranging from 0.5 to 10 kGy to extend its
306 shelf life and refrigeration period.

307 The advantages of radiation include shorter processing times compared to heating, no chemical
308 residues, and easier handling (Yang et al., 2023). However, certain radiation doses can lead to
309 undesirable changes such as discoloration and softening of fruits, oxidation of meat products, and
310 consumer resistance due to concerns about radiation exposure (Ahn et al., 2013; Shahbaz et al., 2016).
311 Therefore, determining the appropriate type and dose of radiation for meat-based pet food is crucial.

312

313 **Setting the pet food radiation sterilization standards**

314 Table 3 presents a prior study on radiation sterilization technology applied to meat and fish products,
315 which are primary sources of animal proteins in pet food. It demonstrates that gamma rays and
316 electron beams below 10 kGy are the most prevalent forms of radiation used for the sterilization of
317 meat and fish, achieving general sterilization at doses above 6 kGy across all cultures. However,
318 doses around 3 kGy were found to produce excessive peroxides and volatiles, leading to off-flavors
319 and other issues affecting palatability (Farkas, 1998). Consequently, research is being conducted on
320 the impact of radiation on samples supplemented with natural or synthetic antioxidants.

321 The inclusion of guava leaf extract, which contains antioxidants, has shown to inhibit aerobic
322 microorganisms and *E. coli* even at a dose of 2 kGy, indicating that natural antioxidants maintain their
323 activity post-radiation (Sadiq et al., 2023). Furthermore, synthetic antioxidants such as inorganic
324 pyrophosphate, cinnamaldehyde, and ascorbic acid have demonstrated significant inhibitory effects
325 against mesophilic and thermophilic microorganisms, with antibacterial activity ranking in the order
326 of inorganic pyrophosphate, cinnamaldehyde, and ascorbic acid (Ayari et al., 2016).

327 Additionally, antioxidants like tea polyphenols, grape seed extract, D-sodium erythorbate, chitosan,
328 carvacrol, and mangosteen pulp have proven effective in reducing microbial presence during radiation
329 (Chen et al., 2023 ; Fitrianto et al., 2024 ; Hu et al., 2021). It is suggested that incorporating these
330 antioxidants could reduce the necessary radiation dose for microorganisms with high radiation
331 resistance or for finished products that require the prevention of external contamination factors during
332 consumption.

333 Harmful microorganisms that form resistant spores or are highly resistant to radiation require
334 treatment at higher doses than common harmful microorganisms (Lv et al., 2021). Notably, *Bacillus*
335 *Cereus*, *Proteus Mirabilis*, and *Enterococcus Faecalis* exhibit D_{10} -values for gamma rays and electron
336 beams that are approximately twice as high as those for general harmful microorganisms such as *E.*
337 *coli* and *Staphylococcus* spp. This differential resistance should be taken into account when selecting
338 the appropriate disinfection radiation dose (de Lara et al., 2002 ; Ebrahim et al., 2022). Moreover, a

339 radiation dose of 45 kGy has been found adequate for sterilizing *Clostridium* spp. in various meat
340 products, including smoked turkey, chicken fillet, chicken luncheon, beef luncheon, ground meat, and
341 raw sausage (Ebrahim et al., 2022).

342 It is recommended that radiation sterilization treatment of pet food raw ingredients should be
343 conducted within the maximum allowable dose of 10 kGy. Furthermore, quality and sensory
344 properties studies should be performed for each type of raw ingredient to ensure safety and
345 acceptability. For finished pet food products, it is necessary to conduct quality and sensory properties
346 studies to determine the appropriate radiation sterilization dose, particularly for microorganisms with
347 high contamination potential, considering the shape and distribution form of the product, such as
348 pouches or cans.

350 Conclusion

351 The commercialization of non-thermal sterilization technology in the food industry is actively
352 underway, and introducing non-thermal sterilization for pet food is considered highly valuable as the
353 culture of pet humanization spreads. The main non-thermal sterilization technologies are HPP, plasma,
354 and radiation. Unlike traditional sterilization methods such as chemical and heat sterilization, these non-
355 thermal methods offer quality and sensory advantages, including leaving no residue on food,
356 minimizing deformation of raw ingredients, and preserving nutrients.

357 Autoclaving involves the application of high-pressure to raw ingredients or finished products,
358 ensuring even sterilization across the entire surface of the sample, regardless of its shape, nature, or
359 packaging status. However, protein gel formation and color changes can occur at pressures of 200-400
360 MPa, making it crucial to identify pressures that effectively sterilize microbes at lower pressures.
361 Therefore, it is necessary to study the sterilization efficiency and quality parameters such as color, water
362 retention, shear force, and sensory properties like flavor and panel evaluation between raw and finished
363 pet food products.

364 Plasma technology applies plasma sterilization to sterilize the surface of raw ingredients or finished

365 products. Dielectric barrier plasma technology, which uses dielectrics to expand the usable range and
366 lower the treatment temperature, is primarily studied. However, dielectric barrier plasma is less
367 economical than PAW. Additionally, PAW can impart a sour taste depending on the type of treated water,
368 necessitating a comparison of the efficiency of both methods. Consequently, studying the quality and
369 sensory properties of pet food according to various plasma methods is essential.

370 Radiation involves penetrating raw ingredients or finished products with radiation and can be
371 conducted while the sample is packaged. However, certain radiation doses may cause discoloration and
372 off-flavors, requiring pet sensory studies. Therefore, selecting the appropriate radiation dose and
373 conducting quality and sensory properties studies for various ingredients and finished products is crucial
374 in the radiation treatment of pet food.

ACCEPTED

375 Conflicts of interest

376 The authors declare no potential conflicts of interest.

377

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387 Ethics Approval

388 This article does not require IRB/IACUC approval because there are no human and animal participants.

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656 **Table 1.** Summary of high-pressure processing standards for meat and fish raw ingredients commonly used as pet food ingredients.

Raw ingredients	Processing conditions	Experimental results	Reference
Chicken (chicken leg)	0.1, 300, 600 MPa (15±3°C, 5 min)	<ul style="list-style-type: none"> ■ Total microorganisms were reduced to undetectable levels when treated at 300 and 600 MPa. ■ When treated at 300 and 600 MPa, total microorganisms decreased by 3 and 4 log CFU/g, respectively, compared to the control after 7 days of storage. ■ Treatment at 300 and 600 MPa increased lightness and decreased redness and yellowness. ■ Treatment at 300 MPa had no effect on TBARS and protein oxidation. ■ The addition of phosvitin during ultra-high pressure treatment increased TBARS, protein oxidation, and microbial suppression. 	Jung et al. (2012)
	Total microorganisms (TSA, 37°C, 48 h)		
	Storage conditions 0, 3, 7 days		
Chicken (chicken breast)	500 MPa (18-20°C, 10 min)	<ul style="list-style-type: none"> ■ <i>Brochothrix Thermosphacta</i>, <i>Pseudomonas</i> spp., enterobacteriaceae, <i>Lactobacillus</i>, and yeasts and molds were detected at 1 log CFU/g when treated at 500 MPa at 4 and 12°C. ■ The pH increased compared to the control when treated at 500 MPa. 	Argyri et al. (2018)
	Total microorganisms (PCA, 30°C, 48-72 h)		
	<i>Lactobacillus</i> (MRS, 30°C, 48-72 h)		
	Yeast and mold (DRBC, 25°C, 48-72 h)		
	Gut bacteria (VRBG, 37°C, 24 h)		
	<i>Pseudomonas</i> spp. (pseudomonas agar base, 25°C, 48 h)		
<i>Salmonella Enteritidis</i> (XLD, 37°C, 16-18 h)			
Storage conditions 0, 5, 10 days 4, 12°C			

Pork	<p>50, 100, 200, 300, 400, 500 MPa (15 min)</p> <hr/> <p>Total microorganisms (NA, 30°C, 48 h)</p>	<ul style="list-style-type: none"> ■ Total microorganisms decreased to less than 3 log CFU/g when treated at over 400 MPa. ■ Total microorganisms of pork treated at 500 MPa for 1, 5, and 15 minutes, decreased from approximately 3 log CFU/g in the control to 2.17, 1.77, and 2.33 log CFU/g, respectively. ■ The pH of the pork increased with higher treatment pressure and longer treatment time compared to the control. When treated at 500 MPa for 15 minutes, the pH was approximately 5.96, which was higher than the control (approximately pH 5.4) and other treatment groups (approximately pH 5.6-5.9). 	Sazonova et al. (2017).
Pork Fuji (cooked O) Smoked Pork Loin (Cooked X)	<p>300, 400, 500, 600 MPa (10 to 60 min)</p> <hr/> <p><i>Escherichia Coli</i> <i>Enterococci</i> Mesophilic and thermophilic bacteria Eosinophils</p> <hr/> <p>Storage conditions 0, 4, 6, 8 week 4 to 6°C</p>	<ul style="list-style-type: none"> ■ Cooked pork traditional ham treated at 600 MPa for 10 minutes showed no detection of coliform, <i>Enterococci</i>, or acidophilic bacteria during the 8-week storage period. ■ Cooked pork traditional ham treated at 300 MPa or higher for more than 10 minutes showed increased redness, yellowness, and shear force, and a decrease in lightness compared to the control. 	Karlowski et al. (2002)
Beef (breast, pectoral)	<p>400, 600 MPa (35, 45, 55°C, 20 min)</p> <hr/> <p>Total microorganisms (PCA, 30°C, 72 h) <i>Lactobacillus</i> (MRS, 37°C, 24 h) Gut bacteria (VRBG, 30°C, 24 h) <i>Listeria</i> <i>Salmonella</i> <i>Campylobacter</i></p> <hr/> <p>Storage conditions 0, 7, 15, 30 days</p>	<ul style="list-style-type: none"> ■ Beef treated at 400 MPa or higher at 35°C showed increased pH, lightness, yellowness, and cooking loss, and a decrease in redness. ■ Cooking loss increased as the treatment pressure increased. 	McArdle et al. (2011)

Sliced beef	<p>300, 450, 600 MPa (5 min)</p> <hr/> <p><i>Listeria Innocua</i>, (Oxford ager, 37°C, 48 h) <i>Enterococcus Faecium</i> (Enterococcosel agar, 37°C, 48 h)</p>	<ul style="list-style-type: none"> ■ Treatment with pressures of 300, 450, and 600 MPa, including 2% citric acid and 2% salt solution, reduced the growth of <i>Enterococcus Faecium</i> by 1, 4, and 6 log cycles, respectively. Increasing the citric acid content was effective in inhibiting <i>Enterococcus Faecium</i>. ■ Treatment with pressures of 300-600 MPa, including 2% citric acid and 2% salt solution, reduced the growth of <i>Listeria Innocua</i> by 6 log cycles. Increasing the salt concentration was effective in inhibiting <i>Listeria Innocua</i>. ■ <i>Listeria Innocua</i> is more sensitive to high-pressure treatment than <i>Enterococcus Faecium</i>. When treated at 450 MPa with a solution containing 1% salt and 2% citric acid, <i>Listeria Innocua</i> was reduced by 7 log cycles. 	Rodrigues et al. (2016)
Salmon, cod, mackerel	<p>0.1, 200, 500 MPa (2 min)</p> <hr/> <p>Thermophilic microorganisms (Long & Hammer agar, 15°C, 5-7 days) Hydrogen sulfide-producing microorganisms (Iron ager, 20°C, 3-4 day)</p> <hr/> <p>Storage conditions Cod: 0, 7, 11, 14, 22 days Salmon: 0, 7, 11, 15, 18, 21, 26 days Mackerel: 0, 4, 7, 11, 14, 19 days</p>	<ul style="list-style-type: none"> ■ Cod and mackerel treated at 500 MPa showed total aerobic bacteria reduced to undetectable levels. ■ All treatment groups showed an increase in TBARS and pH compared to the control when treated at 200 MPa or higher. 	Rode and Hovda (2016)

Table 2. Summary of plasma sterilization standards for meat and fish raw ingredients commonly used as pet food ingredients.

Raw ingredients	Processing conditions	Experimental results	Reference
Chicken (chicken breast)	<p>Normal pressure plasma (10 sec, 30 sec, 1 min, 2 min, 3 min) Gap: 1.0-1.5 cm Voltage: 6.0~11.0 kV Frequency: 23.0 to 38.5 kHz He: 5.0 L/min O₂: 50, 100 mL/min</p> <hr/> <p>Total microorganisms (TSA, 30°C, 24 h) <i>Listeria Innocua</i> (TSB, 30°C, 48 h)</p>	<ul style="list-style-type: none"> ■ The thicker and rougher the surface of the sample, the less effective the plasma treatment. ■ The sterilization of <i>Listeria Innocua</i> on skinless chicken breast has been reported with a gap of 1 cm, voltage of 8-11 kV, frequency of 30-38.5 kHz, and O₂ flow rate of 0-25 mL/min for 4 minutes. ■ The sterilization of <i>Listeria Innocua</i> on chicken breast with skin has been reported with a gap of 1 cm, voltage of 7 kV, frequency of 38.5 kHz for 3 minutes, and with a gap of 1 cm, voltage of 8 kV, frequency of 38.5 kHz for 30 seconds. ■ Voltages above 9 kV caused overheating of the transformer, and stable operation was achieved at a frequency of 23 kHz. 	Noriega et al. (2011)
Chicken/Pork (chicken breast/pork belly)	<p>Plasma-activated water (immersion, 25.5°C, 15 min) Voltage: 125 W, 15 kV Frequency: 50 Hz 60 ppm H₂O₂, 500 mL</p> <hr/> <p><i>Salmonella Typhimurium</i> (XLD, 37°C, 24 h) <i>Campylobacter Jejuni</i> (mCCD, 41.5°C, 48 h) <i>Escherichia Coli</i> (ECD, 44°C, 24 h) <i>Staphylococcus Aureus</i> (BPA, 37°C, 48 h) <i>Pseudomonas Aeruginosa</i> (GSP, 30°C, 48 h)</p> <hr/> <p>Storage conditions 0, 3, 7, 10 days 4 to 6°C</p>	<ul style="list-style-type: none"> ■ Treatment of pork and chicken with hydrogen peroxide PAW (plasma-activated water) resulted in a greater reduction of <i>Campylobacter Jejuni</i> compared to other microorganisms. ■ Hydrogen peroxide PAW treatment on the surface of pork resulted in an increase in pH, lightness, temperature, and water activity, and a decrease in redness and yellowness. ■ Hydrogen peroxide PAW treatment on the surface of chicken resulted in an increase in pH and redness, and a decrease in lightness, yellowness, and temperature. 	Sammanee et al. (2022)

Pigs (Sirloin)	Vacuum Plasma (5, 10 min) Frequency: 20-100 kHz	<ul style="list-style-type: none"> ■ When treated with helium gas and plasma, total microorganisms, psychrophilic bacteria, yeasts, and molds were detected at 2.49, 2.56, and 1.35 log CFU/cm², respectively. ■ When treated with argon gas and plasma, total microorganisms, psychrophilic bacteria, yeasts, and molds were detected at 3.13, 3.13, and 1.86 log CFU/cm², respectively. ■ When treated with nitrogen gas and plasma, total microorganisms, psychrophilic bacteria, yeasts, and molds were detected at 5.02, 5.09, and 3.37 log CFU/cm², respectively. ■ pH showed no significant effect when treated with vacuum plasma. 	Ulbin-Figlewicz et al. (2015)
Beef (10g)	Plasma-activated water (1mL spray, 0, 5, 10, 20, 30 min) Voltage: 10 kV Frequency: 8 kHz 40 mL deionized water	<ul style="list-style-type: none"> ■ A 5 minute treatment with PAW resulted in a pH decrease of 3.7 compared to the control. ■ Longer treatment with PAW led to an increase in VBN and TBARS. ■ PAW treatment resulted in a decrease in redness after 4 days of storage. 	Zhao et al. (2018)
Mackerel (fillet)	Plasma-activated water (immersion, 25°C, 10 min) Voltage: 550 W Frequency: 25 kHz 200 ppm CH ₃ COOH 30 mL	<ul style="list-style-type: none"> ■ A 10 minute treatment with plasma-activated peracetic acid (PA-PAA) resulted in a 3.8 log CFU/g reduction in total microorganisms. ■ PA-PAA treatment had no significant effect on color and TBARS. 	Zhao et al. (2021)

Total microorganisms (TMC/TPC)

(30°C, 48 h/6.5°C, 10 day)

Escherichia Coli

(MacConkey, 37°C, 24 h)

Listeria Innocua

(Oxford, 37°C, 24 h)

Pseudomonas Fluorescens

(CFN, 30°C, 24 h)

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Raw ingredients	Processing conditions	Experimental results	Reference
Chicken (Machine-boned chicken)	Gamma rays (0.30, 0.60, 0.90, 1.20, 1.80, 2.70, 3.60 kGy) <i>Salmonella Typhimurium</i> (TSA, 35°C, 24 h)	<ul style="list-style-type: none"> ■ Gamma irradiation treatment at 20°C exhibited a higher inhibitory effect on <i>Salmonella Typhimurium</i> compared to -20°C and 0°C. ■ Atmospheric pressure conditions showed a higher inhibitory effect on <i>Salmonella Typhimurium</i> compared to vacuum conditions. ■ When irradiated with 3.6 kGy at 20°C, <i>Salmonella</i> was detected at 2.75 log CFU/g under atmospheric pressure conditions and 3.06 log CFU/g under vacuum conditions. ■ When irradiated with 3.0 kGy at -20°C, <i>Salmonella</i> was detected at 3.93 log CFU/g under atmospheric pressure conditions and 4.29 log CFU/g under vacuum conditions. 	Thayer and Boyd (1991)
Chicken (chicken patty) Guava Leaf Extract	Gamma rays (2 kGy) Total microorganisms (PCA) Storage conditions 0, 5, 10 days	<ul style="list-style-type: none"> ■ Under atmospheric pressure conditions, TPC, DPPH, and FRAP were lower, while TVBN, TBARS, POV, total anaerobic bacteria, and fecal coliforms were higher compared to vacuum conditions. ■ As the concentration of guava leaf extract increased, the detection of total anaerobic bacteria and fecal coliforms decreased. ■ Treatment with 2 kGy resulted in an increase in TPC, DPPH, FRAP, lightness, redness, and yellowness compared to the control, while TVBN, TBARS, and POV decreased. 	Sadiq et al. (2023)
Pork (ground pork, 30 g)	Gamma rays (5 kGy) <i>Pseudomonas Aeruginosa</i> (NA, 37°C, 48 h) <i>Lactobacillus Casei</i> (MRS, 37°C, 48 h)	<ul style="list-style-type: none"> ■ The control treated with 5 kGy showed no detection of <i>Pseudomonas Aeruginosa</i> and <i>Lactobacillus Casei</i>. ■ After irradiation treatment and inoculation with <i>Pseudomonas Aeruginosa</i>, there was an increase in pH compared to the control. ■ After irradiation treatment and inoculation with <i>Lactobacillus Casei</i>, there was a decrease in pH compared to the control. 	Kim et al. (2004)

Pork (minced pork, bologna sausage)	Electron beam (2.5 MeV, 2, 4, 6, 8, 10 kGy) X-Ray (5 MeV, 2, 4, 6, 8, 10 kGy)	<ul style="list-style-type: none"> ■ Compared to electron beams, X-rays showed lower protein solubility, detection of total anaerobic bacteria, lightness, and redness, but higher TBARS, yellowness, and cooking yield. ■ Treatment with 2 kGy or more resulted in increased collagen solubility, TBARS, redness, and yellowness, and decreased detection of total anaerobic bacteria and lightness. ■ In fresh pork, treatment with 2-6 kGy using electron beams reduced the detection of total anaerobic bacteria by 1.69-2.88 log CFU/g, and no detection was observed from 8 kGy onwards. ■ In fresh pork, treatment with 2-6 kGy using X-rays reduced the detection of total anaerobic bacteria by 1.99-2.94 log CFU/g, and no detection was observed from 8 kGy onwards. 	Shin et al. (2014)
Beef (ground beef) Antioxidants (cinnamaldehyde , ascorbic acid, Pyrophosphate inorganic)	<p>Gamma rays (2 kGy)</p> <hr/> <p>Mesophilic bacteria (PCA, 30°C, 3 day) Total coliforms (VRBA, 30°C, 24 h) Isolated <i>Escherichia Coli</i> (VRBA, 44°C, 24 h) Yeast and mold (SD, 25°C, 3-5 day) <i>Staphylococcus Aureus</i> (MSA ,35°C, 24-48 h) <i>Listeria Monocytogenes</i> (PALCAM ,35°C, 24-48 h) <i>Bacillus Cereus</i> (EE, 30°C, 24 h)</p> <hr/> <p>Storage conditions 0, 7, 12, 21 days</p>	<ul style="list-style-type: none"> ■ Treatment with 2 kGy resulted in a reduction of mesophilic bacteria, psychrotrophic bacteria, and yeasts and molds by 2.58, 3.76, and 1.32 log CFU/g, respectively. ■ Treatment with 2 kGy showed no significant difference in pH and general components, but there was a decrease in heme iron content and an increase in TBARS, peroxide, lightness, and redness. 	Ayari et al. (2016)
Herring (canned)	Gamma rays (1.5, 3, 6 kGy) Electromagnetic radiation, X-rays (4.8 MeV, 1.5, 3, 6 kGy)	<ul style="list-style-type: none"> ■ Gamma irradiation and electron beam treatment at 6 kGy resulted in the detection of microbes at approximately 1 log CFU/g. 	Sanzharova et al. (2021)

Mesophilic bacteria
Conditions Anaerobes
(PCA, 32°C)
Salmonella spp.
(Bismuth sulphite agar)
Total coliforms
(MacConkey agar)
Listeria spp.
(Oxford agar)
Staphylococci spp.
(Chapman Stone agar)

Storage conditions
15, 30 days

- X-ray treatment at 3-6 kGy resulted in detection of microbes at approximately 1 log CFU/g.
- Treatment at 6 kGy resulted in the generation of a 'smoking' aroma.

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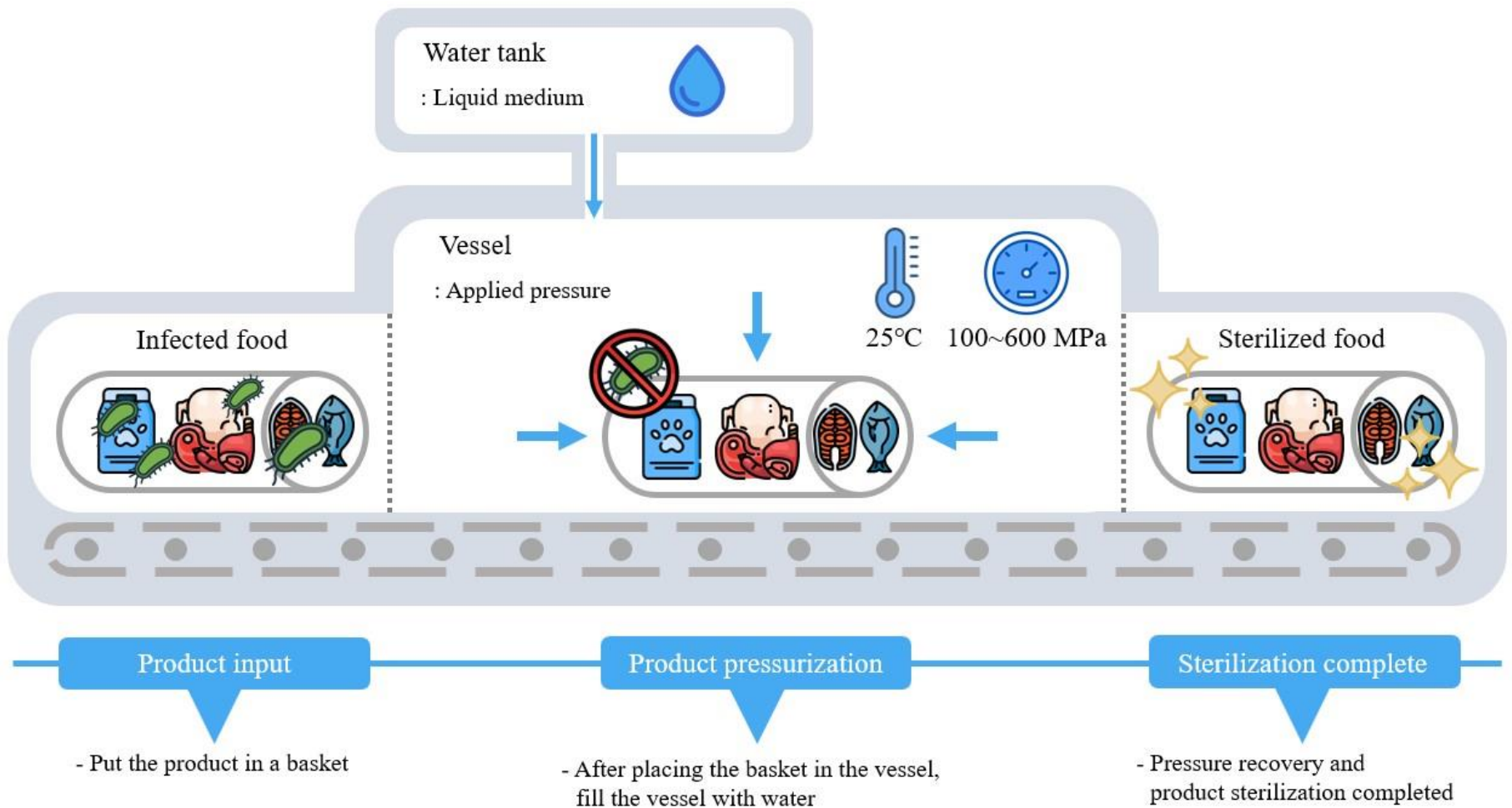


Fig 1. High-pressure processing equipment and principles.

Energy level

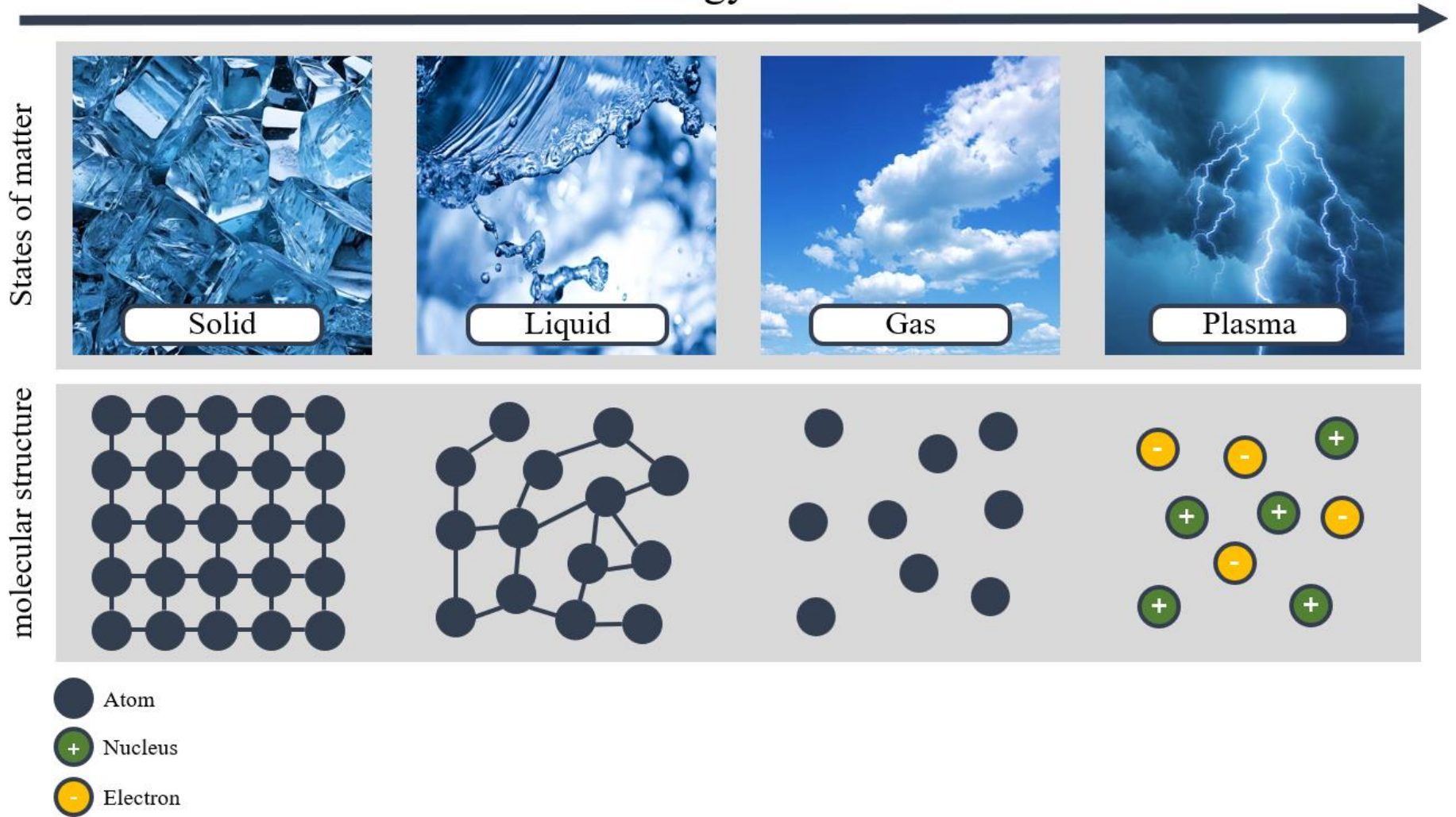
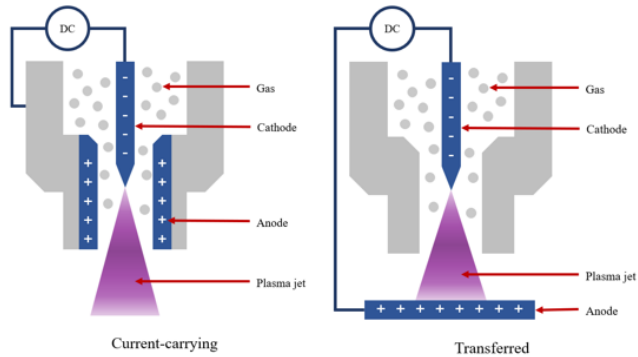


Fig 2. Plasma, is the fourth form of matter.

Local thermodynamic equilibrium plasma

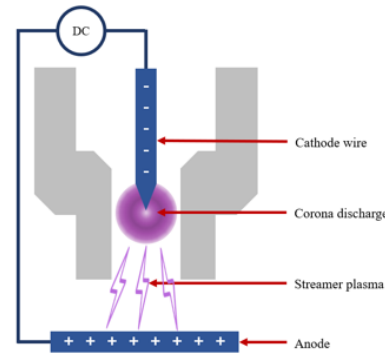
Arc plasma



- Conditions: atmospheric pressure, gas flow
- Use direct current (DC) electrodes
- Plasma treatment area is small and high temperature is generated
- Applied to welders and cutters

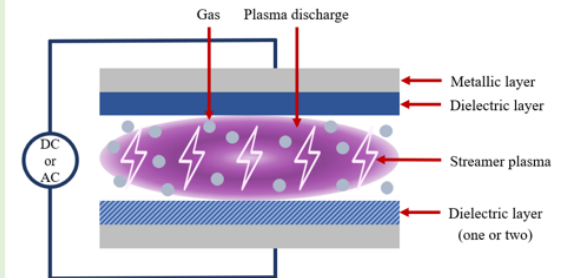
Non-local thermodynamic equilibrium plasma

Corona discharge



- Conditions: atmospheric pressure
- Use direct current (DC) electrodes
- Plasma treatment area is small
- Applied to air purification and sterilization

Dielectric barrier discharge



- Conditions: Available from low to high pressure
- Use alternating current (AC) electrodes
- Plasma treatment area is large
- Applied to pollutant removal and sterilization

Fig 3. Different plasma generation methods and their characteristics.

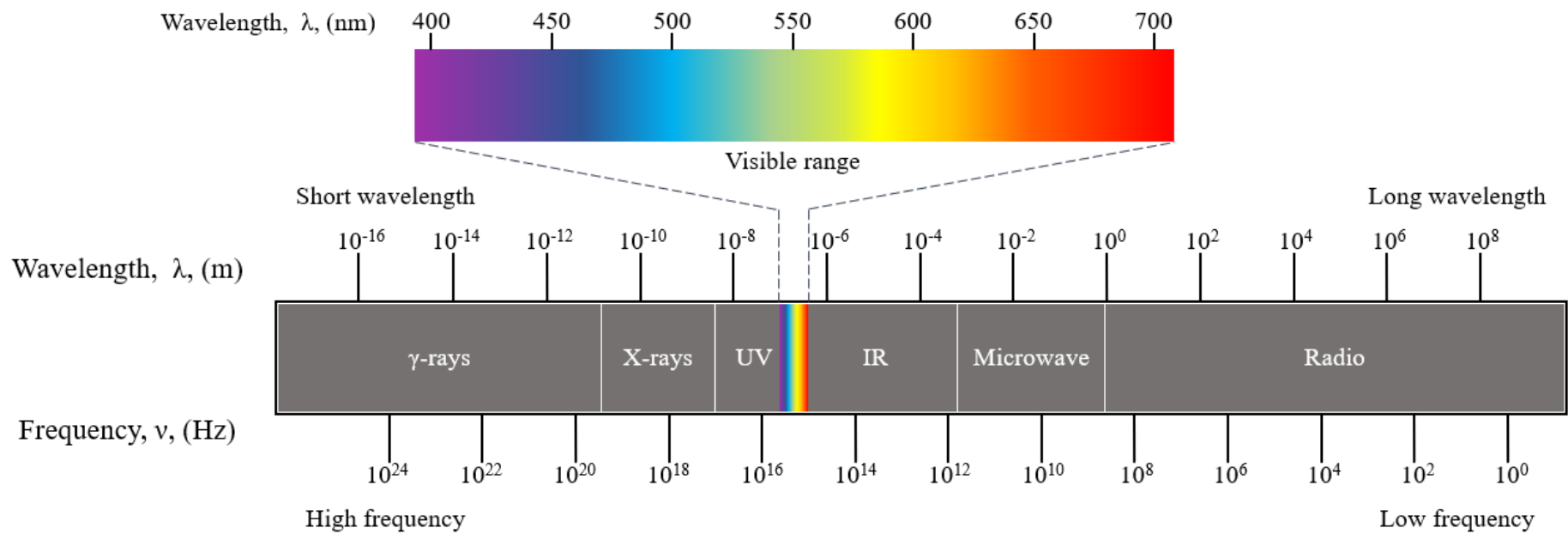


Fig 4. Types and extent of radiation.