Korean Journal for Food Science of Animal Resources

Korean J. Food Sci. An. 2018 June 38(3):442~450

DOI https://doi.org/10.5851/kosfa.2018.38.3.442 DOI https://doi.org/10.5851/kosfa.2018.38.3.442

ARTICLE

Development and Validation of Predictive Model for *Salmonella* Growth in Unpasteurized Liquid Eggs

Young-Jo Kim¹, Hye-Jin Moon², Soo-Kyoung Lee³, Bo-Ra Song¹, Jong-Soo Lim³, Eun-Jeong Heo¹, Hyun-Jung Park¹, Sung-Hwan Wee³, and Jin-San Moon^{3,*}

¹Ministry of Food and Drug Safety, Cheongju 28159, Korea 2Department of Food and Nutrition, Kyung Hee University, Seoul 02447, Korea 3Animal and Plant, Quarantine Agency, Gimcheon 39660, Korea

Abstract Liquid egg products can be contaminated with *Salmonella* spp. during processing. A predictive model for the growth of *Salmonella* spp. in unpasteurized liquid eggs was developed and validated. Liquid whole egg, liquid yolk, and liquid egg white samples were prepared and inoculated with *Salmonella* mixture (approximately 3 Log CFU/mL) containing five serovars (*S.* Bareilly, *S.* Richmond, *S.* Typhimurium monophasic, *S*. Enteritidis, and *S*. Gallinarum). *Salmonella* growth data at isothermal temperatures (5, 10, 15, 20, 25, 30, 35, and 40℃) was collected by 960 h. The population of *Salmonella* in liquid whole egg and egg yolk increased at above 10℃, while *Salmonella* in egg white did not proliferate at all temperature. These results demonstrate that there is a difference in the growth of *Salmonella* depending on the types of liquid eggs (egg yolk, egg white, liquid whole egg) and storage temperature. To fit the growth data of *Salmonella* in liquid whole egg and egg yolk, Baranyi model was used as the primary model and the maximum growth rate and lag phase duration for each temperature were determined. A secondary model was developed with maximum growth rate as a function of temperature. The model performance measures, bias factor (B*f*, 0.96- 0.99) and r^2 (0.96 - 0.99) indicated good fit for both primary and secondary models. In conclusion, it is thought that the growth model can be used usefully to predict *Salmonella* spp. growth in various types of unpasteurized liquid eggs when those are exposed to various temperature and time conditions during the processing.

Keywords liquid whole egg, liquid egg yolk, liquid egg white, *Salmonella* spp., storage temperatures

Introduction

Eggs are a representative perfect food and widely used as raw material of processed foods. There are various processed forms of liquid eggs – whole egg, egg yolk, and egg white. Egg yolk is used for mayonnaise flavoring and additives, and egg white is used for processed meat products, marine products and liquefied beverages. Mass liquid eggs are used as raw materials in various food industries and the demand for liquid eggs is expected to increase further with its convenience and economic efficiency (Kim

OPEN ACCESS

***Corresponding author** : Jin-San Moon Animal and Plant, Quarantine Agency, Gimcheon 39660, Korea Tel: $+82-54-912-0530$ Fax: +82-54-912-0530 E-mail: moonjs727@korea.kr

[©] Korean Society for Food Science of Animal Resources. This is an open access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licences/by-nc/3.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

et al., 2014). However, pasteurization of liquid eggs is not simple since proteins of egg white are denatured at a temperature of 57°C or higher (Kang et al., 2011). In the removal processing of the egg shells, unpasteurized eggs are exposed to pathogenic microorganisms at high risk. Accordingly, caution needs to be taken when unpasteurized liquid eggs are used for processed products. In Sakha et al. (2012), *S.* Enteritidis proliferated more rapidly in unpasteurized liquid eggs than in pasteurized liquid eggs.

Salmonellosis continues to be a major food-borne disease in food industry, especially, poultry related products. In Korea, the criteria for microbes in unpasteurized liquid eggs are defined as less than 500,000 CFU per gram of bacteria, less than 100 CFU per gram of *Escherichia coli* and negative for *Salmonella* (MFDS, 2015). It should also be cooled to below 5℃ after egg breaking and should not be stored for more than 72 hours (MFDS, 2015). However, secondary contamination by *Salmonella* may occur by various factors such as fecal matter, dust, and so on during the process of manufacturing liquid eggs. Actually, three types of *Salmonella* serotypes were isolated as a result of monitoring liquid eggs distributed domestically (Kim et al., 2015). *Salmonella* growth is affected by internal factors such as the moisture activity of food, pH, initial microorganisms and external factors such as the storage temperature, packaging method, and storage humidity. Therefore, proper management in the production, storage, and distribution processes is necessary to control *Salmonella*.

In order to establish the appropriate temperature and time for food storage and distribution, models for predicting the growth of *Salmonella* in egg products were developed (Elias et al., 2016; Na, 2014; Sakha et al., 2012; Singh et al., 2011). But most studied mainly on *S.* Enteritidis growth and did not deal with other serotypes of *Salmonella*. Recent study on the growth and survival of *Salmonella* in unpasteurized egg yolk and egg white (Moon et al., 2016) showed that there is a slight difference in *Salmonella* growth between *S.* Enteritidis and other serotypes of *Salmonella.* In the study of Moon et al. (2016), *S*. Enteritidis proliferated more rapidly than other serotypes, which was accordance with the *Salmonella* growth predictive model of homemade mayonnaise (Malheiros et al., 2007). Accordingly, it is necessary to conduct a study on the development of growth predictive model in liquid eggs for various *Salmonella* types. In particular, in Korea, the expiration dates of unpasteurized liquid egg are the same regardless of egg yolk, egg white, and liquid whole egg. Therefore, it is necessary to compare and analyze the changes in *Salmonella* depending on the storage time in various temperature conditions considering the manufacturing environments, distribution processes of liquid eggs.

In this study, we predicted the growth of *Salmonella* in unpasteurized liquid whole egg and egg yolk stored at 10–40℃, where *Salmonella* mainly proliferate, using a growth predictive model. The purpose of this study was to establish a proper storage temperature and the expiration dates of unpasteurized liquid eggs using the model predicting the growth of *Salmonella.*

Materials and Methods

Preparation of egg samples and bacterial cultures

The 10 liter of unpasteurized liquid whole eggs used in this experiment was purchased from an egg processing plant in Anseong, Gyeonggi-do in 2012. At the same time, 400 shell eggs were prepared and separated into egg yolk and egg white. The conditions of eggs used in this study were as follows: the microbial levels of unpasteurized liquid whole egg, egg yolk, and egg white used in this study were less than 500,000/g for general bacterial count, less than 100/g for *E. coli* count, *Salmonella* negative. The acidity (pH) was 6.39 ± 0.11 in egg yolk, 9.23 ± 0.02 in the egg white, and 7.49 ± 0.05 in liquid whole egg, which all satisfying the quality control standards of South Korea.

To artificially contaminate the unpasteurized liquid whole egg, egg yolk, and egg white described above, 5 kinds of *Salmonella* serovars — the two reference strains of *S.* Enteritidis ATCC 13076 and *S.* Gallinarum ATCC 9184, and *S*. Typhimurium monophasic, *S*. Bareilly, and *S*. Richmond which are the main issues in liquid eggs in South Korea (Kim et al., 2015) were used in the experiment.

Inoculation of a bacterial culture and measurement of bacterial counts

An aliquot (about 8 mL) of the liquid whole egg, egg yolk, and egg white was put into a 10 mL tube and inoculated with the mixed *Salmonella* serovars with the initial concentration of 3 Log CFU/g. A total of 1,000 mL inoculated liquid eggs were stored at 5, 10, 15, 20, 25, 30, 35 and 40℃ (n=15–20 samples per each temperature). Changes in the *Salmonella* growth in unpasteurized liquid egg were measured for up to 960 h. That is, each 10 mL sample was homogenized by adding 20 mL of sterilized 0.1% buffered peptone water (BPW, Oxoid, UK) to a test tube and using a vortexer for 30 s. One millitliter of the homogenized sample was diluted with 9 mL of sterilized 0.1% peptone water for each concentration level using the decimal dilution method and was uniformLy distributed to the selected medium, Xylose Lysine Deoxycholate Agar (Oxoid, UK). Each medium was incubated in a 37℃ incubator for 24 h, and the number of cells was counted to calculate Log CFU/mL.

Development of *Salmonella* **growth predictive model depending on the storage temperatures of unpasteurized liquid whole egg and egg yolk**

The development of a predictive model at each temperature was performed through first and second rounds. The first growth predictive model is based on the Baranyi function equation proposed by Baranyi and Robert (1994). The lag phase duration (LPD) and the maximum specific growth rate (μmax) of *Salmonella* depending on temperature and storage time were calculated with DMFit 3.5 Excel® Add-in (IFR, UK).

$$
y = y0 + \frac{\mu_{\text{max}}}{\ln(10)} - \frac{1}{\ln(10)} \ln \left(1 + \frac{e^{\mu_{\text{max}} A} - 1}{10^{(y_{\text{max}} - y_0)}} \right)
$$
 (1)

$$
A = t + \frac{1}{\mu_{\text{max}}} \text{In} \left[\frac{e^{-\mu_{\text{max}} t + q_0}}{1 + q_0} \right] \tag{2}
$$

$$
LPD = \frac{\ln\left(1 + \frac{1}{q_0}\right)}{\mu_{\text{max}}} \tag{3}
$$

Log*y*0 : Initial populations (Log CFU/mL)

y : Population at any time (Log CFU/mL)

 μ_{max} : Maximum specific growth rate (Log CFU/h)

*y*max : Maximum population (Log CFU/mL)

 q_0 : Measure of the initial physiological state of cell

LPD and μ_{max} , the variables of a primary model calculated in each temperature, were used to develop a secondary growth predictive model. The secondary model was developed on the basis of a Davey model (Davey, 1989) and a square-root model (Ratkowsky et al., 1982), using Graph Pad Prism V4.0 (Graph Pad Software, USA).

 $LPD = a + (b/T) + (c/T^2)$ *LPD* ∶ Lag phase duration (h)

a,*b*,*c* ∶ regression coefficient

T ∶ Temperature (°C)

 $\sqrt{\mu_{max}} = a (T - T_{min})$

 $Log\mu_{\text{max}}$: Maximum specific growth rate(Log CFU/h)

a ∶ regression coefficient

T ∶ Temperature (°C)

 T_{\min} : Theoretical minimum temperature for cell growth (°C)

Validation of growth predictive model

To verify the suitability of the growth predictive model, growth curves of *Salmonella* in liquid whole egg and yolk were fitted to the Baranyi model. Measured values of *Salmonella* in the unpasteurized liquid whole egg and egg yolk stored at 20℃ and 30℃, which were not used in the predictive model, were applied for validation. For assessment between the measured and predicted values, values of Bias factors (B_i) , accuracy factors (A_i) and root mean square error (RMSE) proposed by Baranyi et al. (1999) were compared. If B*f* is less than 0.7 or greater than 1.15, the model is not suitable. A*f* indicates the inaccuracy of the developed model as the A_f value moves away from 1 (Baranyi et al., 1999). If B_f and A_f value is close to 1, and RMSE was close to 0, the developed secondary model is considered to be fit for predicting the growth of *Salmonella*.

$$
B_f = 10^{\sum log(predicted/observed)/n}
$$
 (4)

$$
A_f = 10^{\sum log(predicted/observed)/n}
$$
 (5)

$$
RMSE = \sum_{n} \sqrt{\frac{(predicted - observed)^2}{n}}
$$
 (6)

Statistical analysis

Each experiment was repeated twice and the results were analyzed using SAS program version 9.3 (SAS Institute Inc., USA). A significant difference $(p<0.05)$ between LPD and μ_{max} was analyzed using a one-way ANOVA which utilizes the Duncan's multiple test method. In addition, *t*-test was used to analyze the significant difference in μmax and the LPD of liquid whole egg and egg yolk $(p<0.05)$.

Results and Discussion

Changes in the growth of *Salmonella* **spp. by storage temperature in various liquid eggs**

Salmonella did not proliferate in all liquid egg products stored at 5℃ even after artificially inoculating *Salmonella* (Fig. 1). These results coincided with the previous studies where cryopreservation minimizes the possibility of *Salmonella* proliferation (EFSA Panel on Biological Hazards, 2014) and the minimum growth temperature of *S*. Enteritidis is 6–8℃ (Whiting et al., 1997). In contrast, *Salmonella* at a storage temperature of above 10℃ proliferated in liquid whole egg and

Fig. 1. Growth curve of *Salmonella* **in non-pasteurized liquid whole egg and egg yolk product at different storage temperature and time.** ◆, whole egg product; ○, egg yolk product.

egg yolk over a time period. In liquid whole egg, the bacteria proliferated by more than 1 Log after 168 h (7 days) at 10℃ and 72 h (3 days) at 15℃. In addition, *Salmonella* proliferated more than twice of the initial concentration after 12, 9, and 8 hours at 25, 35, and 40℃, respectively. In egg yolk, *Salmonella* proliferated by more than 1 Log after 96 h (4 days) at 10℃ and after 33 h at 15℃, showing the proliferation about twice faster than in liquid whole egg. This study showed a similar result to Singh et al. (2011), where the mixure of four *Salmonella* serotypes including *S*. Enteritidis did not show proliferation in liquid whole egg stored below 7℃, but proliferated at 10–43℃. In the study of Gumudavelli et al. (2007), 5 types of *S*. Enteritidis proliferated in egg yolk at 10–43℃, but not at 7℃. Therefore, it is important to distribute the liquid whole egg and egg yolk below 10℃ in order to lower the risk of *Salmonella*.

In the case of the liquid egg white (Fig. 2), *Salmonella* at 5 and 10℃ was undetectable after 960 h (40 days). On the other hand, *Salmonella* at 25 and 30℃ survived maintaining the initial concentration (3 Log CFU/g). These results are similar to the results of Moon et al. (2016), where *S.* Enteritidis and *S.* Typhimurium proliferated slightly in eggs stored at 25℃ whereas they did not grow at 8, 10, and 15℃. Kang et al. (2011) reported that the growth of microorganisms in egg white was slower than that of liquid egg yolk and liquid whole egg.

Validation of growth predictive model for unpasteurized liquid eggs on the different storage temperature

The Baranyi model was well fitted to the growth of *Salmonella* in liquid whole egg and egg yolk showing the r² values of 0.98– 0.99. Table 1 shows the growth parameters (LPD and μmax) for *Salmonella*, obtained using the Baranyi model, in liquid whole egg and egg yolk. As the storage temperature increased, LPD significantly $(p<0.05)$ decreased and μ_{max} significantly $(p<0.05)$ increased in both liquid whole egg and egg yolk. At all temperatures, LPD was shorter and μmax was significantly higher (*p<*0.05), in egg yolk than in liquid whole egg, with the largest difference at 35℃. The difference in *Salmonella* spp. growth may be due to the albumen components such as lysozyme and ovotransferrin, which is present only in egg white. It is reported that the albumen can affect the degradation of bacterial cell walls, stunting the growth of *Salmonella* (Zhang et al., 2011).

The LPD and μ_{max} calculated in the primary study was used to derive a secondary growth model by applying to a Davey model (Davey, 1989) and a square-root model (Ratkowsky et al., 1982), respectively (Fig. 3). The r² values of the secondary models ranged from 0.92 to 0.99, indicating the suitability of the model applied. This is similar to trend of the previously

Fig. 2. Growth curve of *Salmonella* **in liquid egg white product at different storage temperature and time.**

Parameters		Temperature $(^{\circ}C)$				
		10	15	25	35	40
LPD	Whole egg	127.264 ± 0.113^a	44.465 ± 0.864^b	6.399 ± 0.037 °	3.904 ± 0.996 ^d	$4.456\pm0.006d$
	Egg yolk	56.822 ± 0.386 ^{a*}	$21.660\pm1.001^{b*}$	2.547 ± 0.141 ^{c*}	2.358 ± 0.068 ^c	2.286 ± 0.045 ^{c*}
μ_{max}	Whole egg	0.031 ± 0.000 ^e	0.116 ± 0.002 ^d	0.478 ± 0.004 ^c	$0.530 \pm 0.030^{\circ}$	0.616 ± 0.002 ^a
	Egg yolk	0.024 ± 0.001 ^{d*}	0.104 ± 0.002 ^{c*}	$0.587 \pm 0.010^{b*}$	0.973 ± 0.013 ^{a*}	0.972 ± 0.001 ^{a*}
	Whole egg	0.999 ± 0.000	0.994 ± 0.006	0.998 ± 0.002	0.965 ± 0.025	0.960 ± 0.001
	Egg yolk	0.995 ± 0.001	0.997 ± 0.000	0.994 ± 0.004	0.997 ± 0.002	0.994 ± 0.004

Table 1. Kinetics parameters for LPD and µmax at each temperature obtained after fitting the *Salmonella* **spp. growth data in liquid whole egg and egg yolk**

Values are means±STD (n=125).

Small letters mean within a row with different superscripts in are significantly different at the $p<0.05$ level.

 $*$ Values between sample are significantly different at the p <0.05 level.

LPD, lag phase duration (h); μ_{max} , maximum specific growth rate (Log CFU/mL/h); r^2 , coefficient of determination.

Fig. 3. Secondary modeling the LPD and µmax for liquid whole egg and egg yolk, derived from Davey model and square root model. LPD lag phase duration (h); μ_{max} , maximum specific growth rate (Log CFU/mL/h); T, temperature; ---, 95% confidence interval.

developed LPD model for *Salmonella* in egg yolk solution and mayonnaise (Elias et al., 2016; Moon et al., 2016). In addition, μ_{max} increased with increasing temperature and the difference between whole egg and egg yolk was larger. The previously developed μ_{max} model of mayonnaise (Elias et al., 2016) was closer to the model of egg yolk than the whole egg. Therefore, it is likely that *Salmonella* may grow rapidly, when egg yolk solution and processed foods containing egg yolk are stored at room temperature.

Further, the second growth predictive model was validated with experimental data at 20 and 30℃ (Fig. 4). When

Fig. 4. Validation of correspondence between liquid whole egg and egg yolk for *Salomonella* **spp.** BF, bias factor; AF, accuracy factor; RMSE, root mean square error; ●, values at 20℃; ○, values at 30℃.

comparing measured values with predictive values, B*f* values of whole egg and egg yolk were 0.968 and 0.993, respectively. A*f* values were also close to 1, indicating that the model is fit for predicting the growth of *Salmonella*. Extrapolating the egg yolk model using the observed value of whole egg indicated that there is a difference in the proliferation pattern of *Salmonella* (B*f*: 1.330). These data indicate that the models utilized in this study were suitable in predicting the growth of *Salmonella* spp. in the unpasteurized liquid whole egg and egg yolk under a variety of storage temperature and time conditions. Therefore, it is thought that the *Salmonella* growth predictive model can be used usefully in setting the storage temperature during the distribution process of unpasteurized liquid whole egg and egg yolk.

Conclusion

This study demonstrated there is a difference in the proliferation of *Salmonella* in unpasteurized liquid eggs depending on the types of liquid eggs (egg yolk, egg white, whole egg) and storage temperature, using *Salmonella* growth predictive model. Therefore, it is desirable to reset the expiration dates of liquid eggs taking into account of the product type, storage conditions (temperature and time), and microbiological factors.

Acknowledgements

This study is the result of the research project (B-FS03-2011-12-01) by the Animal and Plant Quarantine Agency.

References

Baranyi J, Roberts TA. 1994. A dynamic approach to predicting bacterial growth in food. Int J Food Microbiol 23:277-294. Baranyi J, Pin C, Ross T. 1999. Validating and comparing predictive models. Int J Food Microbiol 48:159-166. Choi SW, Ha JS, Kim BY, Lee DH, Park JK, Youn HN, Hong YH, Lee SB, Lee JB, Park SY, Choi IS, Song CS. 2014.

Prevalence and characterization of *Salmonella* species in entire steps of a single integrated broiler supply chain in Korea.

Poult Sci 93:1251-1257.

- Davey KR. 1989. A predictive model for combined temperature and water activity on microbial growth during the growth phase. J Appl Bacteriol 67:483-488.
- Elias SO, Alvarenga VO, Longhi DA, Sant'Ana AS, Tondo EC. 2016. Modeling growth kinetic parameters of *Salmonella* Enteritidis SE86 on homemade mayonnaise under isothermal and nonisothermal conditions. Foodborne Pathog Dis 13:462-467.
- EFSA Panel on Biological Hazards. 2014. Scientific opinion on the public health risks of table eggs due to deterioration and development of pathogens. EFSA J 12:3782.
- Gantois I, Ducatelle R, Pasmans F, Haesebrouck F, Gast R, Humphrey TJ, Van Immerseel F. 2009. Mechanisms of egg contamination by *Salmonella* Enteritidis. FEMS Microbiol Rev 33:718-738.
- Gumudavelli V, Subbiah J, Thippareddi H, Velugoti PR, Froning G. 2007. Dynamic predictive model for growth of *Salmonella* Enteritidis in egg yolk. J Food Sci 72:M254-M262.
- Kang GH, Cho SH, Seong PN, Park BY, Ham JS, Jeong SG, Kim DH, Chae HS. 2011. Microbial and physicochemical properties of liquid egg during cold storage. Korean J Food Sci An 31:557-562.
- Kim KO, Liu YN, Yoon LN, Park HJ. 2014. Comparison of quality characteristics and antioxidative activities of cookies containing blueberry powder and different types of egg yolk. J Korean Soc Food Sci Nutr 43:999-1008.
- Kim YJ, Chon JW, Lim JS, Song BR, Seo KH, Heo EJ, Park HJ, Wee SH, Oh DH, Moon JS. 2015. Traceback investigation for *Salmonella* contamination at egg processing plants in South Korea: Prevalence, antibiotic resistance, and epidemiological tracing by rep-PCR fingerprinting. J Food Sci 80:M759-M764.
- Malheiros PS, de Paula CMD, Tondo EC. 2007. Growth kinetics of *Salmonella* Enteritidis involved in outbreaks of foodborne illness in Rio Grande do Sul, Southern Brazil: A comparision with other serovas strains. Food Sci Technol 27:751-755.
- MFDS. 2015. The processing standards and ingredient specifications for livestock products, Ministry of Food and Drug safety, Korea. Available from: http://www.mfds.go.kr/eng/eng/index.do?nMenuCode=120&page=1&mode=view&board Seq=70016. Accessed at Nov 12, 2015.
- Moon HJ, Lim JG, Yoon KS. 2016. Comparative study of change in *Salmonella* Enteritidis and *Salmonella* Typhimurium populations in egg white and yolk. J Food Hyg Saf 31:342-348.
- Na YJ. 2014. Risk assessment and control of *Salmonella* Enteritidis on ready-to-eat (RTE) quail eggs products. M.S. thesis, Kyunghee Univ., Seoul, Korea.
- Ratkowsky DA, Olley J, McMeekin TA, Ball A. 1982. Relationship between temperature and growth rate of bacterial cultures. J Bacteriol 149:1-5.
- Sakha MZ, Fujikawa H. 2012. Growth characteristics of *Salmonella* Enteritidis in pasteurized and unpasteurized liquid egg products. Biocontrol Sci 17:183-190.
- Singh A, Korasapati NR, Juneja VK, Subbiah J, Froning G, Thippareddi H. 2011. Dynamic predictive model for the growth of *Salmonella* spp. in liquid whole egg. J Food Sci 76:M225-M232.
- Whiting RC, Buchanan RL. 1997. Development of a quantitative risk assessment model for *Salmonella* Enteritidis in pasteurized liquid eggs. Int J Food Microbiol 36:111-125.