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<b>Article Type</b>	Review article
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9 **Abstract**

10 Pork belly is one of the most valuable primal cuts of pork with high preferences. Although  
11 meat quality is becoming increasingly important, defining pork belly quality is challenging  
12 owing to the structure and diversity of the preferred characteristics. This study identified the  
13 factors influencing pork belly quality traits through a literature review. In total, 55 articles  
14 related to pork belly quality were selected and summarized. The quality traits of pork belly  
15 are considered to be various factors, including belly yield (weight, length, thickness, etc.),  
16 firmness, fatty acid composition, color, and sensory properties. The quality of pork belly is  
17 influenced by various factors, such as sex, genetic parameters, carcass weight, and diet. A  
18 more diverse approach is required to comprehensively understand the quality traits and impact  
19 factors of pork bellies.

20

21 **Keywords:** pork, pork belly, quality, endogenous factor, exogenous factor

22

## 23 **Introduction**

24 Meat is an important source of protein and several essential nutrients in the human diet.  
25 Pork is one of the most preferred meats worldwide and its consumption has steadily increased  
26 (Godfray et al., 2018; Jeon et al., 2024). Among the primal cuts of pork, the pork belly is one  
27 of the most valuable cuts that is preferred in many countries (Jo et al., 2022; Jeong et al.,  
28 2024).

29 Consumer interest in food safety, quality, and healthy diets is increasing, and these  
30 changes influence meat consumption and meat industry. While factors, such as individual  
31 income and product price, are expected to have less impact on meat consumption, the  
32 importance of meat quality is anticipated to increase (Henchion et al., 2014; Kim et al., 2023).  
33 Pork belly is composed of multiple muscles and intermuscular fat layers, making it more  
34 challenging to assess meat quality than single-muscle cuts such as pork loin (Jo et al., 2024).  
35 To determine the quality of pork belly, the quality of both the muscle and fat layers must be  
36 considered. Additionally, pork bellies are consumed differently in different countries. In  
37 Western countries, pork belly is primarily consumed as bacon after curing, whereas in some  
38 countries, such as South Korea, consumers prefer grilled pork belly (Choe et al., 2015; Kang  
39 et al., 2015). These varying consumption preferences lead to different expectations regarding  
40 the quality of pork belly. Therefore, to prepare pork belly that satisfies consumer preferences,  
41 it is necessary to understand the quality traits of pork belly and factors that influence them.  
42 This study systematically reviewed and summarized previously published literature on pork  
43 belly quality. In particular, we reviewed only the fresh pork belly quality, excluding  
44 processing effects such as curing, aging, and freezing. Therefore, this review aimed to clarify  
45 the quality properties of pork belly and identify the factors affecting the quality properties.

46

## 47 **Literature selection**

48 This study aimed to systematically search and summarize the previous literature to  
49 identify the quality properties of fresh pork belly and the factors that influence belly quality.  
50 The literature was selected following the Preferred Reporting Items for Systematic Reviews  
51 and Meta-Analysis (PRISMA) guidelines (Page et al., 2021). The search was conducted using  
52 the Web of Science and SCOPUS databases with no restrictions on the year of publication.

53 We used a combination of the terms ‘meat’, ‘quality’, ‘pork’, and ‘belly’ to search the  
54 literature in Web of Science and SCOPUS. The criteria for selecting studies were as follows:  
55 (1) written in English and published in journals and (2) research studies on the quality of fresh  
56 pork belly or influencing factor to belly quality. The pork belly is a major cut of pork carcass;  
57 therefore, it is common to analyze belly quality together with other cuts to describe the overall  
58 quality of pork carcasses. Among these studies, we selected those that allocated at least one  
59 section to the description of pork belly quality to select literature with sufficient consideration  
60 for pork belly quality. In addition, studies that analyzed the quality of fresh pork belly after  
61 processing, such as heating, storage, and aging, were excluded.

62 A total of 735 studies were obtained from the literature search of the database, and some  
63 studies were excluded based on the process of selection and eligibility evaluation (Fig 1). A  
64 total of 150 studies were excluded due to duplication. Studies were excluded if they did not fit  
65 the topic based on the title and abstract, or if the full text was unavailable. The remaining 184  
66 articles were reviewed in full and those meeting the above-mentioned criteria were excluded.  
67 Finally, 55 articles were selected, and the key data were summarized and organized in this  
68 review.

69

## 70 **Quality traits of fresh pork belly**

71 The pork belly is a cut obtained from the central part after removing the shoulder, leg,  
72 and loin of the half carcass. The cut of pork belly from the carcass varies from countries. In

73 South Korea, pork belly is defined as the abdominal muscle from the 5<sup>th</sup> or 6<sup>th</sup> rib to the hind  
74 leg (the 7<sup>th</sup> lumbar vertebrae) with the loin removed. The United Nations Economic  
75 Commission for Europe (UNECE) and the United States Department of Agriculture (USDA)  
76 specify that the pork belly contains 10 to 13 ribs (USDA requiring a minimum of 11 ribs),  
77 depending on the extent of shoulder part removed, and be square or rectangular in shape with  
78 neither side of the belly more than 5 cm longer than the opposite side (UNECE, 2006; USDA,  
79 2014). The pork belly consists of multiple muscle and fat layers, requiring a comprehensive  
80 assessment of both muscle and fat conditions to evaluate the overall belly quality. We  
81 identified and classified the quality traits of the pork belly reported in the selected studies (Fig  
82 2).

83 The most frequently measured quality trait of pork belly was the pork belly yield from  
84 carcasses. This trait was investigated in 46 of the 55 articles. Pork belly yield includes the  
85 weight of the belly, its proportion within the carcass, dimensions (length, width, and  
86 thickness), and the muscle-to-fat ratio . The pork belly yield is an important commercial  
87 attribute that leads to profits from the producer's perspective (Choe et al., 2015). Processors  
88 prefer heavy and thick pork bellies because of their higher processing yields (Soladoye et al.,  
89 2015). However, increased belly weight is generally associated with higher fat content  
90 (Albano-Gaglio et al., 2024; Hoa et al., 2021) which consumers may not prefer, given the  
91 growing concerns about high-fat diets. Therefore, there is a need to achieve a balance between  
92 producer profitability and consumer preferences regarding pork belly yield.

93 Firmness was the second most commonly evaluated quality trait, as referenced in 37 of  
94 55 articles. The firmness of pork belly is an important property that influences its processing  
95 efficiency, yield, and consumer acceptability. It has been reported that soft pork belly is  
96 difficult to process, has a poor appearance owing to the separation of fat layers, and has a  
97 short shelf life with low oxidation stability (Soladoye et al., 2017; Zomeño et al., 2024).

98 Firmness is affected by multiple factors, including the dimensions, thickness, fat content, and  
99 fat saturation of pork belly (Soladoye et al., 2017). The methods used to assess belly firmness  
100 include flop distance and angle analysis, instrumental texture measurement, and finger-press  
101 firmness. Flop distance and angle analyses measure the distance between two dropped  
102 endpoints and the angle at the bend point by placing the pork belly on a horizontal bar with  
103 the skin side up or down (Font-i-Furnols et al., 2023). A greater flop distance and angle  
104 indicated a firmer pork belly. Instrumental texture measurement determines firmness by  
105 measuring the force required to compress the central part of the pork belly using a texture  
106 analyzer (Apple et al., 2011; Font-i-Furnols et al., 2023). Finger-press firmness evaluates  
107 belly firmness on a 5-point scale by applying pressure on the belly with a finger and assessing  
108 the degree to which the finger press mark remains (Soladoye et al., 2017; Zomeño et al.,  
109 2024).

110 Fatty acid composition analysis was also extensively performed to determine pork belly  
111 quality, representing 52.73% of the selected articles. The fatty acid composition includes the  
112 proportions of saturated fatty acids (SFA), such as palmitic acid and steric acid;  
113 monounsaturated fatty acids (MUFA), such as oleic acid; polyunsaturated fatty acids (PUFA),  
114 such as linoleic acid, linolenic acid, and arachidonic acid; PUFA/SFA ratio; and n-6/n-3 ratio.  
115 Fatty acid composition is affected by various factors such as sex, growth rate, and diet  
116 (Browne et al., 2013; Correa et al., 2008; Kellenr et al., 2015). The fatty acid composition  
117 reflects fat accumulation, with higher growth rates and greater fat accumulation leading to  
118 increased fat saturation (Corre et al., 2008). In general, the fatty acid composition of fresh  
119 pork belly was reported to be a PUFA/SFA ratio of 0.48 and an n-6/n-3 ratio of 17.98 (Choe  
120 et al., 2015). However, many countries, including the United States and Europe, recommend  
121 decreasing the consumption of SFA and the n-6/n-3 ratio and increasing the PUFA/SFA ratio  
122 in the diet (Choe et al., 2015; Soladoye et al., 2017). In the United Kingdom, the PUFA/SFA

123 ratio and n-6/n-3 ratio have been recommended to be greater than 0.4 and less than 4.0  
124 respectively (Soladoye et al., 2017). Therefore, it is important to produce pork bellies with a  
125 balanced fatty acid composition considering both belly productivity and consumer health. The  
126 iodine value (IV), which reflects the degree of fat unsaturation, has also been investigated as a  
127 quality trait in pork belly. IV is calculated as the amount of iodine bound of unsaturated fat  
128 because iodine react with the  $\pi$ -electrons of the double bonds (Gatlin et al., 2003). IV is  
129 associated with UFA content, including oleic acid, linoleic acid, linolenic acid, and pork belly  
130 firmness (Font-i-Furnols et al., 2023).

131 Other quality traits of pork belly include physicochemical properties that are typically  
132 measured to assess meat quality, such as color, pH, proximate composition, and water-holding  
133 capacity. Color is an important factor for consumers when judging meat quality at the point of  
134 purchase. Consumers use meat color as an indicator of freshness. Bright red meat and white  
135 fat are preferred as good quality meat (Font-i-Furnols & Guerrero, 2014; Hugo & Roodt,  
136 2007). Among the 55 studies, 13 confirmed the color of the pork belly. Some studies have  
137 confirmed the color of specific muscles, such as the rectus abdominis muscle, external  
138 abdominal oblique muscle, or fat, rather than the color of the entire surface of the meat  
139 because pork belly consists of layers of muscle and fat (Apple et al., 2007; Engel et al., 2001).  
140 Other studies have assessed the subjective visual color of the muscle fat in pork belly  
141 (Browne et al., 2013; Jeong et al., 2011). Additionally, physicochemical properties, such as  
142 pH and proximate composition, were measured by grinding the pork belly or measuring the  
143 pH in the muscle area using solid-state probes (Hoa et al., 2021; Hoa et al., 2023; Jeong et al.,  
144 2011). The pH of pork belly was measured in 5 studies, and the measured pork belly pH  
145 ranges from 5.70 to 5.95 (COSTA E SILVA et al., 2017; Hoa et al., 2023; Lim et al., 2013;  
146 Lim et al., 2014; Jeong et al., 2011). Other quality traits reported in the literature to confirm  
147 the oxidative stability of pork belly fat include cooking loss, volatile compound composition,



148 sensory properties, and malondialdehyde content (Albano-Gaglio et al., 2024; Hoa et al.,  
149 2021; Lim et al., 2013).

150

## 151 **Factors affecting the pork belly quality**

152 Meat quality is affected by multiple factors, including endogenous factors such as sex,  
153 genetic effect, and breed, as well as exogenous factors such as feeding and slaughter methods.  
154 Additionally, post-slaughter processes such as chilling, storage, and aging significantly  
155 impact meat quality. This review divided the factors affecting pork belly quality, as described  
156 in 55 articles, into endogenous and exogenous factors. To maintain a focus on the fresh pork  
157 belly quality, the effects of processing methods such as storage, freezing, and aging after  
158 slaughter were not addressed.

### 159 **Endogenous factor**

#### 160 **Sex effect and castration methods**

161 Animal sex affects various carcass properties such as weight, lean meat yield, and fat  
162 content, which can change the quality of pork belly. Several studies have investigated the  
163 effects of sex on pork belly quality (Table 1). Male pigs are generally castrated to prevent  
164 boar taint caused by androstenone and skatole and to reduce aggressive and sexual behavior,  
165 thereby improving growth performance (Prunier et al., 2006). Barrows (castrated male pig) are  
166 generally heavier than gilts, increasing the proportion of pork belly in the carcass (Bahelka et  
167 al., 2011; Correa et al., 2008; Duziński et al., 2015; Lee et al., 2013; Overholt et al., 2016;  
168 Stupka et al., 2004). Barrows also tend to have a higher fat deposition in the pork belly,  
169 whereas gilts have a higher proportion of lean meat in the belly (Bahelka et al., 2011; Stupka  
170 et al., 2004). These results were reported because barrows require less energy to deposition  
171 lean tissue than gilts and excess energy is accumulated as fat (Overholt et al., 2016).  
172 Additionally, the fatty acid composition of belly fat can be affected by sex (Correa et al.,

173 2008; Lee et al., 2013). Correa et al. (2008) found that compared to barrows, gilts had lower  
174 SFA content and higher linoleic acid and PUFA contents in belly fat. Lee et al. (2013)  
175 reported similar results, with barrows having a higher palmitic acid content, the SFA, and  
176 lower linoleic acid, the PUFA in belly fat than that in gilts. Overall, compared to barrows,  
177 gilts have a higher degree of belly fat unsaturation and a higher iodine value. Therefore,  
178 barrows produce firmer pork bellies with the higher proportion of SFA than the pork bellies  
179 of gilts.

180 Immunocastration is emerging as an alternative to traditional surgical castration, and the  
181 effects of different castration methods on pork belly quality have been widely studied (Costa e  
182 silva et al., 2017; Font-i-Furnols et al., 2023; Jeong et al., 2011; Kyle et al., 2014; Lowe et al.,  
183 2016; Tavárez et al., 2014). Most studies indicate that immunocastration produces softer pork  
184 belly with lower fat content, higher PUFA content, and higher iodine value than pork belly  
185 obtained from surgically castrated pigs (Costa et al., 2017; Font-i-Furnols et al., 2023; Kyle et  
186 al., 2014; Lowe et al., 2016). Jeong et al. (2011) reported the sensory properties of pork belly  
187 based on different castration methods. They reported that pork belly obtained from  
188 immunocastrated pigs had higher visual evaluation traits than the pork belly obtained from  
189 surgically castrated pig, although the former did not significantly affect taste, tenderness, and  
190 overall acceptability (Jeong et al., 2011). In addition, there have been studies on examining  
191 the effects of supplementing ractopamine hydrochloride (RAC) in combination with  
192 immunological castration (Costa e silva et al., 2017; Kyle et al., 2014; Lowe et al., 2016).  
193 RAC, a  $\beta$ -adrenergic agonist, is known to improve feed efficiency and increase lean meat  
194 content (Leick et al., 2010). Kyle et al. (2014) reported that immunologically castrated  
195 barrows fed a diet supplemented with RAC produced a wider and softer pork belly with a  
196 significantly higher percentage of PUFA in pbelly fat. In contrast, Costa et al. (2017) and  
197 Lowe et al. (2016) found no significant effects of RAC supplementation on pork belly quality

198 in immunocastrated barrows. Harris et al. (2018) and Tavárez et al. (2014) reported changes  
199 in pork belly quality based on time interval between administration of the second dose of  
200 Improvest (GnRF analog diphtheria toxoid conjugate) for immunocastration and slaughter. In  
201 a study by Harris et al. (2018), the thickness of pork belly increased as the time interval  
202 increased, and the PUFA percentage and IV increased as the time interval decreased.  
203 Therefore, additional studies are necessary to determine the optimal combination of  
204 immunocastration with other treatments.

205

### 206 **Genetic effect**

207 Genetic factors, including genotype and breed, are key determinants of meat quality.  
208 Understanding genetic factors is important to improve the quality of pork belly effectively.  
209 Studies have investigated the heritability and genetic correlations between pork belly quality  
210 traits (Hermesch, 2008; Kang et al., 2015; Lee et al., 2023a). In their study, the heritability of  
211 traits such as belly weight, dimensions, fat content, and muscle area of pork belly had a  
212 moderate heritability ranging from 0.2 to 0.5 (Hermesch, 2008; Kang et al., 2015; Lee et al.,  
213 2023a). In particular, studies by Kang et al. (2015) and Lee et al. (2023a) identified genetic  
214 parameters of several individual muscles of pork belly. These estimated genetic parameters  
215 suggest that pork belly traits can be improved through genetic selection. Additionally, Lee et  
216 al. (2023b) predicted key genes associated with pork belly traits, including transcription  
217 factors. They determined the traits related to pork belly yield and three muscle areas  
218 (cutaneous trunci muscle, rectus abdominis muscle, and external abdominal oblique muscle)  
219 in pork belly slices, and identified related genetic factors. The results confirmed that  
220 adipogenesis-associated transcription factors affected pork belly composition.

221 Various studies have been conducted on genotypes associated with pork quality to  
222 improve pig genetics (Table 1). Halothane is a well-known gene that influences pork quality

223 traits. Halothane gene is associated with a pale, soft, and exudative (PSE) meat. Pigs carrying  
224 the halothane gene have increased lean meat content, but have significantly negative effects  
225 on water-holding capacity and the color of meat (Swan et al., 2001). Swan et al. (2001)  
226 investigated pork belly quality based on genotype by comparing pigs without the halothane  
227 gene and pigs heterozygous or homozygous recessive for the halothane gene. Consistent with  
228 the known effects of the halothane gene, pigs lacking the halothane gene showed increased fat  
229 accumulation and pork belly firmness. The IGF2 (insulin like growth factor 2) is a gene  
230 involved in myogenesis. The A/G mutation at position 3072 within intron 3 of IGF2 affects  
231 up to 30% of the variation in muscle mass and up to 20% of backfat thickness (Clark et al.,  
232 2014). The quality of pork belly obtained from pigs heterozygous (AG) or homozygous (AA)  
233 for IGF2 mutation has been investigated (Clark et al., 2014). Thicker and firmer pork bellies  
234 were obtained from pigs that were heterozygous (AG) for the IGF2 mutation. The *CRTC*  
235 family regulates mitochondrial metabolic activity, and of the genes of this family, *CRTC3* has  
236 been reported to play an important role in controlling obesity development and energy  
237 metabolism (Lee et al., 2018). Lee et al. (2018) genotyped 360 Yorkshire pigs and identified  
238 the p.V515F mutation in exon 16 of 40 single-nucleotide polymorphisms. The p.V515F  
239 mutation in *CRTC3* gene significantly affected intermuscular fat thickness, total muscle area,  
240 and total fat percentage in the belly.

241 Pig breeds have continuously improved production capacity and meat quality.  
242 Commercial breeds of pigs include many different breeds such as Duroc, Yorkshire,  
243 Hampshire, and Landrace. Commercial purebred pig breeds include many different breeds,  
244 including the Duroc, Yorkshire, Hampshire, and Landrace. Duroc has excellent growth and  
245 muscle quality attributes and is used as a terminal sire (NSR, 2015). Yorkshire and Landrace  
246 have excellent litter size and birth and weaning weight and are used as parent-stock females  
247 (NSR, 2015). The difference in pig breed traits can ultimately affect the meat quality. Studies

248 have been conducted on the influence of pig breeds on meat quality, investigating the  
249 differences between breeds, such as single breeds, crossbreeds, or novel breeds (Bahelka et  
250 al., 2011; Lim et al., 2013; Lim et al., 2014; Lowell et al., 2019; Hoa et al., 2023). Lim et al.  
251 (2014, 2015) investigated the differences in the quality traits of pork belly from two- and  
252 three-way crossbreeds of Yorkshire, Berkshire, Chester White, Landrace, and Duroc pigs,  
253 which are widely used Korean commercial pigs. Yorkshire × Berkshire pigs showed the  
254 lowest moisture content and cooking loss. Yorkshire × Landrace pigs have a high MUFA  
255 composition, whereas Yorkshire × Chester White pigs have a high PUFA composition and  
256 high sensory evaluation results (Lim et al., 2013). On the other hand, in three-way crossbred  
257 pigs, there were no significant differences in most quality traits except for high moisture  
258 content in Yorkshire×Chester White×Yorkshire pigs and high sensory evaluation results in  
259 Yorkshire×Landrace×Duroc (Lim et al., 2014). Lowell et al. (2019) investigated the effects of  
260 breed type (Pietrain or Duroc) on pork quality traits by controlling inherent and  
261 environmental factors. These results confirmed that thicker and firmer pork bellies were  
262 obtained from Duroc sired pigs. This was consistent with the expectations that the Duroc  
263 breed had fast growth rate and higher intramuscular fat content and that the Pietrain breed had  
264 lean meat yield. Studies on crossbreeding between local and commercial breeds to improve  
265 meat quality have also been reported. The difference in pork belly quality was confirmed  
266 between a novel breed (Woori Heukdon, WHD) which crossbreeding between Duroc sow  
267 with Korean native black pig sire and a commercial breed (Landrace×Yorkshire×Duroc,  
268 LYD) (Hoa et al., 2023). The fat content and cooking loss increased in the WHD group.  
269 Additionally, WHD belly had a higher volatile aroma associated with a fatty odor, whereas  
270 LYD belly had a higher compound with a roasty odor (Hoa et al., 2023). There are various  
271 studies on crossbreeding with local breeds, but most studies deal with growth performance or  
272 overall carcass traits rather than pork belly quality (Pugliese & Sirtori, 2012). To enhance

273 pork belly quality, further studies are needed on pork belly traits across various breeds,  
274 including commercial breeds, crossbreeds, and novel breeds. Saikia et al. (2024) reported the  
275 effectiveness of genetic improvement based on the estimated feed conversion ratio breeding  
276 value. The effects of the genetic line (sire or dam) and feed efficiency groups (low,  
277 intermediate, or high) on breeding value were investigated. Belly weight and belly thickness  
278 was highest in the high-efficiency group of the sire line. Pork belly quality is influenced by  
279 various genetic parameters. Therefore, further research is needed to identify the genetic  
280 factors that can improve quality to meet the needs of producers and consumers.

281

## 282 **Growth performance**

283 Many studies have reported that the growth performance of pigs, such as carcass weight  
284 and fat content, is significantly related to the quality of the pork belly (Correa et al., 2008;  
285 Harsh et al., 2017; Hoa et al., 2021; Lee et al., 2023a; Vališ et al., 2005). In a study by Lee et  
286 al. (2023a), carcass weight had a strong positive genetic correlation with belly weight, total  
287 belly volume, and several muscle areas of the pork belly. Similar results have been reported  
288 by Correa et al. (2008) and Harsh et al. (2017). They reported that a thicker and firmer pork  
289 belly was obtained from heavier carcasses, and a higher proportion of SFA was observed in  
290 the group with a faster growth rate.

291 Hoe et al. (2021) and Albano-Gaglio et al. (2024) studied the relationship between fat  
292 content and pork belly quality. The high fat content of the carcass increased the yield of pork  
293 belly, reduces cooking loss, and decreased the proportion of PUFAs in the pork belly fat (Hoe  
294 et al., 2021). Additionally, the content of oleic acid-derived compounds associated with fatty  
295 and oily flavors increases, which can improve the sensory properties. Albano-Gaglio et al.  
296 (2024) reported similar results for fat content. However, these effects may differ depending  
297 on the breed, even if the fat content is similar. In that study (Albano-Gaglio et al., 2024),

298 although the groups had similar amount of fat content, Iberian×Duroc barrows produced  
299 lower width and firmer bellies compared to Duroc pigs. In conclusion, heavier pigs have a  
300 higher pork belly yield. However, increased carcass weight may increase fat accumulation in  
301 the pork belly, which should be considered because it affects consumer preferences.

302

### 303 **Exogenous factors**

#### 304 **Diet**

305 Dietary components can be readily transferred from feed to the muscle and fat tissues of  
306 pigs, thereby affecting pork quality (Soladoye et al., 2015). Changes in pork belly quality  
307 according to diet are summarized in Table 2.

308 Numerous studies have been conducted to improve the quality of pork belly fat and fatty  
309 acid composition by supplementing it with dietary fat. The fat sources used varied from  
310 vegetable oils, such as corn, flaxseed, and sunflower oil, to animal fats, such as poultry fat  
311 and beef tallow. Many studies have confirmed that the supply of fat significantly affects the  
312 fatty acid composition of pork bellies (Apple et al., 2007; Eggert et al., 2001; Gatlin et al.,  
313 2003; Kellner et al., 2014). Supplementing the diet with conjugated linoleic acid oil (CLA)  
314 increased the total CLA and SFA proportions in pork belly fat, decreased IV, and resulted in a  
315 firmer pork belly (Eggert et al., 2001). Eggert et al. (2001) noted that CLA functions as an  
316 anticarcinogen and antiatherogen in animals and can improve the fat properties of pork belly  
317 without significantly affecting lean meat properties. Varying the IV of pig feed changed the  
318 physical characteristics and fatty acid composition of the pork belly (Gatlin et al., 2003). In a  
319 study by Gatlin et al. (2003), the thickness of the pork belly decreased and its length increased  
320 with an increase in IV in pig feed. Additionally, the linoleic acid content in pork belly fat  
321 increased, while palmitic acid and stearic acid content decreased with variations in IV levels  
322 in pig feed. Supplying an animal fat source to pig feed can increase the SFA proportion in

323 pork belly, decrease IV levels, and produce a firmer pork belly (Apple et al., 2007; Kellner et  
324 al., 2014). Kellner et al. (2015) investigated whether feeding unsaturated fat followed by a  
325 withdrawal period could prevent quality deterioration in pork belly but found that the  
326 withdrawal period did not lead to improvement in the quality of pork belly. On the other hand,  
327 in some studies, the supply of dietary fat did not have a clear effect on the quality of pork  
328 belly fat (Engel et al., 2001; Huang et al., 2019; Swan et al., 2001). This may result from  
329 differences in genetic factors, the energy state of animals, or experimental conditions. Further  
330 studies under various conditions are needed to clarify the effect of dietary fat sources on pork  
331 belly quality.

332 Many studies have investigated the effects of feed supplementation with dried distiller  
333 grains with solubles (DDGS). DDGS is a by-product of ethanol production from grains, and  
334 extensive research has been conducted on its feeding value (Stein & Shurson, 2009).  
335 Researchers have conducted studies on DDGS, investigating the effects of treatments such as  
336 DDGS dosage (Overholt et al., 2016; Whitney et al., 2006), supplementation duration (Harris  
337 et al., 2018; Tavárez et al., 2014; Xu et al., 2010), or combination with other dietary sources  
338 (Browne et al., 2013; Davis et al., 2015; Gaffield et al., 2022; Lee et al., 2013; Villela et al.,  
339 2017), on pork belly quality. DDGS contains many unsaturated fatty acids, especially linoleic  
340 acid. Thus, feeding pigs a diet containing DDGS generally increases the IV levels. (Stein and  
341 Shurson, 2009). Whitney et al. (2006) and Overholt et al. (2016) reported similar results, that  
342 state increasing the DDGS content in pig diets increased the IV of pork belly fat, resulting in a  
343 thin and softer pork belly. The authors concluded that the optimal DDGS content in grower-  
344 finisher pig diets, which were formulated based on the total amino acid content, was less than  
345 20% (Whitney et al., 2006). Overholt et al. (2016) also investigated the effect of a diet  
346 supplemented with DDGS and found that pellet-fed pigs had heavier bellies with higher IV.  
347 Several studies have considered feeding strategies that included DDGS, followed by a



348 withdrawal period or a gradual reduction in DDGS (Harris et al., 2018; Tavárez et al., 2014;  
349 Xu et al., 2010). The increased IV and tenderness found in pork belly supplemented with  
350 DDGS were significantly reduced by including a withdrawal period for DDGS (Harris et al.,  
351 2018; Tavárez et al., 2014; Xu et al., 2010). Lee et al. (2012) investigated the characteristics  
352 of pigs fed DDGS and corn germ and found that the pork belly firmness in these pigs  
353 decreased regardless of the DDGS supply. Several studies have been conducted to reduce the  
354 negative effects of DDGS by supplying additional fat sources (beef tallow, palm kernel oil,  
355 glycerol, cottonseed oil, or yellow grease) to pig diets. However, most dietary fat sources did  
356 not reduce the decrease in pork belly firmness (Browne et al., 2013; Davis et al., 2015; Lee et  
357 al., 2013; Villela et al., 2017). Meanwhile, in a study where researchers added high-oleic  
358 soybean oil (HOSO) to diets with DDGS, it was found that the HOSO supplementation  
359 increased the proportion of MUFAs in pork belly fat and improved the physical properties and  
360 firmness (Gaffield et al., 2022).

361 Other dietary treatments for pigs have also been considered. In a study by Apple et al.  
362 (2011), the addition of carnitine with a fat source changed the fatty acid composition but did  
363 not significantly affect the dimensions or firmness of pork belly. Zhu et al. (2021) found that  
364 adding camelina press cake to pig diets decreased the thickness of pork belly but had no  
365 significant effect on firmness. The effect of supplying antioxidants to pig diets on pork belly  
366 quality has been previously investigated (An et al., 2019; Lu et al., 2014). Lu et al. (2014)  
367 investigated the effects of supplying antioxidants to a high-oxidant diet. Pigs fed oxidized  
368 diets had softer pork belly, and the addition of antioxidants tended to slightly improve  
369 firmness; however, the effect was not significant. An et al. (2019) reported that effects of  
370 supplying lycopene and tomato paste as antioxidants to pigs. There was no significant  
371 difference in the belly yield and lipid properties; however, the malondialdehyde content was  
372 reduced, which improved the oxidative stability. With the ban on the use of antibiotics in

373 livestock diets, Lowell et al. (2018) investigated the effect of antibiotic use on pork belly  
374 quality, and found no significant differences in pork belly quality between pigs fed antibiotic-  
375 free, natural antimicrobials, or antibiotics. Methionine (Met), an essential amino acid, is  
376 commonly added as a supplement to growing-finishing pig diets because it is the second most  
377 limiting amino acid in pigs. Remole et al. (2024) found that differences in Met source did not  
378 significantly affect pork belly quality. Therefore, diets with various ingredients and treatments  
379 can significantly affect pork belly quality, and further research should be conducted to  
380 produce high-quality pork belly.

381

### 382 **Others**

383 Pork belly quality is influenced by factors other than feed intake. Bryan et al. (2020)  
384 reported that infection with porcine reproductive and respiratory syndrome viruses (PRRSV)  
385 reduced pork belly firmness. Zomeño et al. (2024) investigated the effect of the boning  
386 processing method (hot or cold) on pork belly quality. They reported that the hot-boned belly  
387 (cut immediately postmortem) was shorter, wider, thicker, and firmer than the cold-boned  
388 belly (cut at 24 h postmortem) due to intense shortening and hardening. These results confirm  
389 that disease and carcass handling can affect pork belly quality. However, since studies  
390 addressing pork belly quality are insufficient, further research on various influencing factors  
391 is necessary.

392

### 393 **Conclusion**

394 This review identified the quality traits of pork belly and the factors that affect them  
395 based on previous studies. Pork belly quality was assessed based on belly yield, dimensions  
396 (length, width, and thickness), firmness, and fatty acid composition. Factors affecting pork  
397 belly quality include endogenous factors such as sex, breed, and carcass weight, and

398 exogenous factors such as diet. Many studies have focused on improving the fatty acid  
399 composition and firmness of pork bellies in the context of dietary effects. The yield and fat  
400 deposition of pork belly were higher in barrow than in gilt and immunocastration had lower  
401 fat content and softer pork belly. The adipogenesis-associated transcription factors and genes  
402 involved in growth affected the pork belly quality. It was confirmed that the pork belly quality  
403 traits differ with various pig breeds. Dietary fat sources can be used to improve the fat quality  
404 and fatty acid composition of pork belly.

405 However, there is still a need for discussion on good-quality pork belly owing to the  
406 differences in perspectives between producers and consumers regarding pork belly quality.  
407 Particularly, discussions are necessary to balance the health aspects with the economic and  
408 sensory attributes according to the fatty acid composition and fat content of pork belly. In  
409 addition, studies focusing only on pork belly quality are significantly lacking. Most studies  
410 considered pork belly quality to be a part of the carcass quality change and often not  
411 addressed as a major issue. Therefore, to clarify the appropriate pork belly quality according  
412 to changing consumption patterns, research focusing on pork belly quality should be  
413 continuously conducted.

414

#### 415 **Conflicts of interest**

416 The authors declare no potential conflicts of interest.

417

418

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425           Conceptualization: Samooel Jung. Writing-original draft: Kyung Jo. Data curation:  
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427 Seong. Writing-review & editing: Kyung Jo, Seonmin Lee, Seul-Ki-Chan Jeong, Hayeon  
428 Jeon, Hyeun Bum Kim, Pil Nam Seong, Samooel Jung.

429

430 **Ethics Approval**

431           This article does not require IRB/IACUC approval because there are no human an  
432 animal participants.

433

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648

649 **Figure legend.**

650

651 **Figure 1. Pork belly quality PRISMA (Preferred Reporting Items for Systematic**  
652 **Reviews and Meta-Analyses) flow diagram.**

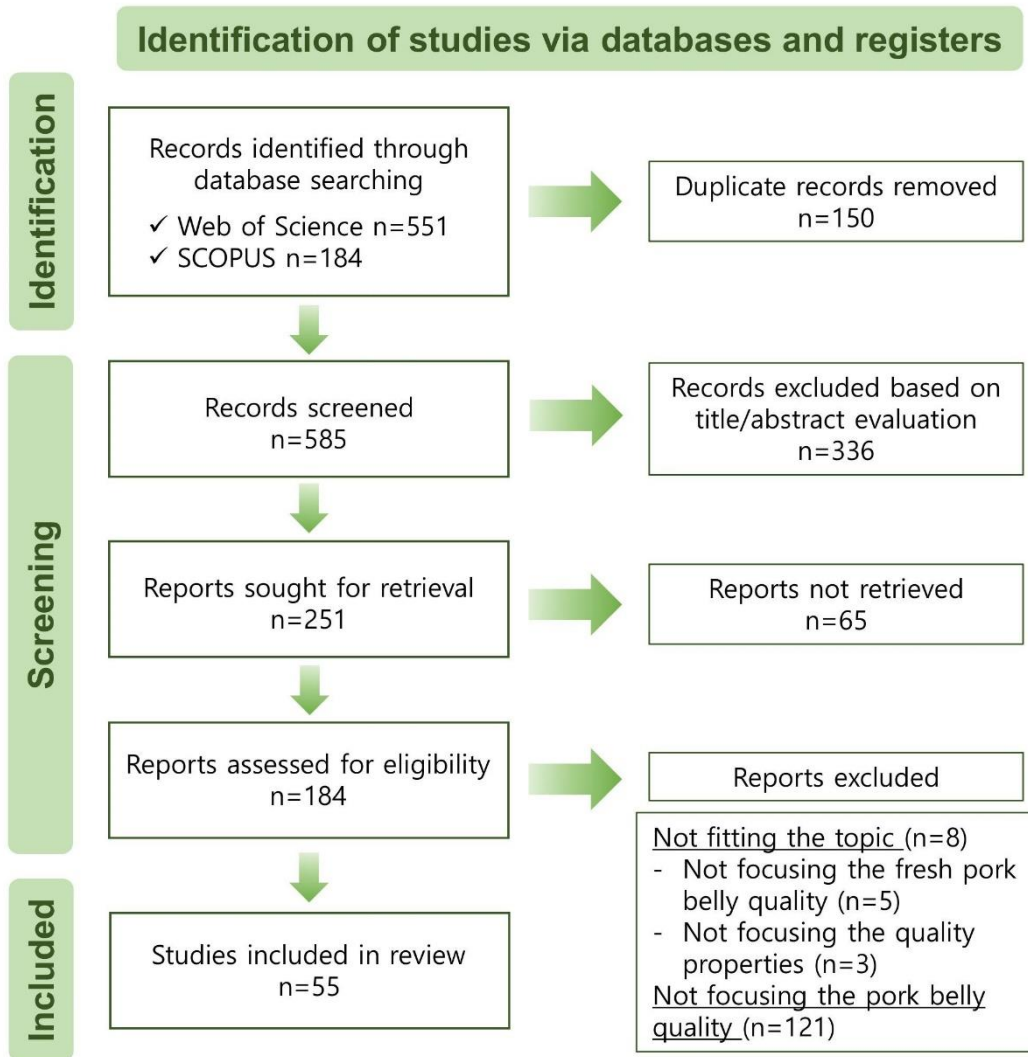
653

654 **Figure 2. Quality traits of pork belly evaluated in 55 selected literature**

655 <sup>1</sup> The number of literature mentioning the quality traits among the total 55 literature

656

ACCEPTED



657

658 **Figure 1.**

A total of 55 literatures on the quality traits of pork belly

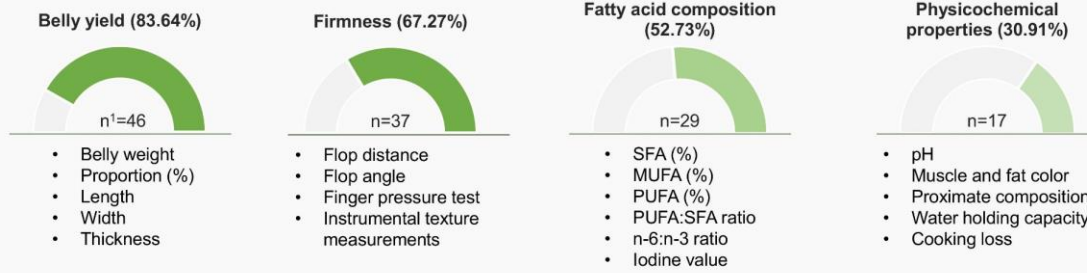


Figure 2.

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**Table 1. Summary of effect of endogenous factors on pork belly quality**

Treatment	Effects on pork belly quality	Reference
<b>Sex effect</b>		
Gilt or barrow	<ul style="list-style-type: none"> <li>• Belly proportion in the carcass: higher in barrow</li> <li>• Lean meat proportion in belly: higher in gilt</li> <li>• Back fat thickness: higher in barrow</li> <li>• Firmness: softer in gilt</li> </ul>	Stupka et al., 2004
Gilt or barrow	<ul style="list-style-type: none"> <li>• Fatty acid composition               <ul style="list-style-type: none"> <li>- ↓ SFA and ↑ linoleic acid and PUFA in gilt</li> <li>- ↑ iodine value (IV) in gilt</li> </ul> </li> <li>• Belly proportion in the carcass: higher in barrow</li> <li>• Meat and fat in the belly (%)               <ul style="list-style-type: none"> <li>- Higher meat proportion in gilt belly</li> <li>- Higher content of fat in barrow belly</li> </ul> </li> <li>• Belly weight: heavier belly in barrow</li> <li>• Firmness: softer in gilt</li> </ul>	Correa et al., 2008
Gilt or barrow	<ul style="list-style-type: none"> <li>• Fatty acid composition               <ul style="list-style-type: none"> <li>- ↑ palmitic acid and ↓ linolenic acid in barrow</li> <li>- ↑ IV in gilt</li> </ul> </li> <li>• Weight, width, and thickness: heavier, wider, and thicker belly in barrows</li> <li>• Firmness: firmer belly in barrows</li> </ul>	Bahelka et al., 2011
Gilt or barrow	<ul style="list-style-type: none"> <li>• Weight, width, and thickness: heavier, wider, and thicker belly in barrows</li> <li>• Firmness: firmer belly in barrows</li> </ul>	Lee et al., 2013
Gilt or barrow	<ul style="list-style-type: none"> <li>• Weight, width, and thickness: heavier, wider, and thicker belly in barrows</li> <li>• Firmness: firmer belly in barrows</li> </ul>	Overholt et al., 2016
<b>Castrated methods</b>		
Immunocastrated males (IC), surgically castrated males (SC), intact males (IM), or females (FE)	<ul style="list-style-type: none"> <li>• pH: IC &gt; SC and FE</li> <li>• Color of IC: ↓ L* value than SC and ↑ a* value than SC and FE</li> <li>• Water holding capacity: IC &lt; SC</li> <li>• Cooking loss: FE ≥ SC ≥ IC = EM</li> <li>• Fat content: highest in SC and lowest in EM belly</li> <li>• Visual evaluation: higher score in IC and FE belly</li> <li>• Sensory evaluations               <ul style="list-style-type: none"> <li>- Tenderness: lower in EM</li> <li>- Juiciness: higher in SC</li> <li>- Overall acceptability: higher in SC and lower in EM</li> </ul> </li> <li>• Width: widest belly in IC barrows fed ractopamine hydrochloride</li> <li>• Thickness: thicker belly in SC barrow than IM</li> </ul>	Jeong et al., 2011
Physically castrated (SC), immunologically castrated barrow (IC), intact male (IM), or gilt	<ul style="list-style-type: none"> <li>• Firmness               <ul style="list-style-type: none"> <li>- Highest flop distances in belly of SC barrow</li> <li>- No differences between IC fed ractopamine and gilts</li> <li>- Lowest flop distance in belly of IM</li> </ul> </li> <li>• Fatty acid composition               <ul style="list-style-type: none"> <li>- ↑ IV in IM and no difference between IC and</li> </ul> </li> </ul>	Kyle et al., 2014

	gilt	
	- ↓ SFA and MUFA and ↑ PUFA in IM	
Physically castrated (SC) or immunologically castrated (IC) and ractopamine hydrochloride diet	<ul style="list-style-type: none"> <li>• Thickness: thicker belly in SC</li> <li>• Firmness (flop): softer belly in IC</li> </ul>	Lowe et al., 2016
Gilt, immunocastrated (IC), or barrow (surgically castrated, SC)	<ul style="list-style-type: none"> <li>• Proximate content: ↑ protein content and ↓ lipid content in IC than SC</li> <li>• Color: ↑ a* value in gilt belly meat</li> <li>• Backfat thickness: higher in IC than gilt</li> <li>• Fatty acid composition <ul style="list-style-type: none"> <li>- ↑ PUFA, omega-3, and omega-6 in IC than SC</li> <li>- ↓ SFA in gilt than SC</li> <li>- ↑ IV in gilt and IC</li> </ul> </li> </ul>	Costa e silva et al., 2017
Surgically castrated males (SCM), entire females (FE), immunocastrated females (ICF)	<ul style="list-style-type: none"> <li>• Belly proportion (%): highest in SCM</li> <li>• Firmness: firmer belly in SCM</li> <li>• Proximate content: ↑ dry matter and fat and ↓ moisture and protein in SCM</li> <li>• Fatty acid composition <ul style="list-style-type: none"> <li>- SFA and MUFA were not significantly different between sexual types</li> <li>- ↑ linoleic acid and PUFA in FE and ICF</li> <li>- IV: FE ≥ ICF ≥ SCM</li> </ul> </li> </ul>	Font-i-Furnols et al., 2023
Immunocastrated males (ICM) or entire males	<ul style="list-style-type: none"> <li>• Belly length: longer belly in ICM</li> <li>• Firmness: Firmer belly in ICM</li> <li>• Fatty acid composition <ul style="list-style-type: none"> <li>- ↑ SFA and ↓ PUFA, PUFA/SFA ratio, and IV in ICM</li> </ul> </li> </ul>	Font-i-Furnols et al., 2023
Physically castrated (SC) or immunologically castrated (IC) barrow	<ul style="list-style-type: none"> <li>• Width: wider belly in IC</li> <li>• Thickness: thicker belly in SC</li> <li>• Firmness (flop): tended to firmer belly in SC</li> </ul>	Tavárez et al., 2014
Time intervals between second Improvest® dose and slaughter	<ul style="list-style-type: none"> <li>• Thickness: increases linearly as time interval increase</li> <li>• Fatty acid composition <ul style="list-style-type: none"> <li>- ↑ PUFA and IV as time interval decrease</li> </ul> </li> </ul>	Harris et al., 2018
<b>Genetic effect, Genotype</b>		
Stress genotype		
<ul style="list-style-type: none"> <li>• Negative = NN (halothane-free), carrier = Nn, or positive = nn</li> </ul>	<ul style="list-style-type: none"> <li>• Firmness: increased in stress-negative genotype</li> <li>• Proximate content: ↓ moisture and protein and ↑ lipid in stress-negative genotype</li> </ul>	Swan et al., 2001

(homozygous recessive for the halothane gene)		
Genotype, IGF2-G3072A mutation	<ul style="list-style-type: none"> <li>• Thickness: thicker belly in AG pigs than AA pigs</li> </ul>	Clark et al., 2014
<ul style="list-style-type: none"> <li>• Heterozygous (AG) or homozygous (AA)</li> </ul>	<ul style="list-style-type: none"> <li>• Firmness: firmer belly in AG pigs</li> <li>• IV: tended to higher IV in AA pigs</li> </ul>	
Genotype, CRT3-p.V515F mutation	<ul style="list-style-type: none"> <li>• Intermuscular fat thickness: thinner in pigs with the TT genotype</li> </ul>	Lee et al., 2018
<ul style="list-style-type: none"> <li>• GG, TG, or TT</li> </ul>	<ul style="list-style-type: none"> <li>• Total muscle area: greater in pigs with heterozygous genotype (GG and TT)</li> </ul>	
	<ul style="list-style-type: none"> <li>• Total fat percentage: TG&gt;GG&gt;TT</li> </ul>	
<b>Genetic effect, Breed</b>		
Sire line	<ul style="list-style-type: none"> <li>• Belly proportion in the carcass: LA&gt;HA×PN&gt;YO×PN</li> </ul>	Bahelka et al., 2011
<ul style="list-style-type: none"> <li>• Hampshire (HA)× Pietrain (PN), Landrace (LA), or Yorkshire (YO)×PN</li> </ul>	<ul style="list-style-type: none"> <li>• Meat and fat in the belly (%) <ul style="list-style-type: none"> <li>- HA×PN: highest percentage of meat</li> <li>- YO×PN: highest percentage of fat, skin, and bones</li> </ul> </li> </ul>	
	<ul style="list-style-type: none"> <li>• pH: lowest in YC</li> <li>• Proximate content: ↓ moisture content in YB belly</li> <li>• Cooking loss: lower in YB</li> </ul>	
Two-way crossbreeds	<ul style="list-style-type: none"> <li>• TBARS values: higher in YB at 14 d</li> </ul>	
<ul style="list-style-type: none"> <li>• Yorkshire×Landrace (YL), Yorkshire×Berkshire (YB), or Yorkshire×Chester White (YC)</li> </ul>	<ul style="list-style-type: none"> <li>• Fatty acid composition <ul style="list-style-type: none"> <li>- YL: ↑ stearic acid, oleic acid, and MUFA</li> <li>- YB and YC: ↑ myristic acid, linoleic acid, linolenic acid, and n-6 fatty acids</li> <li>- YC: ↑ PUFA</li> </ul> </li> </ul>	Lim et al., 2013
	<ul style="list-style-type: none"> <li>• Free amino acid composition: ↑ concentrations of most free amino acids in YB</li> <li>• Sensory evaluation: higher score in YC</li> </ul>	
Three-way crossbreeds		Lim et al., 2014
<ul style="list-style-type: none"> <li>• Yorkshire × Landrace × Duroc (YLD), Yorkshire × Chester White × Yorkshire (YCY), and Yorkshire × Berkshire × Duroc (YBD)</li> </ul>	<ul style="list-style-type: none"> <li>• Proximate content: highest moisture content in YCY belly</li> </ul>	
	<ul style="list-style-type: none"> <li>• Sensory evaluation: higher score in YLD</li> </ul>	
Sire line	<ul style="list-style-type: none"> <li>• Thickness: thicker belly in Duroc sired pigs</li> </ul>	Lowell et al., 2019
<ul style="list-style-type: none"> <li>• Pietrain or Duroc ancestry</li> </ul>	<ul style="list-style-type: none"> <li>• Firmness: greater flop distance in Duroc sired pigs</li> </ul>	

Breed	<ul style="list-style-type: none"> <li>• LYD (Landrace × Yorkshire or novel pig breed (Woori Heukdon, WHD))</li> </ul>	<ul style="list-style-type: none"> <li>• Belly yield (%): higher in WHD</li> <li>• Proximate composition: ↑ fat content and ↓ moisture, protein, and collagen in WHD</li> <li>• Cooking loss: lower in WHD</li> <li>• Color: ↓ L* value and ↑ a* value in WHD</li> <li>• Fatty acid composition               <ul style="list-style-type: none"> <li>- ↑ MUFA and UFA and ↓ SFA in WHD</li> </ul> </li> <li>• Volatile aroma composition               <ul style="list-style-type: none"> <li>- ↑ compounds associated with fatty odor in WHD</li> <li>- ↑ compounds associated with roasty odor in LYD</li> </ul> </li> </ul>	Hoa et al., 2023
Genetic line effect	<ul style="list-style-type: none"> <li>• Sire or dam line</li> </ul> <p>Estimated breeding value</p> <ul style="list-style-type: none"> <li>• Feed efficiency: low, intermediate, or high</li> </ul>	<ul style="list-style-type: none"> <li>• Belly weight: heaviest belly in sire high efficiency group</li> <li>• Thickness: thickest belly in sire high efficiency group</li> </ul>	Saikia et al., 2024
<b>Growth performance</b>			
Growth rate Slaughter weight		<ul style="list-style-type: none"> <li>• Back fat thickness: higher with weight increase</li> <li>• Fatty acid composition               <ul style="list-style-type: none"> <li>- ↑ PUFA:SFA ratios and n-6:n-3 ratios in slow growing</li> <li>- ↑ stearic fatty acid and SFA proportions in fast growing</li> </ul> </li> </ul>	Correa et al., 2008
Carcass weight		<ul style="list-style-type: none"> <li>• Thickness: thicker belly with increase carcass weight</li> <li>• Firmness: firmer belly with increase carcass weight</li> <li>• IV: decreased IV with increase carcass weight</li> <li>• Belly yield: higher yield with increased fat level</li> <li>• Proximate composition: ↑ fat and ↓ moisture, protein, and collagen in high fat level</li> <li>• Color: ↑ b* value in high fat level</li> <li>• Cooking loss: decreased with increased fat level</li> </ul>	Harsh et al., 2017
Fat levels		<ul style="list-style-type: none"> <li>• Fatty acid composition               <ul style="list-style-type: none"> <li>- ↑ oleic acid and ↓ PUFA in high fat level</li> </ul> </li> <li>• Volatile aroma composition               <ul style="list-style-type: none"> <li>- ↑ Maillard reaction-derived flavor compound (meaty and roasty flavors) in low fat level group</li> <li>- ↑ oleic acid-derived compounds (fatty and oily flavors) in high fat level group</li> </ul> </li> </ul>	Hoa et al., 2021
Fatness and genetic effect	<ul style="list-style-type: none"> <li>• F1: 12.3-25.9%, F2: 26.0-33.9%, and F3:</li> </ul>	<ul style="list-style-type: none"> <li>• Sensory properties: higher score in high fat level group</li> <li>• Belly weight: heaviest in F5 and lightest in F1 pigs</li> <li>• Belly proportion: lowest in F5 and no significant difference between commercial pigs (F1-3)</li> <li>• Length: increased across the bellies from F1 to F4</li> </ul>	Albano-Gaglio et al., 2024

- 34.0-47.1% of fat content from commercial pigs
  - F4: 36.4-56.3% of fat content from Duroc pigs
  - F5: 55.0-69.1% of fat content from Iberian×Duroc barrows
  - Width: lowest in F5 and no significant difference between F1-F4
  - Firmness: firmer belly in F5 and softer belly in F1
  - Proximate composition: ↑ lipid content and ↓ moisture, protein, and ash content with increased fatness
  - Fatty acid composition
    - ↑ SFA and MUFA and ↓ PUFA with increased fatness in commercial pigs
    - ↑ oleic acid and ↓ linoleic acid in F5
  - IV: F1>F2>F3>F4>F5
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**Table 2. Summary of effect of diets on pork belly quality**

Treatment	Effects on pork belly quality	Reference
<b>Dietary fat source</b>		
<p>Conjugated linoleic acid (CLA)</p> <ul style="list-style-type: none"> <li>Control or 0.75% CLA</li> </ul> <p>Dietary fat source</p> <ul style="list-style-type: none"> <li>Choice white grease or poultry fat</li> <li>Level: 2, 4, or 6%</li> </ul>	<ul style="list-style-type: none"> <li>Proximate content: ↑ moisture and protein content and ↓ lipid content</li> </ul>	Swan et al., 2001
<p>Conjugated linoleic acid</p> <ul style="list-style-type: none"> <li>1% CLA oil, 1% sunflower oil, or fed the sunflower oil-supplemented diet restricted to the amount consumed by pigs fed the CLA diet</li> </ul>	<ul style="list-style-type: none"> <li>Firmness: increased in the CLA group</li> <li>Fatty acid composition: <ul style="list-style-type: none"> <li>↑ total CLA and SFA and ↓ MUFA and UFA in the CLA group</li> <li>IV: lower in the CLA group</li> </ul> </li> </ul>	Eggert et al., 2001
<p>Hydrogenated dietary fat</p> <ul style="list-style-type: none"> <li>Supplement with 5% choice white grease to IV of 20, 40, 60, or 80</li> </ul> <p>Dietary fat source</p> <ul style="list-style-type: none"> <li>5% Beef tallow (BT) or soybean oil (SBO)</li> </ul>	<ul style="list-style-type: none"> <li>Thickness: decreased with increasing IV of diet</li> <li>Length: increased with increasing IV of diet</li> <li>Fatty acid composition <ul style="list-style-type: none"> <li>↑ IV with increasing IV of diet</li> <li>↑ linoleic acid and ↓ palmitic acid and steric acid with increasing IV of diet</li> </ul> </li> </ul>	Gatlin et al., 2003
<p>Dietary fat source</p> <ul style="list-style-type: none"> <li>3 or 6% of choice white grease (CWG), corn oil (CO), or beef tallow (TAL)</li> </ul>	<ul style="list-style-type: none"> <li>Firmness: firmer belly in BT group</li> <li>Color of belly fat: lighter and redder in BT group</li> <li>Fatty acid composition <ul style="list-style-type: none"> <li>↓ PUFA and ↑ SFA and MUFA in BT group</li> </ul> </li> </ul>	Apple et al., 2007
	<ul style="list-style-type: none"> <li>Weight: increased in pigs fed dietary fat source</li> <li>Firmness: firmer belly in pigs fed beef tallow</li> <li>IV: increased in pigs fed corn oil</li> </ul>	Kellner et al., 2014

Dietary fat source		
<ul style="list-style-type: none"> <li>0 or 1% flaxseed oil + 1,3, or 5% poultry fat</li> </ul>	<ul style="list-style-type: none"> <li>Width and thickness: ↑ width and thickness with ↑ dietary lipids</li> </ul>	Huang et al., 2019
Vitamin E		
<ul style="list-style-type: none"> <li>11 or 220 IU/kg</li> </ul>		
Dietary fat withdrawal times		
<ul style="list-style-type: none"> <li>21, 42, or 63 d before slaughter</li> </ul>		
Dietary fat unsaturation loads	<ul style="list-style-type: none"> <li>Belly weight and thickness: no effect</li> <li>Firmness: ↓ belly firmness with increasing the dietary fat unsaturation loads</li> </ul>	Kellner et al., 2015
5% corn oil (HIGH), 5% animal-vegetable blend (MED), or 2.5% corn oil (LOW)		

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### Dried distillers grains with solubles (DDGS)

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DDGS	<ul style="list-style-type: none"> <li>Thickness: ↓ thickness with ↑ DDGS concentration</li> <li>Firmness: ↓ firmness with ↑ DDGS concentration</li> <li>IV: increased with increased DDGS concentration</li> </ul>	Whitney et al., 2006
<ul style="list-style-type: none"> <li>0, 10, 20, or 30%</li> </ul>		
DDGS with withdrawal period	<ul style="list-style-type: none"> <li>Firmness: Softer belly with feeding 30% DDGS without withdrawal period</li> <li>Fatty acid composition <ul style="list-style-type: none"> <li>↑ PUFA and IV and ↓ SFA and MUFA with ↑ DDGS</li> <li>↓ IV with ↑ DDGS withdrawal period</li> </ul> </li> <li>Weight: tend to decreased with feeding DDGS</li> <li>Length: decreased with increase corn germ in diet without DDGS</li> </ul>	Xu et al., 2010
<ul style="list-style-type: none"> <li>DDGS: 0, 15, or 30%</li> <li>Withdrawal: 0, 3, 6, or 9 week</li> </ul>		
DDGS	<ul style="list-style-type: none"> <li>Firmness <ul style="list-style-type: none"> <li>↓ flop distance in feeding DDGS</li> <li>↓ flop distance with corn germ without DDGS supplement</li> </ul> </li> </ul>	Lee et al., 2012
<ul style="list-style-type: none"> <li>0 or 30%</li> </ul>		
Corn germ		
<ul style="list-style-type: none"> <li>0, 10, 20, or 30%</li> </ul>		
DDGS + Dietary fat source	<ul style="list-style-type: none"> <li>Firmness: softer belly in YG fed during all 5 feeding phases than BT</li> <li>Fat color: ↓ b* value as time fed BT increased</li> <li>Fatty acid composition <ul style="list-style-type: none"> <li>↑ SFA and MUFA concentrations in belly fat with BT fed during all 5 feeding phases</li> <li>↑ PUFA and IV in belly fat with YG fed during all 5 feeding phases</li> </ul> </li> </ul>	Browne et al., 2013
<ul style="list-style-type: none"> <li>Beef tallow (BT, 5%) or yellow grease (YG, 4.7%)</li> <li>5 feeding phases</li> </ul>		

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DDGS			
• DDGS: 0 or 30%	• Firmness: ↑ flop distance in pigs fed DDGS		
Dietary treatment	• Fatty acid composition		
• Corn germ, beef tallow, palm kernel oil, or glycerol	- ↑ oleic acid content in control group than pigs fed DDGS except pigs fed beef tallow	Lee et al., 2013	
	- ↑ MUFA in control group than pigs fed DDGS or corn germ		
	- ↑ MUFA in pigs fed beef tallow than pigs fed DDGS		
DDGS			
• 0%, 30% DDGS with withdrawal, or 30% DDGS without withdrawal	• Width: tended to wider belly with fed DDGS	Tavárez et al., 2014	
	• Firmness: softer belly with fed DDGS		
	• IV: increased with fed DDGS without withdrawal		
	• Thickness: tend to be thicker in tallow fed pigs		
	• Length: decreased in tallow fed pigs		
	• Firmness: softer belly in DDGS fed pigs and tend to decreased of flop angle in tallow fed pigs		
DDGS	• Fatty acid composition		
• 0 or 30%	- ↓ oleic acid, MUFA, and SFA and ↑ PUFA in DDGS fed pigs	Davis et al., 2015	
Tallow	- ↑ oleic acid and MUFA and ↓ SFA in tallow fed pigs		
• 0 or 5%	• IV: increased in DDGS fed pigs and decreased when tallow added to diets with DDGS		
	• Fat color: ↓ L*, a*, and b* value in pigs fed DDGS		
	• Belly weigh: heavier belly in pellet-fed pigs		
Diet form	• Thickness: reduced in 30 % DDGS-fed pigs		
• Meal or pelleted	• Firmness: ↓ flop distance in 30% DDGS-fed pigs	Overholt et al., 2016	
	• Fatty acid composition		
DDGS	- ↑ PUFA and ↓ MUFA and SFA in pellet-fed pigs and 30% DDGS-fed pigs		
• 0 or 30%	- ↑ IV in pellet-fed pigs and 30% DDGS-fed pigs		
	• Thickness: highest in pig fed cottonseed oil		
40% DDGS and dietary treatment	• Firmness		
• Cottonseed oil or crude glycerol	- Compression force tended to be less in pigs fed glycerol than 40% DDGS	Villela et al., 2017	
	• Fatty acid composition		
	- ↑ SFA, PUFA, and IV and ↓ MUFA in pigs fed cottonseed oil		
DDGS feeding strategies	• Belly percentage: lowest in NCon and highest in PCon		
• Corn-soybean meal with 0% DDGS (PCon)	• Thickness: thinner belly in NCon and similar thickness in PCon and WD		
	• Firmness: flop distance, PCon> SD>WD>NCon, softer belly in PCon	Harris et al., 2018	
• Progressive reduction in	• Color of belly fat: ↓ L* value in NCon		
	• Fatty acid composition		



<ul style="list-style-type: none"> <li>DDGS supply (SD)</li> <li>DDGS 40% with withdrawal period (WD)</li> <li>DDGS 40% in all phase (NCon)</li> </ul>	<ul style="list-style-type: none"> <li>↓ SFA and MUFA and ↑ PUFA in NCon</li> <li>IV: NCon&gt;WD=SD&gt;PCon</li> </ul>	
<p>High oleic soybean oil (HOSO)</p> <ul style="list-style-type: none"> <li>25% DDGS or HOSO (2, 4, or 6%)</li> </ul>	<ul style="list-style-type: none"> <li>Width: higher in DDGS fed pigs and lower in 2 and 4% HOSO fed pigs</li> <li>Thickness: Thicker belly in HOSO fed pigs</li> <li>Firmness: firmer belly in HOSO fed pigs</li> <li>Fatty acid composition               <ul style="list-style-type: none"> <li>↓ SFA with increasing HOSO levels in pig diet</li> <li>↑ MUFA with increasing HOSO levels in pig diet</li> <li>↑ PUFA in DDGS fed pigs</li> <li>IV: highest in 6% HOSO fed pigs and lowest in 2% HOSO fed pigs</li> </ul> </li> </ul>	Gaffield et al., 2022
<b>Others</b>		
<p>L-carnitine (CARN)</p> <ul style="list-style-type: none"> <li>0 or 100 mg/kg</li> </ul> <p>Corn oil</p> <ul style="list-style-type: none"> <li>0, 2, or 4%</li> </ul>	<ul style="list-style-type: none"> <li>Firmness: decrease linearly with corn oil content in diet</li> <li>Fatty acid composition               <ul style="list-style-type: none"> <li>CARN supplement: ↑ SFA in the intermuscular fat layer, ↑ MUFA in the lean layers, and ↓ PUFA in the intermuscular fat and <i>cutaneous trunci</i> muscle</li> <li>Corn oil: ↓ SFA and MUFA composition and ↑ PUFA content</li> </ul> </li> </ul>	Apple et al., 2011
<p>Antioxidant</p> <ul style="list-style-type: none"> <li>High oxidant diet, 11 IU/kg vitamin E, antioxidant blend, vitamin E + antioxidant blend, and standard corn-soy control diet</li> </ul>	<ul style="list-style-type: none"> <li>Length: decreased belly length in pigs fed high oxidant diet and vitamin E</li> <li>Width: lowest in pigs fed high oxidant diet</li> <li>Firmness: firmer belly in pigs fed corn-soy diet and softer belly in vitamin E pigs</li> </ul>	Lu et al., 2014
<p>Antibiotic or antimicrobial</p> <ul style="list-style-type: none"> <li>Antibiotic free, natural antimicrobial (0.025% oregano), or antibiotic (40</li> </ul>	No significant effect	Lowell et al., 2018

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mg/kg tylosin phosphate)		
Lycopene, tomato paste, or both	<ul style="list-style-type: none"> <li>• MDA content: lower MDA concentrations in feeding lycopene or tomato paste</li> <li>• Fatty acid composition: no effect</li> </ul>	An et al., 2019
Camelina press cake (CPC) 0, 5, 10, or 15% Methionine (Met) source	<ul style="list-style-type: none"> <li>• Thickness: decreased thickness with increased CPC level</li> <li>• Firmness: no effect</li> </ul>	Zhu et al., 2021
L-Met, DL-Met, or calcium salt of DL-Met hydroxyl analog	No significant effect	Remole et al., 2024

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